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## Supporting Information

### Appendix S1

#### Expanded Materials and Methods

**Appendix S1.A. Local Sites.** We obtained data on the local species composition of 123 sites throughout the range of Hylidae (Tables S1–S3). Our major source of data was published studies of the amphibian faunas of local sites. We focused on well-studied site, typically of several km<sup>2</sup> in size, that represent a single biome or habitat (e.g. tropical lowland rainforest), but include multiple microhabitats (e.g. forest, stream edge, pond). We generally excluded sites spanning multiple biomes, and from poorly known regions in which the observed hylid richness was much lower than for other sites in the same region (possibly reflecting poor sampling or human impacts). In some cases, we also included species lists from national parks or reserves, particularly for regions where we could corroborate these lists with published range maps (e.g. U.S., Australia). For some areas having few obvious sites, we picked localities where large numbers of hylid species have been collected, based on literature or museum records. For areas with very low hylid diversity (e.g., Europe, Asia, Western North America), we used museum records.

For most analyses, we used a single, well-studied locality to represent each major biogeographic region (Table 1), in order to reduce potential problems of uneven numbers of sites among biogeographic regions, spatial autocorrelation, and inadequately surveyed sites. For a given region, we used

23 the site with the maximum local diversity of hylids within that region. Although  
24 this could lead to our analysis being influenced by outliers, we found that for  
25 each region, the site with the second highest diversity generally has diversity  
26 very similar to that of the site with the highest diversity (e.g. the maximum  
27 diversity for the Amazon region is 36 at Santa Cecilia, Ecuador, but Cocha  
28 Cashu, Manu National Park, Peru has 35; Table S1). For regions in which there  
29 are >1 sites with the same maximum diversity, we either used the more well  
30 studied site or one that was centrally located within the region. We then  
31 confirmed that these focal sites did not show spatial autocorrelation in species  
32 richness (Appendix S1.B). We also conducted some analyses using sites  
33 representing the mean richness for each region (see Appendix S1.L).

34 We considered 12 major biogeographic regions. Six correspond to major  
35 large-scale regions where hylids occur, exclusive of South America (North  
36 America [north of Mexico], Middle America [Mexico to Panama], the West  
37 Indies, Europe [including north Africa and the Middle East], Asia, Australia).  
38 We divided South America into six regions (Amazon, Andes, Atlantic rainforest,  
39 Cerrado, Chocó, and Guyana highlands) based on the standard domains used by  
40 Duellman (1999; see his Fig. 5:3) and previous authors, but with the following  
41 modifications: (i) regions without hylids were excluded, (ii) the Caribbean  
42 Coastal Forest and Llanos regions were included within the Amazonia-Guyana  
43 region (Amazonia hereafter), as they have relatively few endemic hylid species  
44 and have hylid faunas very similar to the Amazon region, (iii) the Guyanan

45 highlands were treated as separate from Amazonia, given their many endemic  
46 hylid species and genera, and (iv) the four contiguous grassland regions in  
47 southern South America were combined (Cerrado, Chaco, Monte, Pampas;  
48 referred to as Cerrado hereafter for brevity), given that only a limited number of  
49 regions can be used in likelihood analysis of ancestral areas (see Appendix S1.E).  
50 We also performed a limited set of analyses in which some of these regions were  
51 subdivided (Appendix S1.M), and confirmed that our main results were not  
52 overturned by doing so. The number of regions was somewhat limited due to  
53 the limitations of our method for biogeographic reconstruction (see below).  
54 Nevertheless, the regions used seem to correspond to major areas of endemism  
55 for hylids and other amphibian species (e.g. Duellman 1999).

56 We acknowledge that there is considerable debate about what constitutes  
57 a community (e.g. Ricklefs 2008; Brooker *et al.* 2009). Here we use “community”  
58 simply as the assemblage of species occurring at a given local site. Given the size  
59 and dispersal ability of hylids and the general scale of sites, all hylid species  
60 should have the potential to interact within these sites. For example, a single  
61 species may occur in more than one habitat (e.g., both pond and forest), and a  
62 single individual may occur in multiple habitats over the course of a single day.  
63 However, species within a site may partition microhabitats and resources in such  
64 a way that they may or may not actually interact or compete (e.g., sets of species  
65 utilizing ponds vs. streams as breeding sites). Our definition of most sites  
66 follows from previous studies of amphibians (e.g. Duellman 1978, 1988, 2001,

67 2005; Moen *et al.* 2009) and our goal is to understand variation in richness  
68 between sites at this general spatial scale.

69 We determined the latitude and longitude of each locality from the  
70 original literature sources (Table S2). Prior to analysis, we confirmed that the  
71 elevation for each site (and associated climatic data) was broadly consistent with  
72 values reported in the literature (when available) for that locality. For a few  
73 larger sites that are not as clearly defined (e.g., larger national parks), we selected  
74 a given point within that larger site for georeferencing, based on additional  
75 locality information and/or by choosing sites within the known elevational  
76 ranges of the relevant species. However, these localities were not used to  
77 represent any regions in the main analyses of the paper.

78 For some sites, the exact spatial extent was not clearly defined by the  
79 original authors. However, we found that for almost all regions, the maximum  
80 local richness is similar across multiple sites, regardless of their area. This  
81 strongly suggests that patterns of local richness among regions are not simply  
82 the result of differences in area of local sites.

83

84 **Appendix S1.B. Spatial Autocorrelation of Sites.** We conducted analyses using  
85 SAM (Spatial Analysis in Macroecology; Rangel *et al.* 2010) to test whether  
86 species richness values of the 12 focal sites are potentially similar due to  
87 geographic proximity alone. We used Moran's I test, including 1,000  
88 randomized replicates to generate a 95% confidence interval and then evaluated

89 whether observed values of  $I$  were more positive or negative than expected (and  
90 whether they fell outside the 95% confidence interval). We used a geographic  
91 coordinate matrix with six distance classes and with all options set to defaults for  
92 SAM. The results showed no significant spatial autocorrelation in species  
93 richness among sites ( $P > 0.312$  for all six classes, all values outside the 95%  
94 confidence interval). Results were generally similar for the potential correlates of  
95 species richness including climatic variables and summed clade ages (described  
96 below).

97

98 **Appendix S1.C. Selection of Climatic Variables.** We initially chose to use  
99 annual mean temperature (Bio1) and annual precipitation (Bio12) as two obvious  
100 and intuitive descriptors of climate. These two variables also showed significant  
101 relationships with local species richness, when all 123 sites are considered (see  
102 Results). Other variables, such as temperature seasonality (Bio4) and  
103 precipitation seasonality (Bio15) showed substantially weaker relationships with  
104 local richness than Bio1 and Bio12 (Bio4:  $r^2 = 0.194$ ;  $P < 0.0001$ ; Bio15:  $r^2 = 0.031$ ;  $P$   
105  $= 0.0525$ ).

106 We also conducted principal components analysis (PCA) of all 19 climatic  
107 variables simultaneously, using data from all 123 local sites. PCA was conducted  
108 with JMP version 7. We found that PC1 explained 47.3% of the climatic variation  
109 among sites, and was significantly correlated with local species richness (see  
110 Results). PCs 2–4 explained 20.6, 10.8, and 8.9% of the remaining variance, and

111 showed only weak relationships with local richness (PC2:  $r^2 = 0.004$ ;  $P = 0.475$ ;  
112 PC3:  $r^2 = 0.051$ ;  $P = 0.012$ ; PC4:  $r^2 = 0.006$ ;  $P = 0.408$ ). These PCs were not  
113 considered further. Eigenvectors for PC1 (Table S10) indicate that sites with high  
114 values for PC1 have relatively high annual mean temperatures (Bio1), warm  
115 temperatures in their coldest (Bio6, Bio11) and driest (Bio9) periods, limited  
116 temperature seasonality and annual range (Bio4, Bio7) and high isothermality  
117 (Bio3), and high annual precipitation (Bio12). Localities with the highest values  
118 for PC1 typically correspond to wet, lowland, tropical rainforest sites.

119

120 **Appendix S1.D. Divergence-Time Estimation.** We performed Bayesian  
121 divergence time estimation using BEAST version 1.4.7 (Drummond & Rambaut  
122 2007). In order to avoid repeating the time-intensive search for the best topology,  
123 we used the estimated maximum likelihood tree (Wiens *et al.* 2010a) as a  
124 constraint in the dating analyses. We used separate partitions within and  
125 between genes, and utilized the GTR + I +  $\Gamma$  model (generalized time reversible  
126 with parameters for invariant sites and gamma distribution of rates for variable  
127 sites) for each partition (based on Wiens *et al.* 2010a). Divergence times were  
128 estimated under the uncorrelated relaxed-clock tree model (Drummond *et al.*  
129 2006) with a Yule process speciation prior and Jeffrey's priors on the substitution  
130 model parameters. Lognormal date priors were placed on the root of the tree,  
131 with a broad prior distribution (lognormal standard deviation of 0.25) to allow for  
132 the possibility of substantial rate variance.

133 We used the following 10 fossil calibration points in estimating dates on  
134 the chronogram, including five in the ingroup and five among the outgroups. (i)  
135 Crown-group age of clade consisting of pipoids, pelobatoids, and neobatrachians  
136 is at least 144 Myo (Million years old), given the fossil taxon *Rhadinosteus parvus*  
137 (ostensibly a rhinophrynid, but clearly a pipoid) from the Late Jurassic  
138 (Tithonian; Rocek 2000). (ii) Crown-group age of *Caudiverbera* (=   
139 *Calyptocephalella*) and Myobatrachidae, at least 61.0 Myo, given fossil *Caudiverbera*  
140 (= *Calyptocephalella*) from the Early Paleocene (Baez 2000). (iii) Crown-group age  
141 of Bufonidae at least 23.8 Myo, given fossil *Bufo* from the Early Miocene (Rocek &  
142 Rage 2000), and that *Bufo* is nested inside of Bufonidae. (iv) Crown-group age of  
143 Ranidae + Microhylidae (*Rana catesbiana* and *Gastrophryne carolinensis*) at least  
144 33.7 Myo, given fossil *Rana* from the Late Eocene (Rocek & Rage 2000). (v) Stem-  
145 group age of Eleutherodactylidae, at least 35 Myo, given an amber-preserved  
146 specimen of *Eleutherodactylus* (sensu lato) from the La Toca formation in the  
147 Dominican Republic, with an estimated age from 35–40 Myo (Poinar &  
148 Cannatella 1987). (vi) Crown-group age of clade of Pelodyadinae +  
149 Phyllomedusinae within Hylidae, at least 28 Myo, given that the last terrestrial  
150 connection between Australia and South America was sundered at least 28 Myo  
151 (Sanmartin & Ronquist 2004). (vii) Crown-group age of *Acris-Pseudacris* clade, at  
152 least 15 Myo; Holman (2003) suggested that the extinct fossil taxon *Acris barbouri*  
153 is likely the sister group to extant *Acris* species and is at least 15 Myo (end of  
154 Miocene Hemingfordian North American Land Mammal Age; NALMA). Thus,

155 the split between *Acris* and *Pseudacris* is at least 15 Myo. Various *Pseudacris*  
156 fossils are known from the middle Miocene Barstovian of North America (~12–15  
157 Myo; Holman 2003), but given that these fossils cannot be assigned confidently  
158 to clades within *Pseudacris* we did not use this information (and given that the  
159 *Acris* fossils already show the *Acris-Pseudacris* clade to be at least 15 Myo). (viii)  
160 Crown-group age of Asian and European *Hyla* clade, at least 16 Myo; given fossil  
161 *Hyla* similar to extant *H. arborea* and *H. meridionalis* in the Lower Miocene of  
162 Austria (~16 Myo; Sanchiz 1998). We assume that these *Hyla* are closely related  
163 to *Hyla* presently extant in Europe. However, we cannot assume that these  
164 fossils are younger than the crown-group age of the extant European species.  
165 We assume instead that the crown group of the clade of European and Asian  
166 *Hyla* is at least 16 Myo based on these European fossils. (ix) Crown-group age of  
167 *H. squirella-H. gratiosa-H. cinerea* clade, at least 15 Myo; *H. goini* is a fossil species  
168 from Miocene Hemingfordian NALMA (15–19 Myo) thought to be closely  
169 related to, if not actually conspecific with, extant *H. squirella* (Holman 2003). *Hyla*  
170 *miofloridana* (Miocene, Hemingfordian NALMA; 15–19 Myo) is similar to *H.*  
171 *gratiosa* (Holman 2003). In our phylogeny, *H. cinerea*, *H. gratiosa*, and *H. squirella*  
172 form a clade. Thus, we assume that the crown group age of this clade is at least  
173 15 Myo. (x) Stem-group age of *H. avivoca-H. chrysocelis-H. versicolor* clade; *H.*  
174 *miocenica* is thought to be closely related to *H. chrysocelis* and *H. versicolor* and  
175 occurs in the early Miocene Barstovian (14–16 Myo; Holman 2003). In our



176 phylogeny *H. avivoca*, *H. chrysocelis*, and *H. versicolor* form a clade. We assume  
177 that the stem group age of these three species is at least 14 Myo.

178 We also considered three possible root ages for the tree, where the root  
179 consists of the crown group including Pipoidea + Pelobatoidea + Neobatrachia.  
180 Roelants et al. (2007) estimated this clade to be ~230 Myo, using MultiDivTime  
181 (Thorne & Kishino 2002). Wiens (2007) estimated ages of 155.3, 183.0, and 221.43  
182 Myo for the equivalent clade using penalized likelihood analysis with r8s  
183 (Sanderson 2003), depending on the root age of the tree. In the present analyses,  
184 three root ages for the tree were used: the youngest of these dates (155.3 Myo),  
185 the oldest (~230 Myo) and a date that is intermediate between these two (192.65  
186 Myo). However, given that these three root ages gave similar clade ages within  
187 Hylidae, all analyses were based on the intermediate age (192.65 Myo).

188

189 **Appendix S1.E. Biogeographic Reconstructions.** The timing and pattern of  
190 colonization of each region was estimated using LAGRANGE (Ree & Smith  
191 2008). Current versions of LAGRANGE allow fewer regions (~8) than the  
192 number of interest here (~12). We therefore performed two analyses. In the first  
193 analysis, South America was treated as a single area, and the six other regions  
194 were each treated as separate areas (results in Appendix S2.A). In the second  
195 (Appendix S2.B), South America was divided up into the six regions described  
196 above, and only two regions outside of South America were used (Australia;  
197 Northern Hemisphere regions [Middle America, West Indies, North America,

198 Europe, and Asia]). We combined the results of these two analyses to infer  
199 biogeographic patterns both in South America and in other areas. This  
200 combination was straightforward, as >90% of all species outside of South  
201 America are clearly the result of only two colonization events: the colonization of  
202 Australia by Pelodyadinae and the colonization of Middle America, North  
203 America, Europe, and Asia by Hylini (Wiens *et al.* 2006).

204       Species were assigned to regions based on range maps and descriptions of  
205 distributions in the database of the Global Amphibian Assessment  
206 ([www.globalamphibians.org](http://www.globalamphibians.org)). Several species had ranges extending into two or  
207 more areas, but such multi-state taxa are easily incorporated in LAGRANGE.  
208 The division between some areas was necessarily arbitrary. For example, species  
209 were considered as occurring in the Andes if their ranges extended to >1000 m in  
210 elevation (given our observation that many endemic Andean species occur at  
211 >1,000 m), but many species had ranges that extended both above and below  
212 1,000 m. Similarly, the distinction between the Atlantic forest and Cerrado  
213 regions in southern Brazil is not always clear. We assigned species to these two  
214 regions based partly on range maps, but also depending on descriptions of  
215 habitat (i.e. forest species assigned to Atlantic forest, grassland species to  
216 Cerrado).

217       For analyses using LAGRANGE, we created a matrix of geographically  
218 adjacent regions, such that direct dispersal between non-adjacent regions was not  
219 allowed. For example, transitions from Middle America to Europe or Asia must

220 pass through North America. For transitions to Australia, we assumed that these  
221 occurred through South America, but we considered direct dispersal from any  
222 region in South America to be theoretically possible, excluding the landlocked  
223 Andes mountains and Guyanan highlands. Although the Amazon rainforest and  
224 Atlantic forest are presently not adjacent (but only barely), we nevertheless  
225 allowed dispersal between these two regions of rainforest habitat. We did not  
226 place restrictions on the temporal pattern of dispersal between regions.

227         For many branches, LAGRANGE estimated more than one region or  
228 combination of regions as being possible (i.e. within two log-likelihood units of  
229 the optimal solution). For a given branch, we considered a given region to be  
230 unambiguously reconstructed if all the possible reconstructions included this  
231 region, either alone or in combination with other regions.

232

233 **Appendix S1.F. Biogeography and Local Richness.** We combined the  
234 biogeographic reconstructions with divergence-time estimates to determine to  
235 what extent the time that hylids have been present in each region determines  
236 their patterns of local diversity (i.e., the time-for-speciation effect; review in  
237 Stephens & Wiens 2003). We used two main approaches to quantify the timing  
238 of colonization of hylids in each region. Both have their own advantages and  
239 disadvantages, but both showed a significant relationship between the local  
240 richness of each region and time. Most localities within a region should share  
241 similar values for these temporal indices (especially given that our biogeographic

242 reconstruction methods are based on these same regions). Therefore, we only  
243 included the most diverse locality within each region for these analyses (but see  
244 Appendix S1.L)

245         First, we tested the relationship between the local richness of hylids in  
246 each region and how long hylids have been present in each region, based on the  
247 oldest reconstructed occurrence of hylids in each region (the oldest clade  
248 unambiguously reconstructed as present in that region). In some ways, this  
249 approach is the most conservative. However, this approach effectively assumes  
250 that only the first colonization of each region is relevant to explaining hylid  
251 species richness there, and the possible impact of multiple colonizations of the  
252 region is ignored.

253         Second, to better account for the impact of multiple colonizations of a  
254 given region by different clades at different points of time on richness, we tested  
255 the relationship between local richness and how long each clade has been present  
256 in each region, summed across clades. For this analysis, we determined the  
257 oldest occurrence of each clade in each region, and then summed these values  
258 across clades to estimate the overall time that hylids have been present in each  
259 region. One might be tempted to suggest that these colonization times should be  
260 averaged across clades rather than summed. However, doing so effectively  
261 eliminates the very information on colonizations by multiple clades that this  
262 approach was designed to incorporate. Furthermore, the number of clades alone  
263 is not the sole determinant of local richness, as their timing in each region is

264 important as well. We found that local richness is uncorrelated with the number  
265 of clades ( $r^2 = 0.232$ ;  $P = 0.113$ ) and mean clade age also shows a significant  
266 relationship with local richness ( $r^2 = 0.416$ ;  $P = 0.024$ ). There is a significant  
267 relationship between summed clade ages and number of clades in a region ( $r^2 =$   
268  $0.389$ ;  $P = 0.030$ ), but this significant relationship disappears when the two  
269 regions with single species (Asia, Europe) are eliminated ( $r^2 = 0.268$ ;  $P = 0.125$ ),  
270 and the strong relationship between summed clade ages and richness remains ( $r^2$   
271  $= 0.643$ ;  $P = 0.005$ ). Thus, consideration of summed clade ages offers valuable  
272 information that is potentially independent of the number of clades alone  
273 (although this approach is useful specifically because it incorporates information  
274 from multiple clades). We note that this approach does not, however,  
275 incorporate colonization events that are older than the origins of the major clades  
276 used.

277       As a potential third approach we estimated the total amount of time that  
278 hylids have been present in each region, based on the summed lengths of the  
279 terminal and internal branches reconstructed unambiguously as occurring in that  
280 region across all clades. However, this approach is potentially influenced by  
281 both the total number of species in each region and by the number of species  
282 sampled in each region. Therefore, we do not emphasize this approach here, but  
283 we note that it gives similar results to the other two approaches, especially the  
284 summed clade ages (first colonization:  $r^2 = 0.454$ ;  $P = 0.0162$ ; summed clade ages:  
285  $r^2 = 0.705$ ;  $P = 0.0006$ ; total time:  $r^2 = 0.648$ ;  $P = 0.0016$ )

286           The second approach required dividing hylids into major clades. We used  
287 the following clades: (i) subfamily Pelodryadinae, (ii) the subfamily  
288 Phyllomedusinae, (iii) the tribe Cophomantini, (iv) the *Dendropsophus* clade  
289 (*Dendropsophus* and *Xenohyla*), (v) the *Scinax* clade (*Scinax* and *Sphaenorhynchus*),  
290 (vi) the *Pseudis* clade (*Scarthyla*, *Pseudis*, *Lysapsus*, and *Podonectes*), (vii) the tribe  
291 Lophiohylini, and (viii) the tribe Hylini. Each of these clades is supported as  
292 monophyletic (Wiens *et al.* 2010a) and the species composition of each is  
293 relatively unambiguous. Together, these eight clades include all known hylid  
294 species. Admittedly, there are many ways that hylids could be divided into  
295 clades, but this set of clades is the most practical for several reasons. For  
296 example, the Pelodryadinae and Hylini occur primarily in Australia and Middle  
297 America, respectively, and are logical units.

298           One important limitation of these analyses is that they may be influenced  
299 by the ambiguity of the biogeographic reconstructions. For example, the Cerrado  
300 and Atlantic rainforest regions appear to have been colonized relatively recently,  
301 but this may reflect the ambiguity at many nodes because of the frequent  
302 dispersals between these two regions and between these regions and Amazonia.

303

304 **Appendix S1.G. Climate and Diversification Rates.** A relationship between  
305 climate and local richness may be explained by higher rates of diversification in  
306 the more species-rich climatic conditions. We estimated the diversification rate

307 for each genus given the total number of described species (Amphibiaweb;  
308 <http://amphibiaweb.org/>) and  
309 estimated age (from our chronogram). We used stem-group ages (which are  
310 similar to crown-group ages but can be accurately estimated with fewer species),  
311 and the method-of-moments estimator of diversification rates (Magallón &  
312 Sanderson 2001). This estimator includes a parameter ( $e$ ) for the (unknown) rate  
313 of extinction relative to the rate of speciation. We initially used a low, medium,  
314 and high value ( $e = 0, 0.45, 0.90$ ), but different values gave similar results, and  
315 only results using  $e = 0.45$  are presented. Preliminary analyses showed that  
316 diversification rates of genera are significantly related to their species richness ( $r^2$   
317  $= 0.275, P = 0.0001$ ) but not their age ( $r^2 = 0.049; P = 0.1285$ ), suggesting that  
318 comparisons of these rates are relevant for understanding richness patterns, and  
319 are not compromised by potentially faster rates in younger clades. Further, we  
320 are interested in the net accumulation of species from the time of origin of the  
321 clade to the present day (net diversification), and not instantaneous rates of  
322 speciation and extinction. Information on climatic data for genera is provided in  
323 the main text.

324

325 **Appendix S1.H. Climate, Time, and Local Richness.** We used our climatic  
326 reconstructions to test if the observed relationships between local richness and  
327 climate are ultimately related to greater time-for-speciation in environments that  
328 have been inhabited longer (as opposed to climate-diversity relationship being

329 caused by the influence of climate on diversification (speciation - extinction)).  
330 We focused on PC1, given that this variable shows the strongest relationships  
331 between climate and local richness (see Results). PC1 for local sites and PC1 for  
332 species are based on separate analyses, but they are based on the same climatic  
333 variables and yield very similar eigenvectors (compare Tables S10, S11)

334 To evaluate whether time drives the climate-diversity relationship, we  
335 first divided the observed range of PC1 into 14 bins of equal size (range), based  
336 on the range of observed PC1 values among local sites (-8–6). We then  
337 determined the mean richness of local sites that occur in each bin (Table S12).  
338 We found a strong relationship between mean local richness and PC1 using this  
339 approach ( $r^2 = 0.864$ ;  $P < 0.0001$ ), demonstrating that the climate-diversity  
340 relationship is not obfuscated by binning. We acknowledge that the number of  
341 bins is somewhat arbitrary.

342 We then used the reconstructions for PC1 across the tree to determine the  
343 age of the oldest clade occurring in a given climatic zone (bin), for each of the  
344 eight major clades. We used reconstructed values for internal nodes only and  
345 did not directly incorporate values in extant species. In a few cases, a single,  
346 relatively old branch spanned >1 bins, and the age of the this clade was used for  
347 both bins. Given that most clades span a broad range of values for PC1  
348 (indicating multiple colonizations of each climatic zones by different hylid  
349 clades), we summed our time estimates for each band across clades to obtain an  
350 index of the overall time that each band has been occupied by hylids. Thus,



351 climatic zones that have been colonized by multiple clades should have higher  
352 values than those colonized by a single clade, all other things being equal. We  
353 used linear regression to test for a relationship between the estimated time and  
354 mean richness for each bin.

355 Overall, this approach has some important limitations that should be  
356 noted. First, the analysis is based on species mean values, and the reconstructed  
357 values do not reflect the whole range of their ancestral or current climatic  
358 distributions. These differences between mean values and actual climatic  
359 distributions could be important in determining local species richness patterns  
360 (even if the reconstructions of ancestral mean values are completely accurate,  
361 which is also debatable). Second, the climate-diversity relationship is only  
362 moderately strong in hylids and may vary among regions (e.g., tropical  
363 rainforest sites in different regions may differ in richness by two or three-fold),  
364 and this regional variation is not directly incorporated. Third, some estimates of  
365 time for some bands may reflect common ancestry rather than independent  
366 colonization. Similarly, colonizations older than the age of these eight clades are  
367 not counted. Nevertheless, despite the various limitations of this approach, it  
368 may provide useful information beyond the reconstructed climatic distributions  
369 of the major clades. Further phylogenetic exploration of the causes of the  
370 climate-diversity relationship should be an important area for future research.

371

372 **Appendix S1.I. Diversification Rates and Local Richness in Allopatric and**  
373 **Sympatric Clades.** We tested the hypothesis that clades that do not co-occur  
374 with other hylid clades will have higher rates of diversification (all other things  
375 being equal, a higher local density of clades will lead to a higher local density of  
376 species). Two major hylid clades are either partially or entirely allopatric with  
377 respect to other hylid clades. Pelodryadines are the only hylids occurring in  
378 Australia and New Guinea. The tribe Hylini is the only clade of hylids occurring  
379 in North America, Europe, and Asia, and in many communities in Middle  
380 America. In many other communities in Middle America, particularly those in  
381 the tropical lowlands, hylines co-occur with some of species of other hylid  
382 clades. However, even in Middle America, hylines make up ~80% of hylid  
383 diversity, and appear to have colonized Middle America much earlier than other  
384 hylid clades (Moen *et al.* 2009). Therefore, we assigned Hylini to the allopatric  
385 category. The six other clades each occur with one or more other hylid clades in  
386 South America in most communities (Table S3). Given the estimated stem-group  
387 ages and species richness of these clades (Table 2), we then estimated the  
388 diversification rates for these eight clades as described for genera (Table 2), using  
389 the method-of-moments estimator (Magallón & Sanderson 2001). We then  
390 compared the allopatric and sympatric rates using a one-tailed non-parametric  
391 Mann-Whitney U test and using phylogenetic generalized least squares (Martins  
392 & Hansen 1997). This latter method allows testing of relationships between  
393 continuous and categorical variables in a phylogenetic context. We report the

394 results using stem-group ages with  $e = 0.45$ , but use of crown-group ages and  
395 other values of  $e$  gave generally similar results. These diversification rates were  
396 also strongly related to clade richness ( $r^2 = 0.794$ ;  $P = 0.003$ ) but not clade age ( $r^2 =$   
397  $0.112$ ;  $P = 0.4181$ ), suggesting that these rates are relevant to explaining species  
398 richness patterns among clades.

399 We used higher clades for these analyses (rather than genera) because one  
400 of the two allopatric clades (Pelodryadinae) effectively consists of a single genus  
401 and cannot readily be subdivided. Further, we expect to see the strongest effects  
402 at the level of these major allopatric clades, and not necessarily within them.

403 To address the robustness of our results to an alternate approach, we also  
404 conducted a set of analyses using the BiSSE method (Binary-state Speciation  
405 Extinction) of Maddison et al. (2007), as implemented in Mesquite version 2.52  
406 (Maddison & Maddison 2007). This method can test the effect of a trait on  
407 diversification rates using the entire tree. We used our time-calibrated  
408 phylogeny for 362 taxa to whether the likelihood estimate of net diversification  
409 rates was significantly improved by allowing for different rates in sympatric vs.  
410 allopatric clades (with sympatry vs. allopatry treated as a binary variable, as  
411 described above). We found no significant difference (ln likelihoods basically  
412 identical), confirming the results from PGLS analyses. However, this analysis  
413 may be compromised by incomplete sampling (<50%) of hyloid species in our  
414 phylogeny (Wiens *et al.* 2010a). We therefore performed another analysis in  
415 which described but unsampled species were added to their respective genera

416 (with their placement within genera being arbitrary), and branch lengths were  
417 then re-estimated in Mesquite (using the ultrametricize option and constraining  
418 ages of major clades and genera to those from the original chronogram). The  
419 final tree contained 902 hylid species (all described species plus several  
420 undescribed taxa included in our phylogeny). BiSSE analyses on this expanded  
421 tree confirmed that net diversification rates are higher in allopatric clades when  
422 these rates are estimated separately (sympatric rate = 0.04615; allopatric rate =  
423 0.0570), but that the overall ln likelihood is not significantly different from a  
424 model in which rates are set to be equal (3214.17 under both models).

425         We also tested whether sympatry between major clades influenced  
426 patterns of within-clade diversification over time. Previous authors (e.g.  
427 Phillimore & Price 2008) have suggested that diversification rates will decrease  
428 over time due to an increasing density of species within a clade, and have used  
429 the gamma statistic (Pybus & Harvey 2000) to test for slowing diversification  
430 (with strongly negative values of gamma indicating a significant slowdown  
431 relative to a model of constant diversification over time). In theory, sympatry  
432 between clades should also increase decrease diversification over time, as  
433 sympatry between clades could contribute significantly to increasing the density  
434 of species and decreasing opportunities for niche expansion and subsequent  
435 evolutionary radiation. Using our time-calibrated phylogeny, we quantified  
436 gamma in each of the eight major clades of hylids, and tested whether gamma  
437 was significantly negative. However, our phylogeny includes only a fraction of

438 the species for most clades (Table S9). Therefore, we also used simulations  
439 (based on the known number of described species in each clade and a model of  
440 diversification with constant rates and speciation only) to generate expected  
441 critical values for significantly negative gamma, given the known number of  
442 species in the clade and the number of species sampled. To correct for  
443 incomplete sampling in our comparisons of gamma between clades, we used the  
444 difference between the observed gamma and the critical value as our index (with  
445 negative values indicating gamma values more negative than expected; Table  
446 S9). We used the one-tailed Mann-Whitney test to evaluate whether this index  
447 was more strongly negative in the six sympatric clades relative to the two largely  
448 allopatric clades. All simulations and estimates of gamma were conducted in the  
449 R package LASER (Rabosky 2006). We acknowledge that this approach has some  
450 limitations. For example, limited taxon sampling can potentially lead to  
451 estimating significantly negative values of gamma for simulated data generated  
452 under the constant rates model, even when using the simulation approach  
453 applied here (Monte-Carlo constant-rates test; Cusimano & Renner 2010).  
454 However, we found no relationship between the proportion of species sampled  
455 from each clade and the clade's corrected index for gamma ( $r^2 = 0.050$ ;  $P =$   
456  $0.5945$ ).

457 We also tested whether the maximum local richness of clades was higher  
458 in clades that are primarily allopatric versus sympatric relative to other clades.  
459 We determined the maximum local diversity of each of the eight clades (Table 2)

460 from our survey of local communities (Table S3), and assigned Hylini and  
461 Pelodyadinae to the allopatric category. We used the one-tailed Mann-Whitney  
462 U test but not the phylogenetic test here, given that the maximum local diversity  
463 of a clade is not a trait that evolves on a tree.

464

465 **Appendix S1.J. Body-Size Variation.** We tested the hypothesis that greater  
466 phenotypic diversity among species might permit the co-existence of larger  
467 numbers of species in a local community. A previous morphometric analysis of  
468 hylids (Moen et al. 2009), including 14 variables for representatives of all hylid  
469 genera, showed that body-size (principal component 1; PC1) explains >90% of  
470 overall morphometric variation among hylids. Other principal components each  
471 accounted for less than 2.5% of the variation. Furthermore, previous dietary  
472 studies of hylids (e.g. Duellman 2005; Moen & Wiens 2009) suggest that hylids  
473 are generalist predators in which body size determines prey size and the specific  
474 prey that are eaten. Therefore, body size appears to be a critical variable in  
475 determining dietary resource use and resource overlap in hylids, and in  
476 describing overall phenotypic variation. We used the maximum snout-vent  
477 length (SVL) reported in the males of each species as a measure of body size.  
478 SVL and PC1 are strongly correlated among hylids ( $r = 0.991$ ; Moen *et al.* 2009),  
479 and SVL is frequently reported for hylid species (whereas PC1 is particular to a  
480 given study). Furthermore, given their conspicuous calling behavior, male  
481 hylids seem to be collected more frequently than females. Body sizes are

482 generally similar between males and females of a given hylid species, but not  
483 identical (e.g. Duellman 1978, 2001, 2005). Given that average adult body sizes  
484 for species are not always reported for hylids, we focused on maximum sizes.  
485 We obtained data on maximum male SVL (SVL hereafter) for almost every  
486 described species in the 123 communities that we surveyed (Table S3). Literature  
487 sources for body-size data are given after Table S7.

488         The maximum body size reported for a species might not match their  
489 body size in a given local community. However, species with small maximum  
490 SVL will be small in all communities. Further, geographic variation in SVL in  
491 hylids appears to be limited relative to overall variation in body size (Moen *et al.*  
492 2009).

493         We used linear regression to test the hypothesis that maximum SVL  
494 (among species) and the range of SVLs among species were greater in more  
495 diverse communities, and the minimum SVL is smaller in more diverse  
496 communities (Table S8). However, given stochastic sampling of species, we  
497 would expect, the minimum, maximum, and range of SVL to all be more extreme  
498 in more diverse communities. To partially account for this, we excluded  
499 communities with less than 3 species, leaving 89 species for this analysis.

500         In addition, we created a null distribution of test statistics for the  
501 regression that accounts for artifacts due to random sampling of body sizes, and  
502 we used this null distribution to assess the statistical significance of our results.  
503 To create this distribution, we first constructed random “communities” in which

504 species were randomly sampled with replacement from all hylids for which  
505 maximum male SVL was available (757 species; combining our lists of species  
506 from the phylogeny and from local communities); these null communities give  
507 an expected distribution of body sizes that would result from random sampling.  
508 Ten thousand null communities were constructed for all sizes of communities  
509 between 1 and 36 species (i.e. the observed range of local richness). Second, we  
510 constructed 10,000 random vectors of 89 communities (with the distribution of  
511 sizes corresponding to those in our actual data) by taking random samples of the  
512 previously simulated communities. Third, we conducted linear regressions of  
513  $\log(\text{species number})$  versus body size range/min./max. on these random vectors  
514 of communities to create a distribution of test statistics (in this case, the t-value).  
515 Finally, we compared the test statistic from our actual data to this null  
516 distribution to obtain a  $P$ -value. Again, the idea here was to ask what kind of  
517 regression results one would get from a sample of communities in which the  
518 body sizes are only a consequence of randomly sampling species of hylids for  
519 each community. In summary, these simulations account for the fact that  
520 random sampling by itself will lead to a larger range of body sizes in large  
521 communities. The results of these simulations confirm that the observed  
522 relationship between the range of body sizes in communities and their richness is  
523 not merely an artifact of sampling ( $P = 0.0152$ ).

524



525 **Appendix S1.K. Average Diversification Rates in Communities.** Local richness  
526 may depend upon the diversification rates of the clades that are present. For  
527 example, a given community may have high richness because it is dominated by  
528 one or more rapidly diversifying clades, especially if ecological conditions in that  
529 region or locality favor higher speciation rates. To test this hypothesis, we first  
530 estimated the diversification rate for each hylid genus (Table S6), as described  
531 above. We then came up with a weighted average diversification rate for each of  
532 the 12 focal localities with maximum richness in each region, by adding the  
533 diversification rates for each species of each genus, and dividing by the total  
534 number of species in each community. We then tested the hypothesis that  
535 average rates of diversification will be higher in more diverse communities using  
536 linear regression.

537 We acknowledge some limitations of this approach. First, the overall  
538 diversification rate for a wide-ranging genus may not reflect its diversification  
539 rate in the area of the local community. Second, our estimate does not address  
540 temporal variation in diversification rates within clades (although these are not  
541 necessarily of interest here). Third, we assume a similar relative extinction rate  
542 ( $e$ ) across the tree (a standard assumption). Despite these limitations, the results  
543 of these analyses are corroborated by those showing a lack of relationship  
544 between climate and diversification rates and larger-scale analyses showing that  
545 diversification rates are lower where multiple clades co-exist.

546

547 **Appendix S1.L. Analyses of Sites Representing Mean Richness.** For these  
548 analyses, we determined the mean local species richness of each the 12 regions  
549 based on our sampled sites. We then selected the site with the value closest to  
550 that mean in that region. For North America and Middle America, multiple sites  
551 had values equivalent to the mean richness; in these cases, we chose  
552 representative sites at low elevations that are centrally located within each  
553 region. Several regions had only a single site available (Chocoan, Guyana  
554 highlands) or all sampled sites already had equivalent richness (Asia, Europe,  
555 West Indies). The sites selected and their basic properties are shown in Table  
556 S13.

557 The results show that richness of these communities is not correlated with  
558 any of the environmental variables considered (temperature:  $r^2 = 0.082$ ;  $P = 0.366$ ;  
559 precipitation:  $r^2 = 0.207$ ;  $P = 0.132$ ; PC1:  $r^2 = 0.161$ ;  $P = 0.195$ ). Local richness is  
560 correlated with the summed clade ages for the region ( $r^2 = 0.439$ ;  $P = 0.019$ ), but  
561 not the timing of first colonization ( $r^2 = 0.199$ ;  $P = 0.146$ ). Like the results for  
562 maximum local richness, these results also show stronger relationships between  
563 summed clade ages and diversity than between climate and diversity. However,  
564 we strongly prefer those analyses using maximum richness, as there seems to be  
565 some obvious distortion using communities representing the mean richness. For  
566 example, using mean richness, southeastern Brazil has much higher local  
567 richness than any other region, including Amazonia. However, this is not  
568 necessarily because richness is generally higher there, but rather because we

569 were more cautious about including low-diversity sites in southeastern Brazil,  
570 given the possibility that sites there were not adequately sampled. In fact, when  
571 southeastern Brazil is removed from the analysis, the relationship between the  
572 timing of first colonization and mean local richness becomes significant ( $r^2 =$   
573  $0.457$ ;  $P = 0.022$ ). In addition, our definition of the Amazonian region includes  
574 some parts with relatively low richness (e.g. llanos grasslands). Thus, use of  
575 mean richness within a region may be more subject to the vagaries of regional  
576 delimitation and sampling.

577

578 **Appendix S1.M. Sensitivity to Regional Delimitation.** We also performed a  
579 series of analyses addressing the sensitivity of our results to how regions were  
580 delimited. In general the number of regions we used was determined by the  
581 limited number of regions that could be included in biogeographic analyses  
582 using LAGRANGE. Nevertheless, we also performed an analysis in which some  
583 regions were subdivided and the maximum local richness within each  
584 subdivision was used. A total of 19 regions were used (Table S14). We  
585 subdivided Amazonia into Western Amazonia (arbitrarily delimited as occurring  
586 in those countries bordering the Andes), Eastern Amazonia, and the llanos  
587 (tropical grasslands in Colombia and Venezuela). We subdivided the Cerrado  
588 region into Cerrado, Chaco, and Pampean Monte (following Duellman 1999).  
589 We subdivided Australia into Australia and New Guinea, Middle America into  
590 Mexico and Lower Central America and North America into Eastern North

591 America (east of the Rocky Mountains) and Western North America. Regions  
592 with single or very few localities were not subdivided (e.g., Guyana,  
593 Southeastern Brazil) nor were regions that were basically invariant in local  
594 richness (e.g., Asia, Europe, West Indies).

595         These analyses show significant relationships between local richness and  
596 climatic variables (temperature:  $r^2 = 0.267$ ;  $P = 0.024$ ; precipitation:  $r^2 = 0.208$ ;  $P =$   
597  $0.050$ ; PC1:  $r^2 = 0.235$ ;  $P = 0.035$ ), which are generally similar to those from  
598 analyzing the original 12 regions. There is also a significant relationship between  
599 local richness and time ( $r^2 = 0.221$ ;  $P = 0.042$ : for hylids overall;  $r^2 = 0.331$ ;  $P =$   
600  $0.010$ : for summed clade ages). However, these latter relationships are greatly  
601 weakened relative to the analysis of the original 12 regions. This weakening  
602 seems to occur for two reasons. First, the Llanos region is nested inside of  
603 Amazonia, and is treated as such for clade ages. However, this dry grassland  
604 region has low diversity relative to rainforest sites, and removing this data point  
605 dramatically strengthens this relationship ( $r^2 = 0.369$ ;  $P = 0.008$ : hylids overall;  $r^2$   
606  $= 0.575$ ;  $P = 0.003$ : summed clades). Second, the timing of colonization of  
607 different subregions may differ from that of the overall regions. Unfortunately,  
608 this is difficult to address, given that the number of regions that can be used in  
609 current likelihood-based biogeographic reconstructions is limited.

610

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703

704 Table S1. The 123 local sites considered in this study, including the  
 705 biogeographic region in which they occur, number of species, and the source  
 706 describing their hylid composition. Literature sources cited are listed  
 707 immediately following the table. The 12 regions are abbreviated as follows:  
 708 AMA = Amazon Basin; ANDES = Andes; ASIA = Asia; AUS = Australia; CERR =  
 709 Cerrado and adjacent grasslands; CHOCO = Chocoan region; EUR = Europe and  
 710 adjacent Palearctic areas; GUY = Guyanan highlands; MA = Middle America;  
 711 NA = North America; SEB = Atlantic rainforest of southeastern Brazil; WIN =  
 712 West Indies.

713

Location	Region	Species	Source
			Zimmerman and
Brazil: Manaus INPA	AMA	16	Rodrigues (1990)
Brazil: Para: Belem	AMA	23	Crump (1971)
Ecuador: Napo: Santa Cecilia	AMA	36	Duellman (1978)
			Toft and Duellman (1979);
Peru: Huanoco: Panguana	AMA	21	Aichinger (1987)
			Duellman and Mendelson
Peru: Loreto: Teniente Lopez	AMA	26	(1995)
Peru: Madre de Dios: Manu	AMA	35	Rodriguez and Cadle

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National Park (Cocha Cashu)			(1990)
Peru: Madre de Dios: Cuzco			
Amazonico Reserve	AMA	31	Duellman (2005)
			Hoogmoed and Gorzula
Venezuela: Bolivar: El Manteco	AMA	11	(1979)
Venezuela: Gaurico: Hato			
Masaguaral	AMA	5	Staton and Dixon (1977)
Venezuela: La Escalera Region	AMA	16	Duellman (1997)
Venezuela: Sierra de Lema	AMA	6	Duellman (1997)
	ANDE		
Argentina: La Rioja: Chilecito	S	1	Cei (1980)
Colombia: Boyaca: Parama de la	ANDE		
Rusia	S	1	Duellman (1988)
	ANDE		
Ecuador: Napo: Rio Salado	S	7	Duellman (1988)
Ecuador: Pichincha: Quebrada	ANDE		
Zapadores	S	1	Duellman (1988)
	ANDE		
Peru: Cuzco: Rio Cosnipata	S	3	Duellman (1988)
China: Fujian Prov.: Kuliang,			
near Fuzhou	ASIA	1	Museum data (MVZ)

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China: Guangdong Prov.: Tai- Yong, E Kwantun	ASIA	1	Museum data (MVZ)
China: Guangdong Prov.:Yim- Na-San	ASIA	1	Museum data (MVZ)
China: Guangdong: Dinghushan: Cha Chang	ASIA	1	Museum data (MCZ)
China: Jiangxi: Hong-San, SE Kiangsi Prov.	ASIA	1	Museum data (MVZ)
China: Yunnan Province, Nu Jiang Prefecture	ASIA	1	Museum data (CAS)
China: Yunnan, Baoshan Prefecture, Qushi	ASIA	1	Museum data (CAS)
Japan: Kanagawa: Odawara City	ASIA	1	Museum data (LACM)
Japan: Miyagi Pref.: Honshu: near Takayama	ASIA	1	Museum data (MVZ)
Korea: Kyonggi Prov.; Mt. Buckak, near Seou	ASIA	1	Museum data (MVZ)
Myanmar: Chin State: Falam Township, Lon Pi	ASIA	1	Museum data (CAS)
Australia: New South Wales:	AUS	2	Swan and Foster (2005)

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Mutawintji National Park

			<a href="http://www.nationalparks.nsw.gov.au/npws.nsf/content/yengo_nth Vertebrate_fauna">http://www.nationalparks.nsw.gov.au/npws.nsf/content/yengo_nth Vertebrate_fauna</a>
Australia: New South Wales:			
Northern Yengo National Park	AUS	6	
Australia: Northern Territory:			
Kakadu National Park	AUS	15	Finlayson et al. (2006) <a href="http://www.nt.gov.au/nr/eta/parks/manage/pdf/nps4_values.pdf">http://www.nt.gov.au/nr/eta/parks/manage/pdf/nps4_values.pdf</a>
Australia: Northern Territory:			
Nitmiluk National Park	AUS	17	
Australia: Queensland:			
Lamington National Park	AUS	14	Hero (2006) OzCam ( <a href="http://www.ozcam.gov.au/">http://www.ozcam.gov.au/</a> ); C. Hoskin (pers. comm.) OzCam ( <a href="http://www.ozcam.gov.au/">http://www.ozcam.gov.au/</a> ); C. Hoskin (pers. comm.)
Australia: Queensland:			
Townsville	AUS	12	
Australia: Queensland: Cairns	AUS	14	
Australia: Queensland: Kuranda	AUS	14	C. Hoskin (pers. comm.)

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Australia: Queensland: Dinden			
Forest Reserve	AUS	15	C. Hoskin (pers. comm.)
Australia: Queensland:			
Wooroonooran National Park	AUS	8	C. Hoskin (pers. comm.)
New Guinea: Lakekamu Basin,			
Ivimka Research Station	AUS	9	Allison et al. (1998)
New Guinea: Utai	AUS	6	Austin et al. (2008)
Argentina: Buenos Aires: Bahia			
Blanca	CER	1	Cei (1980)
Argentina: Cordoba: Capilla de			
Monte	CER	1	Cei (1980)
Argentina: Parque Nacional			
Chaco	CER	10	Cespedez et al. (2001)
Brazil: Goias: Espora Power			
Plant	CER	16	Vaz-Selva et al. (2007)
Brazil: Mato Grosso: Dardanelos			
Dam	CER	20	Sao Pedro et al. (2009)
Brazil: Rio Grande do Sul:			
Parque Nacional Aparados da			
Serra	CER	16	Deiques et al. (2007)
Brazil: Tocantins: Lajeado, Luis			
	CER	17	Brandao and Peres (2001)

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 Eduardo Magalhaes Dam

Uruguay: Montevideo: Pajas

Blancas CER 5 Nunez et al. (2004)

Uruguay: Salto: El Espinillar CER 9 Nunez et al. (2004)

Ecuador: Los Rios: Rio Palenque CHOC

Biological Station O 10 Duellman (1988)

France: Provence-Alpes-Cote-

d'Azur; 3 mi W EUR 1 Museum data (MVZ)

Georgia; ca. 10 km SSE

Borzhomi EUR 1 Museum data (MVZ)

Germany: Rostock,

Mecklenburg EUR 1 Museum data (MVZ)

Hungary: Budapest EUR 1 Museum data (MVZ)

Morocco: Settat; Ben-Slimane, 5

km N of EUR 1 Museum data (USNM)

Spain: Andalusia: Cadiz Prov.:

Benalup de Sidonia EUR 1 Museum data (MVZ)

Ukraine: Kiev EUR 1 Museum data (CM)

Venezuela: Gran Sabana GUY 8 Duellman (1997)

Costa Rica: Cartago: Moravia MA 11 Duellman (2001)

Costa Rica: Heredia: La Selva MA 12 Scott et al. (1983)

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Costa Rica: Heredia: Volcan			
Barba	MA	5	Duellman (2001)
Costa Rica: Las Canas: Finca			
Taboga	MA	6	Scott et al. (1983)
Costa Rica: Puntarenas: Las			
Cruces	MA	10	Scott et al. (1983)
Costa Rica: Puntarenas: Rincon			
de Osa	MA	11	Scott et al. (1983)
Guatemala: El Peten: Tikal	MA	8	Lee (1996)
			McCranie and Wilson
Honduras: Atlantida: La Ceiba	MA	6	(2002)
Honduras: Atlantida: Quebrada			McCranie and Wilson
de Oro	MA	4	(2002)
Honduras: Copan: Laguna de			McCranie and Wilson
Cerro	MA	8	(2002)
Honduras: Copan: Quebrada			McCranie and Wilson
Grande	MA	8	(2002)
Honduras: Gracias de Dios:			McCranie and Wilson
Barra Patuka	MA	7	(2002)
Mexico: Chiapas: Rayon			
Mescalapan	MA	6	Duellman (2001)

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Mexico: Distrito Federal: Lago			
Xochimilco	MA	1	Duellman (2001)
Mexico: Durango: El Salto	MA	1	Duellman (2001)
Mexico: Guerrero: Puerto del			
Gallo	MA	8	Duellman (2001)
Mexico: Hidalgo: El Chico	MA	3	Duellman (2001)
Mexico: Jalisco: Chamela	MA	8	Ramirez-Bautista (1994)
Mexico: Michoachan: Nueva			
Italia	MA	5	Duellman (2001)
Mexico: Oaxaca: Puerto			
Escondido	MA	6	Duellman (2001)
Mexico: Oaxaca: San Gabriel			
Mixtepec	MA	8	Duellman (2001)
Mexico: Oaxaca: Tehuantepec	MA	4	Duellman (2001)
Mexico: Oaxaca: Tuxtepec	MA	8	Duellman (2001)
Mexico: Oaxaca: Vista Hermosa	MA	9	Duellman (2001)
Mexico: Puebla: 14.4 km W			
Huachinango	MA	4	Duellman (2001)
Mexico: Sinaloa: Mazatlan	MA	5	Duellman (2001)
Mexico: Sonora: Alamos	MA	3	Duellman (2001)
Mexico: Veracruz: Acultzingo (1	MA	4	Duellman (2001)

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km W)			
Mexico: Veracruz: Cuatlapan	MA	9	Duellman (2001)
Mexico: Veracruz: Estacion Los			
Tuxtlas	MA	8	Vogt et al. (1997)
Mexico: Veracruz: Huatusco (3k			
SW)	MA	7	Duellman (2001)
Mexico: Veracruz: Mata de			
Oscuara	MA	4	Duellman (2001)
Mexico: Veracruz: Volcan San			
Martin	MA	6	Duellman (2001)
Mexico: Yucatan: Piste	MA	7	Lee (1996)
Panama: Barro Colorado Island	MA	10	Rand and Myers (1990)
Panama: Cocle: El Valle	MA	9	Duellman (2001)
Panama: Colon: Achiote	MA	7	Duellman (2001)
Panama: Darien:Rio Tuira at Rio			
Mono	MA	7	Duellman (2001)
USA: California: Contra Costa			
Co.; 1 mi NW Alamo	NA	1	Museum data (MVZ)
USA: California: San Diego Co.:			
Spring, Pine Mountain	NA	2	Museum data (MVZ)
USA: Colorado: Commanche	NA	1	<a href="http://www.npwrc.usgs">http://www.npwrc.usgs</a> .

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National Grassland			<a href="http://www.nps.gov/resource/herps/comaherp/index.htm">gov/resource/herps/comaherp/index.htm</a>
USA: Florida: Everglades			<a href="http://www.nps.gov/arc">http://www.nps.gov/arc</a>
National Park	NA	4	<a href="http://www.nps.gov/arc/hive/ever/eco/herps.htm">hive/ever/eco/herps.htm</a>
USA: Florida: Timucuan			
Ecological and Historic Preserve	NA	7	Tuberville et al. (2005) <a href="http://fl.biology.usgs.gov/armi/Okefenokee/Okefenokee_Tables/okefenokee_tables.html">http://fl.biology.usgs.gov/armi/Okefenokee/Okefenokee_Tables/okefenokee_tables.html</a>
USA: Georgia: Okefenokee			
Swamp	NA	10	
USA: Georgia: Okmulgee			
National Monument	NA	9	Tuberville et al. (2005)
USA: North Carolina: Moores			
Creek National Battlefield	NA	10	Tuberville et al. (2005) <a href="http://www.nps.gov/arc">http://www.nps.gov/arc</a>
USA: North Dakota: Badlands			
National Park	NA	1	<a href="http://www.nps.gov/cuva/naturescience/amphibians.htm">hive/badl/exp/rept-amph.htm</a>
USA: Ohio: Cuyahoga Valley			
National Park	NA	3	<a href="http://www.nps.gov/cuva/naturescience/amphibians.htm">ans.htm</a>
USA: South Carolina: Congaree	NA	12	Tuberville et al. (2005)

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Swamp National Monument			
USA: South Carolina: Savanna			Gibbons and Semlitsch
River Ecology Laboratory	NA	12	(1991)
			<a href="http://www.npwrc.usgs.gov/resource/wildlife/sa">http://www.npwrc.usgs.</a>
USA: South Dakota: Sand Lake			<a href="http://www.npwrc.usgs.gov/resource/wildlife/sa">gov/resource/wildlife/sa</a>
National Wildlife Refuge	NA	3	<a href="http://www.npwrc.usgs.gov/resource/wildlife/sa">ndlake/herp.htm</a>
			<a href="http://www.dlia.org/atbi">http://www.dlia.org/atbi</a>
USA: Tennessee: Great Smoky			<a href="http://www.dlia.org/atbi">/species/Animalia/Chord</a>
Mountains National Park	NA	4	<a href="http://www.dlia.org/atbi">ata/Amphibia/Anura/Hy</a>
			<a href="http://www.dlia.org/atbi">lidae/index.shtml</a>
USA: Texas: Padre Island			<a href="http://www.nps.gov/pais/naturescience/amphibia">http://www.nps.gov/pai</a>
National Seashore	NA	2	<a href="http://www.nps.gov/pais/naturescience/amphibia">ns.htm</a>
			<a href="http://www.nps.gov/pais/naturescience/amphibia">http://www.nps.gov/zio</a>
USA: Utah: Zion National Park	NA	1	<a href="http://www.nps.gov/pais/naturescience/amphibia">n/naturescience/reptiles.</a>
			<a href="http://www.nps.gov/pais/naturescience/amphibia">htm</a>
			<a href="http://www.nps.gov/pais/naturescience/amphibia">http://www.nps.gov/she</a>
USA: Virginia: Shenandoah			<a href="http://www.nps.gov/pais/naturescience/amphibia">n/naturescience/upload/</a>
National Park	NA	4	<a href="http://www.nps.gov/pais/naturescience/amphibia">Amphibians_of_Shenando</a>
			<a href="http://www.nps.gov/pais/naturescience/amphibia">ah_National_Park.pdf</a>
USA: Washington: Pierce Co.	NA	1	Museum data (MVZ)

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Parkland, Wake Lake

			<a href="http://www.nps.gov/yell/naturescience/reptiles.htm">http://www.nps.gov/yell/naturescience/reptiles.htm</a>
USA: Wyoming: Yellowstone National Park	NA	1	
			Silvano and Pimenta (2003)
Brazil: Bahia: Estacion VeraCruz	SEB	28	
Brazil: Bahia: Zumbidos			Silvano and Pimenta (2003)
Palmares	SEB	20	
Brazil: Boriacea	SEB	26	Heyer et al. (1990)
Dominican Republic: Samana: Laguna	WIN	3	Museum data (USNM)
Dominican Republic: Samana: Rio San Juan	WIN	3	Museum data (USNM)
Haiti: Ouest: Furcy	WIN	3	Museum data (USNM)
Jamaica, Manchester Parish, Mandeville	WIN	3	Museum data (USNM)
Jamaica, Quick Step	WIN	3	Museum data (USNM)
Jamaica: Trelany Parish: Windsor	WIN	3	Museum data (USNM)

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- 821

822 Table S2. The 123 local communities considered in this study, including the  
 823 latitude, longitude, elevation (m above sea level), annual mean temperature ( $^{\circ}$  C),  
 824 and annual precipitation (mm).

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Location	Latitude	Longitude	Elevation	Annual mean temp.	Annual precipitation
Brazil: Manaus, INPA	-3.217	-60.033	27	27.4	2381
Brazil: Para: Belem	-1.350	-48.500	4	26.9	2728
Ecuador: Napo: Santa Cecilia	0.050	-76.983	327	25.4	3670
Peru: Huanoco: Panguana	-9.617	-74.933	269	25.7	2179
Peru: Loreto: Teniente Lopez	-2.594	-76.116	211	25.1	2978
Peru: Madre de Dios: Manu National Park (Cocha Cashu)	-11.900	-71.367	343	25.1	3031
Peru: Madre de Dios: Cuzco Amazonico Reserve	-12.550	-69.050	175	25.6	2479
Venezuela: Bolivar: El Manteco	7.417	-62.350	238	26.0	1263
Venezuela: Gaurico: Hato Masaguaral	8.550	-67.583	67	27.5	1336
Venezuela: La Escalera Region	6.617	-61.550	151	26.5	1730
Venezuela: Sierra de Lema	6.000	-61.383	776	22.9	2155

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Argentina: La Rioja: Chilecito	-29.167	-67.500	1125	18.5	280
Colombia: Boyaca: Parama de la					
Rusia	5.900	-73.200	2466	15.3	1556
Ecuador: Napo: Rio Salado	-0.196	-77.729	1425	19.8	2742
Ecuador: Pichincha: Quebrada					
Zapadores	-0.288	-78.779	2034	16.7	2276
Peru: Cuzco: Rio Cosnipata	-13.104	-71.321	1694	18.4	1814
China: Fujian Prov.: Kuliang,					
near Fuzhou	26.100	119.383	458	16.8	1457
China: Guangdong Prov.: Tai-					
Yong, E Kwantun	23.622	115.993	666	17.6	1718
China: Guangdong Prov.:Yim-					
Na-San	24.417	116.400	596	18.3	1659
China: Guangdong:					
Dinghushan: Cha Chang	23.167	112.533	374	19.8	1674
China: Jiangxi: Hong-San, SE					
Kiangsi Prov.	28.100	117.433	528	15.5	1911
China: Yunnan Province, Nu					
Jiang Prefecture	26.812	98.886	1292	18.1	1493
China: Yunnan, Baoshan					
Prefecture, Qushi	25.2833	98.600	1466	16.6	1449

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Japan: Kanagawa: Odawara					
City	35.254	139.148	68	14.5	1814
Japan: Miyagi Pref.: Honshu:					
Takayama, near	38.297	141.073	17	11.1	1274
Korea: Kyonggi Prov.; Mt.					
Buckak, near Seou	37.5667	127.000	33	10.4	1332
Myanmar: Chin State: Falam					
Township, Lon Pi	22.877	93.628	1866	15.5	2078
Australia: New South Wales:					
Mutawintji National Park	-31.283	142.250	193	19.3	283
Australia: New South Wales:					
Northern Yengo National Park	-33.033	150.783	179	17.2	1016
Australia: Northern Territory:					
Kakadu National Park	-12.683	132.483	12	28.0	1767
Australia: Northern Territory:					
Nitmiluk National Park	-14.109	132.255	266	26.7	1417
Australia: Queensland:					
Lamington National Park	-28.146	153.113	648	16.6	1749
Australia:					
Queensland:Townsville	-19.267	146.817	5	24.4	1333
Australia: Queensland: Cairns	-16.917	145.767	8	24.9	2868

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Australia: Queensland: Kuranda	-16.817	145.650	379	22.6	2731
Australia: Queensland: Dinden Forest Reserve	-16.960	145.649	518	22.0	2739
Australia: Queensland: Wooroonooran National Park	-17.400	145.820	1471	16.0	4431
New Guinea: Lakekamu Basin, Ivimka Research Station	-7.735	146.496	124	26.7	1835
New Guinea: Utai	-3.396	141.583	229	25.6	2636
Argentina: Buenos Aires: Bahia Blanca	-38.717	-62.283	20	16.2	642
Argentina: Cordoba: Capilla de Monte	-30.850	-64.517	976	14.9	729
Argentina: Parque Nacional Chaco	-27.967	-59.583	57	21.5	1336
Brazil: Goias: Espora Power Plant	-18.674	-51.881	574	24.2	1913
Brazil: Mato Grosso: Dardanelos Dam	-10.000	-60.000	201	24.4	2256
Brazil: Rio Grande do Sul: PN Aparados da Serra	-29.183	-50.083	961	14.7	1888
Brazil: Tocantins: Lajeado, Luis	-21.017	-46.933	934	19.6	1740

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Eduardo Magalhaes Dam					
Uruguay: Montevideo: Pajas					
Blancas	-34.867	-56.367	0	17.0	988
Uruguay: Salto: El Espinillar	-30.933	-57.867	35	19.6	1434
Ecuador: Los Rios: Rio Palenque					
Biological Station	-0.180	-79.183	485	23.8	3463
France: Provence-Alpes-Cote-					
d'Azur; 3 mi W	43.550	6.957	1	14.3	900
Georgia; ca. 10 km SSE					
Borzhomi	43.883	40.083	1249	6.5	1609
Germany: Rostock,					
Mecklenburg	54.083	12.133	14	7.7	623
Hungary: Budapest	47.500	19.044	106	10.1	570
Morocco: Settat; Ben-Slimane, 5					
km N of	33.600	-7.100	289	17.1	499
Spain: Andalusia: Cadiz Prov.:					
Benalup de Sidonia	36.344	-5.810	108	16.6	854
Ukraine: Kiev	50.433	30.517	121	6.9	666
Venezuela: Gran Sabana	5.883	-61.383	1543	18.4	2162
Costa Rica: Cartago: Moravia	9.8625	-83.438	1007	20.6	3562
Costa Rica: Heredia: La Selva	10.417	-83.950	54	26.1	4147

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Costa Rica: Heredia: Volcan					
Barba	10.129	-84.071	2110	14.8	2883
Costa Rica: Las Canas: Finca					
Taboga	10.333	-85.200	13	26.7	1938
Costa Rica: Puntarenas: Las					
Cruces	8.800	-83.000	1349	19.0	3988
Costa Rica: Puntarenas: Rincon					
de Osa	8.700	-83.483	38	26.2	4504
Guatemala: El Peten: Tikal	17.225	-89.613	254	24.6	1324
Honduras: Atlantida: La Ceiba	15.783	-86.783	10	26.5	2407
Honduras: Atlantida: Quebrada					
de Oro	15.633	-86.792	1132	20.1	2078
Honduras: Copan: Laguna de					
Cerro	15.079	-88.955	672	22.9	1637
Honduras: Copan: Quebrada					
Grande	15.083	-88.917	1324	19.2	1655
Honduras: Gracias de Dios:					
Barra Patuka	15.800	-84.300	2	26.9	2956
Mexico: Chiapas: Rayon					
Mescalapan	17.146	-93.012	1768	17.0	2283
Mexico: Distrito Federal.: Lago	19.300	-99.117	2240	15.8	809

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Xochimilco					
Mexico: Durango: El Salto	23.800	-105.400	2603	11.5	1086
Mexico: Guerrero: Puerto del Gallo	17.471	-100.212	1761	19.7	1605
Mexico: Hidalgo: El Chico	20.367	-98.733	2007	15.1	654
Mexico: Jalisco: Chamela	19.533	-105.083	1	26.3	774
Mexico: Michoachan: Nueva Italia	19.020	-102.100	412	27.4	776
Mexico: Oaxaca: Puerto Escondido	15.850	-97.070	2	27.7	938
Mexico: Oaxaca: San Gabriel Mixtepec	16.204	-97.129	1768	18.1	1328
Mexico: Oaxaca: Tehuantepec	16.333	-95.233	53	27.0	781
Mexico: Oaxaca: Tuxtepec	18.090	-96.120	30	25.3	2637
Mexico: Oaxaca: Vista Hermosa	17.888	-96.388	1416	17.7	3615
Mexico: Puebla: 14.4 km W Huachinango	20.183	-98.250	2253	13.3	1259
Mexico: Sinaloa: Mazatlan	23.217	-106.417	9	24.8	717
Mexico: Sonora: Alamos	29.217	-110.133	597	22.2	504
Mexico: Veracruz: Acultzingo (1 km W)	18.700	-97.317	2093	15.7	784

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Mexico: Veracruz: Cuatlapan	18.870	-97.030	1041	20.4	1757
Mexico: Veracruz: Estacion Los Tuxtlas	18.583	-95.100	350	24.2	2947
Mexico: Veracruz: Huatusco (3k SW)	19.141	-96.991	1369	18.4	1816
Mexico: Veracruz: Mata de Oscuara	19.208	-96.808	767	21.4	1623
Mexico: Veracruz: Volcan San Martin	18.572	-95.169	1015	20.0	2952
Mexico: Yucatan: Piste	20.700	-88.467	30	25.4	1234
Panama: BCI	9.167	-79.833	31	26.7	2757
Panama: Cocle: El Valle	8.600	-80.133	643	23.9	2076
Panama: Colon: Achiote	9.223	-80.019	27	26.7	3281
Panama: Darien: Rio Tuirra at Rio Mono	7.704	-77.546	142	25.8	2944
USA: California: Contra Costa Co.; 1 mi NW Alamo	37.859	-122.046	125	14.3	551
USA: California: San Diego Co.: Spring, Pine Mountain	33.340	-116.650	1136	13.8	508
USA: Colorado: Commanche National Grassland	37.4600	-102.620	1337	10.6	382

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USA: Florida: Everglades					
National Park	25.391	-80.681	7	23.4	1431
USA: Florida: Timucuan					
Ecological and Historic Preserve	30.350	-81.502	19	19.7	1314
USA: Georgia: Okefenokee					
Swamp	30.667	-82.333	37	19.0	1442
USA: Georgia: Okmulgee					
National Monument	32.840	-83.540	122	16.9	1267
USA: North Carolina: Moores					
Creek National Battlefield	34.458	-78.112	4	16.1	1432
USA: North Dakota: Badlands					
National Park	43.744	-101.941	740	7.8	415
USA: Ohio: Cuyahoga Valley					
National Park	41.240	-81.550	243	8.8	1014
USA: South Carolina: Congaree					
Swamp National Monument	33.930	-80.840	73	16.3	1268
USA: South Carolina: Savanna					
River Ecology Laboratory	33.344	-81.735	115	16.3	1290
USA: South Dakota: Sand Lake					
National Wildlife Refuge	45.813	-98.219	391	4.3	500
USA: Tennessee: Great Smoky					
	35.710	-83.510	420	12.5	1384

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Mountains National Park

## USA: Texas: Padre Island

National Seashore	26.950	-97.383	1	21.7	728
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USA: Utah: Zion National Park	37.459	-113.224	1625	10.3	386
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## USA: Virginia: Shenandoah

National Park	38.500	-78.450	914	6.8	1153
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## USA: Washington: Pierce Co.

Parkland, Wake Lake	47.156	-122.433	115	9.8	1273
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## USA: Wyoming: Yellowstone

National Park	44.565	-110.400	2438	-1.2	603
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Brazil: Bahia: Estacion VeraCruz	-16.383	-39.167	98	24.0	1540
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## Brazil: Bahia: Zumbidos

Palmares	-14.033	-39.150	133	24.1	2218
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Brazil: Boriacea	-23.638	-45.838	907	17.2	2468
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## Dominican Republic: Samana:

Laguna	19.250	-69.417	280	24.3	2099
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## Dominican Republic: Samana:

Rio San Juan	19.233	-69.317	128	25.2	2068
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Haiti: Ouest: Furcy	18.411	-72.288	1378	18.3	1417
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## Jamaica, Manchester Parish,

Mandeville	18.033	-77.500	639	22.0	1590
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Jamaica, Quick Step	18.250	-77.717	446	23.0	1742
Jamaica: Trelany Parish:					
Windsor	18.350	-77.650	192	24.1	1631

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828 Table S3. Species composition, clade composition, and the maximum reported  
 829 male body size (mm) for species in the 123 local communities considered in this  
 830 study. Literature sources for data on species composition of each community are  
 831 provided in Table S1, whereas sources for data on body sizes are provided in  
 832 Table S7.  
 833

Locality	Species	Clade	SVL
Brazil: Manaus, INPA	<i>Hypsiboas boans</i>	Cophomantini	132.0
	<i>Hypsiboas geographica</i>	Cophomantini	62.0
	<i>Hypsiboas granosa</i>	Cophomantini	44.0
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>minutus</i>	clade	23.0
	<i>Dendropsophus sp.</i>	<i>Dendropsophus</i>	
	( <i>microcephalus</i> group)	clade	
	<i>Dendropsophus sp.</i>	<i>Dendropsophus</i>	
	( <i>parviceps</i> group)	clade	
	<i>Osteocephalus</i>		
	<i>buckleyi</i>	Lophiohylini	64.1
	<i>Osteocephalus</i>		
<i>taurinus</i>	Lophiohylini	85.0	
<i>Osteocephalus sp.</i>	Lophiohylini		

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	<i>Trachycephalus</i>		
	<i>coriaceus</i>	Lophiohylini	63.0
	<i>Trachycephalus</i>		
	<i>resinifictrix</i>	Lophiohylini	100.5
	<i>Phyllomedusa bicolor</i>	Phyllomedusinae	115.0
	<i>Phyllomedusa tarsius</i>	Phyllomedusinae	97.0
	<i>Phyllomedusa</i>		
	<i>tomopterna</i>	Phyllomedusinae	48.0
	<i>Phyllomedusa</i>		
	<i>vaillanti</i>	Phyllomedusinae	59.8
	<i>Scinax cruentommus</i>	<i>Scinax</i> clade	28.0
Brazil: Para: Belem	<i>Hypsiboas calcaratus</i>	Cophomantini	47.5
	<i>Hypsiboas</i>		
	<i>geographicus</i>	Cophomantini	62.0
	<i>Hypsiboas granosa</i>	Cophomantini	44.0
	<i>Hypsiboas</i>		
	<i>multifasciatus</i>	Cophomantini	57.3
	<i>Hypsiboas raniceps</i>	Cophomantini	71.0
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>brevifrons</i>	clade	22.0
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	44.0

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<i>leucophyllatus</i>	clade	
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>melanargyreus</i>	clade	34.0
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>minutus</i>	clade	23.0
	<i>Dendropsophus</i>	
<i>Dendropsophus nanus</i>	clade	22.0
<i>Osteocephalus</i>		
<i>taurinus</i>	Lophiohylini	85.0
<i>Trachycephalus</i>		
<i>venulosus</i>	Lophiohylini	100.5
<i>Phyllomedusa bicolor</i>	Phyllomedusinae	115.0
<i>Phyllomedusa</i>		
<i>hypochondrialis</i>	Phyllomedusinae	37.7
<i>Phyllomedusa</i>		
<i>vaillanti</i>	Phyllomedusinae	59.9
<i>Scinax baumgardneri</i>	<i>Scinax</i> clade	29.0
<i>Scinax boesemanni</i>	<i>Scinax</i> clade	31.1
<i>Scinax egleri</i>	<i>Scinax</i> clade	30.0
<i>Scinax ruber</i>	<i>Scinax</i> clade	41.2
<i>Scinax sp. 1</i>	<i>Scinax</i> clade	

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	<i>Scinax sp. 2</i>	<i>Scinax</i> clade	
	<i>Sphaenorhynchus</i>		
	<i>lacteus</i>	<i>Scinax</i> clade	41.5
Ecuador: Napo: Santa Cecilia	<i>Hypsiboas boans</i>	Cophomantini	132.0
	<i>Hypsiboas calcarata</i>	Cophomantini	47.5
	<i>Hypsiboas fasciata</i>	Cophomantini	40.3
	<i>Hypsiboas geographica</i>	Cophomantini	62.0
	<i>Hypsiboas granosa</i>	Cophomantini	44.0
	<i>Hypsiboas lanciformis</i>	Cophomantini	80.0
	<i>Hypsiboas punctata</i>	Cophomantini	40.0
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>bifurcus</i>	clade	28.0
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>bokermanni</i>	clade	24.0
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>brevifrons</i>	clade	22.0
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>leucophyllatus</i>	clade	44.0
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>marmorata</i>	clade	44.0
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	23.0

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<i>minuta</i>	clade	
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>parviceps</i>	clade	21.9
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>rhodopeplus</i>	clade	24.2
	<i>Dendropsophus</i>	
<i>Dendropsophus riveroi</i>	clade	20.0
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>rossalleni</i>	clade	20.0
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>sarayacuensis</i>	clade	29.0
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>triangulum</i>	clade	28.0
<i>Osteocephalus</i>		
<i>alboguttatus</i>	Lophiohylini	34.0
<i>Osteocephalus</i>		
<i>buckleyi</i>	Lophiohylini	64.1
<i>Osteocephalus</i>		
<i>leprieurii</i>	Lophiohylini	48.0
<i>Osteocephalus</i>		
<i>taurinus</i>	Lophiohylini	85.0

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	<i>Nyctimantis rugiceps</i>	Lophiohylini	67.5
	<i>Trachycephalus</i>		
	<i>coriaceus</i>	Lophiohylini	63.0
	<i>Trachycephalus</i>		
	<i>venulosus</i>	Lophiohylini	100.5
	<i>Phyllomedusa palliata</i>	Phyllomedusinae	49.1
	<i>Phyllomedusa tarsius</i>	Phyllomedusinae	97.0
	<i>Phyllomedusa</i>		
	<i>tomopterna</i>	Phyllomedusinae	48.0
	<i>Phyllomedusa</i>		
	<i>vaillanti</i>	Phyllomedusinae	59.9
	<i>Scinax cruentommus</i>	<i>Scinax</i> clade	28.0
	<i>Scinax funereus</i>	<i>Scinax</i> clade	37.0
	<i>Scinax garbei</i>	<i>Scinax</i> clade	42.2
	<i>Scinax ruber</i>	<i>Scinax</i> clade	41.2
	<i>Sphaenorhynchus</i>		
	<i>carneus</i>	<i>Scinax</i> clade	20.0
	<i>Sphaenorhynchus</i>		
	<i>lacteus</i>	<i>Scinax</i> clade	41.5
Peru: Huanoco: Panguana	<i>Hypsiboas boans</i>	Cophomantini	132.0
	<i>Hypsiboas fasciatus</i>	Cophomantini	40.3

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<i>Hypsiboas granosa</i>	Cophomantini	44.0
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>brevifrons</i>	clade	22.0
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>leucophyllatus</i>	clade	44.0
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>marmoratus</i>	clade	44.0
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>parviceps</i>	clade	21.9
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>rhodopeplus</i>	clade	24.2
	<i>Dendropsophus</i>	
<i>Dendropsophus riveroi</i>	clade	20.0
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>rossalleni</i>	clade	20.0
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>sarayacuensis</i>	clade	29.0
<i>Osteocephalus</i>		
<i>leprieurii</i>	Lophiohylini	48.0
<i>Osteocephalus</i>		
<i>taurinus</i>	Lophiohylini	85.0

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	<i>Trachycephalus</i>		
	<i>coriaceus</i>	Lophiohylini	63.0
	<i>Trachycephalus</i>		
	<i>venulosus</i>	Lophiohylini	100.5
	<i>Phyllomedusa tarsius</i>	Phyllomedusinae	97.0
	<i>Phyllomedusa</i>		
	<i>tomopterna</i>	Phyllomedusinae	48.0
	<i>Phyllomedusa</i>		
	<i>vaillanti</i>	Phyllomedusinae	59.9
	<i>Scinax cruentomma</i>	<i>Scinax</i> clade	28.0
	<i>Scinax garbei</i>	<i>Scinax</i> clade	42.2
	<i>Scinax rubra</i>	<i>Scinax</i> clade	41.2
Peru: Loreto: Teniente Lopez	<i>Hypsiboas boans</i>	Cophomantini	132.0
	<i>Hypsiboas calcaratus</i>	Cophomantini	47.5
	<i>Hypsiboas fasciatus</i>	Cophomantini	40.3
	<i>Hypsiboas</i>		
	<i>geographicus</i>	Cophomantini	62.0
	<i>Hypsiboas lanciformis</i>	Cophomantini	80.0
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>brevifrons</i>	clade	22.0
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	24.0

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<i>koechlini</i>	clade	
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>leucophyllata</i>	clade	44.0
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>marmorata</i>	clade	44.0
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>parviceps</i>	clade	21.9
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>rhodopepla</i>	clade	24.2
	<i>Dendropsophus</i>	
<i>Dendropsophus riveroi</i>	clade	20.0
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>sarayacuensis</i>	clade	29.0
<i>Osteocephalus</i>		
<i>buckleyi</i>	Lophiohylini	64.1
<i>Osteocephalus</i>		
<i>leprieurii</i>	Lophiohylini	48.0
<i>Osteocephalus</i>		
<i>planiceps</i>	Lophiohylini	65.9
<i>Osteocephalus</i>		
<i>taurinus</i>	Lophiohylini	85.0

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	<i>Osteocephalus sp.</i>	Lophiohylini	
	<i>Phyllomedusa coelestis</i>	Phyllomedusinae	64.8
	<i>Phyllomedusa hulli</i>	Phyllomedusinae	37.1
	<i>Phyllomedusa tarsius</i>	Phyllomedusinae	97.0
	<i>Phyllomedusa</i>		
	<i>tomopterna</i>	Phyllomedusinae	48.0
	<i>Phyllomedusa</i>		
	<i>vaillanti</i>	Phyllomedusinae	59.9
	<i>Scinax cruentommus</i>	<i>Scinax</i> clade	28.0
	<i>Scinax funereus</i>	<i>Scinax</i> clade	37.0
	<i>Scinax ruber</i>	<i>Scinax</i> clade	41.2
Peru: Madre de Dios: Manu			
National Park (Cocha Cashu)	<i>Hypsiboas boans</i>	Cophomantini	132.0
	<i>Hypsiboas calcaratus</i>	Cophomantini	47.5
	<i>Hypsiboas fasciatus</i>	Cophomantini	40.3
	<i>Hypsiboas</i>		
	<i>geographicus</i>	Cophomantini	62.0
	<i>Hypsiboas granosa</i>	Cophomantini	44.0
	<i>Hypsiboas punctatus</i>	Cophomantini	40.0
	<i>Hypsiboas sp.</i>		
	( <i>geographica</i> group)	Cophomantini	

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<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>bifurcus</i>	clade	28.0
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>koechlini</i>	clade	24.0
	<i>Dendropsophus</i>	
<i>Dendropsophus leali</i>	clade	23.0
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>leucophyllatus</i>	clade	44.0
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>parviceps</i>	clade	21.9
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>rhodopeplus</i>	clade	24.2
	<i>Dendropsophus</i>	
<i>Dendropsophus riveroi</i>	clade	20.0
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>sarayacuensis</i>	clade	29.0
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>triangulum</i>	clade	28.0
<i>Osteocephalus</i>		
<i>leprieurii</i>	Lophiohylini	48.0
<i>Osteocephalus</i>	Lophiohylini	85.0

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<i>taurinus</i>		
<i>Trachycephalus</i>		
<i>coriaceus</i>	Lophiohylini	63.0
<i>Osteocephalus</i>		
<i>resinifictrix</i>	Lophiohylini	76.0
<i>Osteocephalus</i>		
<i>venulosus</i>	Lophiohylini	100.5
<i>Cruziohylla</i>		
<i>craspedopus</i>	Phyllomedusinae	57.0
<i>Phyllomedusa</i>		
<i>atelopoides</i>	Phyllomedusinae	37.4
<i>Phyllomedusa palliata</i>	Phyllomedusinae	49.1
<i>Phyllomedusa</i>		
<i>tomopterna</i>	Phyllomedusinae	48.0
<i>Phyllomedusa</i>		
<i>vaillanti</i>	Phyllomedusinae	59.9
<i>Phyllomedusa camba</i>	Phyllomedusinae	70.0
<i>Scarthylla goinorum</i>	Pseudis clade	21.0
<i>Scinax cruentommus</i>	Scinax clade	33.5
<i>Scinax epacrorrhinus</i>	Scinax clade	31.5
<i>Scinax garbei</i>	Scinax clade	49.1

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	<i>Scinax ruber</i>	<i>Scinax</i> clade	45.0
	<i>Sphaenorhynchus</i>		
	<i>dorisae</i>	<i>Scinax</i> clade	29.0
	<i>Sphaenorhynchus</i>		
	<i>lacteus</i>	<i>Scinax</i> clade	41.5
Peru: Madre de Dios: Cuzco			
Amazonico Reserve	<i>Hypsiboas boans</i>	Cophomantini	132.0
	<i>Hypsiboas calcaratus</i>	Cophomantini	47.5
	<i>Hypsiboas fasciatus</i>	Cophomantini	40.3
	<i>Hypsiboas</i>		
	<i>geographicus</i>	Cophomantini	62.0
	<i>Hypsiboas granosa</i>	Cophomantini	44.0
	<i>Hypsiboas punctatus</i>	Cophomantini	40.0
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>allenorum</i>	clade	21.4
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>bokermanni</i>	clade	24.0
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>brevifrons</i>	clade	22.0
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>koechlini</i>	clade	24.0

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	<i>Dendropsophus</i>	
<i>Dendropsophus leali</i>	clade	23.0
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>leucophyllatus</i>	clade	44.0
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>marmoratus</i>	clade	44.0
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>parviceps</i>	clade	21.9
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>rhodopeplus</i>	clade	24.2
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>schubarti</i>	clade	19.5
<i>Osteocephalus</i>		
<i>leprieuri</i>	Lophiohylini	48.0
<i>Osteocephalus</i>		
<i>taurinus</i>	Lophiohylini	85.0
<i>Trachycephalus</i>		
<i>coriaceus</i>	Lophiohylini	63.0
<i>Trachycephalus</i>		
<i>venulosus</i>	Lophiohylini	100.5
<i>Cruzihyla craspedopus</i>	Phyllomedusinae	57.0

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	<i>Phyllomedusa</i>		
	<i>atelopoides</i>	Phyllomedusinae	37.4
	<i>Phyllomedusa camba</i>	Phyllomedusinae	70.0
	<i>Phyllomedusa palliata</i>	Phyllomedusinae	49.1
	<i>Phyllomedusa</i>		
	<i>tomopterna</i>	Phyllomedusinae	48.0
	<i>Phyllomedusa</i>		
	<i>vaillanti</i>	Phyllomedusinae	59.9
	<i>Pseudis paradoxa</i>	Pseudis clade	21.0
	<i>Scarthyla goinorum</i>	Pseudis clade	55.0
	<i>Scinax chiquitanus</i>	<i>Scinax</i> clade	36.2
	<i>Scinax garbei</i>	<i>Scinax</i> clade	49.1
	<i>Scinax ictericus</i>	<i>Scinax</i> clade	33.5
	<i>Scinax pedromedinai</i>	<i>Scinax</i> clade	31.5
	<i>Scinax ruber</i>	<i>Scinax</i> clade	45.0
	<i>Sphaenorhynchus</i>		
	<i>lacteus</i>	<i>Scinax</i> clade	41.5
Venezuela: Bolivar: El			
Manteco	<i>Hypsiboas crepitans</i>	Cophomantini	63.0
	<i>Hypsiboas</i>		
	<i>geographica</i>	Cophomantini	62.0

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	<i>Hypsiboas</i>		
	<i>multifasciata</i>	Cophomantini	57.3
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>microcephalus</i>	clade	24.5
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>minusculus</i>	clade	20.6
	<i>Trachycephalus</i>		
	<i>venulosus</i>	Lophiohylini	100.5
	<i>Phyllomedusa</i>		
	<i>hypochondrialis</i>	Phyllomedusinae	37.7
	<i>Pseudis paradoxus</i>	<i>Pseudis</i> clade	55.0
	<i>Scinax trilineatus</i>	<i>Scinax</i> clade	22.5
	<i>Scinax x-signatus</i>	<i>Scinax</i> clade	42.6
	<i>Sphaenorhynchus</i>		
	<i>lacteus</i>	<i>Scinax</i> clade	41.5
Venezuela: Gaurico: Hato			
Masaguaral	<i>Hypsiboas crepitans</i>	Cophomantini	63.0
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>microcephalus</i>	clade	24.5
	<i>Trachycephalus</i>		
	<i>venulosus</i>	Lophiohylini	100.5

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	<i>Pseudis paradoxus</i>	Pseudis clade	55.0
	<i>Scinax ruber</i>	<i>Scinax</i> clade	41.2
Venezuela: La Escalera Region	<i>Hypsiboas boans</i>	Cophomantini	132.0
	<i>Hypsiboas crepitans</i>	Cophomantini	63.0
	<i>Hypsiboas</i>		
	<i>geographicus</i>	Cophomantini	62.0
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>minusculus</i>	clade	20.6
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>minutus</i>	clade	23.0
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>microcephalus</i>	clade	24.5
	<i>Osteocephalus</i>		
	<i>taurinus</i>	Lophiohylini	85.0
	<i>Trachycephalus</i>		
	<i>venulosus</i>	Lophiohylini	100.5
	<i>Phyllomedusa bicolor</i>	Phyllomedusinae	115.0
	<i>Phyllomedusa</i>		
	<i>hypochondrialis</i>	Phyllomedusinae	37.7
	<i>Phyllomedusa tarsi</i>	Phyllomedusinae	97.0
	<i>Phyllomedusa</i>	Phyllomedusinae	48.0

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	<i>tomopterna</i>		
	<i>Scinax rostratus</i>	<i>Scinax</i> clade	45.7
	<i>Scinax ruber</i>	<i>Scinax</i> clade	41.2
	<i>Scinax x-signatus</i>	<i>Scinax</i> clade	42.5
	<i>Sphaenorhynchus</i>		
	<i>lacteus</i>	<i>Scinax</i> clade	41.5
Venezuela: Sierra de Lema	<i>Hypsiboas boans</i>	Cophomantini	132.0
	<i>Hypsiboas lemai</i>	Cophomantini	30.2
	<i>Hypsiboas</i>		
	<i>multifasciata</i>	Cophomantini	57.3
	<i>Hypsiboas sibleszi</i>	Cophomantini	34.9
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>minutus</i>	clade	23.0
	<i>Scinax danae</i>	<i>Scinax</i> clade	27.4
Argentina: La Rioja: Chilecito	<i>Hypsiboas riojanus</i>	Cophomantini	56.0
Colombia: Boyaca: Parama de la Rusia	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>labialis</i>	clade	43.0
	<i>Hyloscirtus</i>		
Ecuador: Napo: Rio Salado	<i>phyllognathus</i>	Cophomantini	34.0
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>brevifrons</i>	clade	22.0

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	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>sarayacuensis</i>	clade	29.0
	<i>Osteocephalus</i>		
	<i>verrucigerus</i>	Lophiohylini	58.6
	<i>Hylomantis buckleyi</i>	Phyllomedusinae	44.5
	<i>Phyllomedusa</i>		
	<i>perinesos</i>	Phyllomedusinae	51.5
	<i>Scinax ruber</i>	<i>Scinax</i> clade	41.2
Ecuador: Pichincha: Quebrada			
Zapadores	<i>Hyloscirtus alytolylax</i>	Cophomantini	37.0
Peru: Cuzco: Rio Cosnipata	<i>Hyloscirtus armatus</i>	Cophomantini	68.5
	<i>Hyloscirtus</i>		
	<i>phyllognatha</i>	Cophomantini	34.0
	<i>Hypsiboas balzani</i>	Cophomantini	50.4
China: Fujian Prov.: Kuliang,			
near Fuzhou;	<i>Hyla chinensis</i>	Hylini	32.0
China: Guangdong Prov.: Tai-			
Yong, E Kwantun	<i>Hyla chinensis</i>	Hylini	32.0
China: Guangdong Prov.: Yim-			
Na-San [=Yin Na S	<i>Hyla chinensis</i>	Hylini	32.0
China: Guangdong:	<i>Hyla simplex</i>	Hylini	34.0

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 Dinghushan: Cha Chang

China: Jiangxi: Hong-San, SE

Kiangsi Prov.	<i>Hyla chinensis</i>	Hylini	32.0
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China: Yunnan Province, Nu

Jiang Prefecture	<i>Hyla annectans</i>	Hylini	35.0
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China: Yunnan, Baoshan

Prefecture, Qushi	<i>Hyla annectans</i>	Hylini	35.0
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Japan: Kanagawa: Odawara

City	<i>Hyla japonica</i>	Hylini	39.0
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Japan: Miyagi Pref.: Honshu:

Takayama, near Matushima

Bay	<i>Hyla japonica</i>	Hylini	39.0
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Korea: Kyonggi Prov.; Mt.

Buckak, near Seoul	<i>Hyla japonica</i>	Hylini	39.0
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Myanmar: Chin State: Falam

Township, Lon Pi village	<i>Hyla annectans</i>	Hylini	35.0
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Australia: New South Wales:

Mutawintji National Park	<i>Litoria careulea</i>	Pelodryadinae	80.0
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	<i>Litoria rubella</i>	Pelodryadinae	37.0
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Australia: New South Wales:

Northern Yengo National Park	<i>Litoria caerulea</i>	Pelodryadinae	80.0
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	<i>Litoria dentata</i>	Pelodryadinae	40.0
	<i>Litoria fallax</i>	Pelodryadinae	26.0
	<i>Litoria latopalmata</i>	Pelodryadinae	39.0
	<i>Litoria lesueuri</i>	Pelodryadinae	43.0
	<i>Litoria peronii</i>	Pelodryadinae	53.0
Australia: Northern Territory:			
Kakadu National Park	<i>Cyclorana australis</i>	Pelodryadinae	79.0
	<i>Cyclorana longipes</i>	Pelodryadinae	46.0
	<i>Litoria bicolour</i>	Pelodryadinae	27.0
	<i>Litoria caerulea</i>	Pelodryadinae	80.0
	<i>Litoria coplandi</i>	Pelodryadinae	36.0
	<i>Litoria dahlia</i>	Pelodryadinae	63.0
	<i>Litoria inermis</i>	Pelodryadinae	33.0
	<i>Litoria meiriana</i>	Pelodryadinae	20.0
	<i>Litoria microbelos</i>	Pelodryadinae	16.0
	<i>Litoria nasuta</i>	Pelodryadinae	45.0
	<i>Litoria pallida</i>	Pelodryadinae	34.0
	<i>Litoria personata</i>	Pelodryadinae	29.0
	<i>Litoria rothii</i>	Pelodryadinae	48.0
	<i>Litoria rubella</i>	Pelodryadinae	37.0
	<i>Litoria tornieri</i>	Pelodryadinae	36.0

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 Australia: Northern Territory:

Nitmiluk National Park	<i>Cyclorana australis</i>	Pelodryadinae	79.0
	<i>Cyclorana longipes</i>	Pelodryadinae	46.0
	<i>Cyclorana maculosa</i>	Pelodryadinae	55.0
	<i>Cyclorana</i>		
	<i>platycephala</i>	Pelodryadinae	64.0
	<i>Litoria bicolor</i>	Pelodryadinae	27.0
	<i>Litoria caerulea</i>	Pelodryadinae	80.0
	<i>Litoria coplandi</i>	Pelodryadinae	36.0
	<i>Litoria inermis</i>	Pelodryadinae	33.0
	<i>Litoria meiriana</i>	Pelodryadinae	20.0
	<i>Litoria microbelos</i>	Pelodryadinae	16.0
	<i>Litoria nasuta</i>	Pelodryadinae	45.0
	<i>Litoria pallida</i>	Pelodryadinae	34.0
	<i>Litoria personata</i>	Pelodryadinae	29.0
	<i>Litoria rothii</i>	Pelodryadinae	48.0
	<i>Litoria rubella</i>	Pelodryadinae	37.0
	<i>Litoria tornieri</i>	Pelodryadinae	36.0
	<i>Litoria wotjulumensis</i>	Pelodryadinae	38.0

## Australia: Queensland:

Lamington National Park	<i>Litoria caerulea</i>	Pelodryadinae	80.0
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	<i>Litoria chloris</i>	Pelodryadinae	62.0
	<i>Litoria dentata</i>	Pelodryadinae	40.0
	<i>Litoria fallax</i>	Pelodryadinae	26.0
	<i>Litoria gracilentata</i>	Pelodryadinae	42.0
	<i>Litoria latopalmata</i>	Pelodryadinae	39.0
	<i>Litoria lesuerii</i>	Pelodryadinae	43.0
	<i>Litoria nasuta</i>	Pelodryadinae	45.0
	<i>Litoria pearsoniana</i>	Pelodryadinae	29.0
	<i>Litoria peroni</i>	Pelodryadinae	53.0
	<i>Litoria revelata</i>	Pelodryadinae	28.0
	<i>Litoria rubella</i>	Pelodryadinae	37.0
	<i>Litoria tyleri</i>	Pelodryadinae	48.0
	<i>Litoria verreauxi</i>	Pelodryadinae	36.0
Australia: Queensland:	<i>Cyclorana</i>		
Townsville	<i>novaehollandiae</i>	Pelodryadinae	81.0
	<i>Cyclorana brevipes</i>	Pelodryadinae	45.0
	<i>Litoria alboguttata</i>	Pelodryadinae	67.0
	<i>Litoria bicolor</i>	Pelodryadinae	27.0
	<i>Litoria caerulea</i>	Pelodryadinae	80.0
	<i>Litoria fallax</i>	Pelodryadinae	26.0
	<i>Litoria gracilentata</i>	Pelodryadinae	42.0

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	<i>Litoria inermis</i>	Pelodryadinae	33.0
	<i>Litoria nasuta</i>	Pelodryadinae	45.0
	<i>Litoria rothii</i>	Pelodryadinae	48.0
	<i>Litoria rubella</i>	Pelodryadinae	37.0
	<i>Litoria wilcoxii</i>	Pelodryadinae	
Australia: Queensland: Cairns	<i>Litoria bicolor</i>	Pelodryadinae	27.0
	<i>Litoria caerulea</i>	Pelodryadinae	80.0
	<i>Litoria genimaculata</i>	Pelodryadinae	41.0
	<i>Litoria gracilentata</i>	Pelodryadinae	42.0
	<i>Litoria infrafronata</i>	Pelodryadinae	102.0
	<i>Litoria jungguy</i>	Pelodryadinae	
	<i>Litoria microbelos</i>	Pelodryadinae	16.0
	<i>Litoria nannotis</i>	Pelodryadinae	48.0
	<i>Litoria nasuta</i>	Pelodryadinae	45.0
	<i>Litoria rheocola</i>	Pelodryadinae	32.0
	<i>Litoria rothii</i>	Pelodryadinae	48.0
	<i>Litoria rubella</i>	Pelodryadinae	37.0
	<i>Litoria xanthomera</i>	Pelodryadinae	56.0
	<i>Nyctimystes dayi</i>	Pelodryadinae	42.0
Australia: Queensland:			
Kuranda	<i>Litoria bicolor</i>	Pelodryadinae	27.0

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	<i>Litoria caerulea</i>	Pelodryadinae	80.0
	<i>Litoria genimaculata</i>	Pelodryadinae	41.0
	<i>Litoria gracilentata</i>	Pelodryadinae	42.0
	<i>Litoria infrafrinata</i>	Pelodryadinae	102.0
	<i>Litoria jungguy</i>	Pelodryadinae	
	<i>Litoria microbelos</i>	Pelodryadinae	16.0
	<i>Litoria myola</i>	Pelodryadinae	
	<i>Litoria nasuta</i>	Pelodryadinae	45.0
	<i>Litoria rheocola</i>	Pelodryadinae	32.0
	<i>Litoria rothii</i>	Pelodryadinae	48.0
	<i>Litoria rubella</i>	Pelodryadinae	37.0
	<i>Litoria xanthomera</i>	Pelodryadinae	56.0
	<i>Nyctimystes dayi</i>	Pelodryadinae	42.0
Australia: Queensland:			
Dinden Forest Reserve			
	<i>Litoria bicolor</i>	Pelodryadinae	27.0
	<i>Litoria caerulea</i>	Pelodryadinae	80.0
	<i>Litoria fallax</i>	Pelodryadinae	26.0
	<i>Litoria genimaculata</i>	Pelodryadinae	41.0
	<i>Litoria gracilentata</i>	Pelodryadinae	42.0
	<i>Litoria jungguy</i>	Pelodryadinae	
	<i>Litoria microbelos</i>	Pelodryadinae	16.0

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	<i>Litoria nannotis</i>	Pelodryadinae	48.0
	<i>Litoria nasuta</i>	Pelodryadinae	45.0
	<i>Litoria latopalmata</i>	Pelodryadinae	39.0
	<i>Litoria rheocola</i>	Pelodryadinae	32.0
	<i>Litoria rothii</i>	Pelodryadinae	48.0
	<i>Litoria rubella</i>	Pelodryadinae	37.0
	<i>Litoria xanthomera</i>	Pelodryadinae	56.0
	<i>Nyctimystes dayi</i>	Pelodryadinae	42.0
Australia: Queensland:			
Wooroonooran National Park	<i>Litoria genimaculata</i>	Pelodryadinae	41.0
	<i>Litoria infrafrenata</i>	Pelodryadinae	102.0
	<i>Litoria jungguy</i>	Pelodryadinae	
	<i>Litoria nannotis</i>	Pelodryadinae	48.0
	<i>Litoria revelata</i>	Pelodryadinae	28.0
	<i>Litoria rheocola</i>	Pelodryadinae	32.0
	<i>Litoria xanthomera</i>	Pelodryadinae	56.0
	<i>Nyctimystes dayi</i>	Pelodryadinae	42.0
New Guinea: Lakekamu Basin,			
Ivimka Research	<i>Litoria dorsalis</i>	Pelodryadinae	24.0
	<i>Litoria genimaculata</i>	Pelodryadinae	41.0
	<i>Litoria infrafrenata</i>	Pelodryadinae	102.0

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	<i>Litoria modica</i>	Pelodryadinae	30.0
	<i>Litoria pygmaea</i>	Pelodryadinae	30.0
	<i>Litoria sp. 1</i>	Pelodryadinae	
	<i>Litoria sp. 2</i>	Pelodryadinae	
	<i>Litoria sp. 3</i>	Pelodryadinae	
	<i>Nyctimystes</i>		
	<i>cheesmani</i>	Pelodryadinae	60.0
	<i>Litoria cf.</i>		
New Guinea: Utai	<i>genimaculata</i>	Pelodryadinae	41.0
	<i>Litoria huntorum</i>	Pelodryadinae	
	<i>Litoria infrafrenata</i>	Pelodryadinae	102.0
	<i>Litoria nigropunctata</i>	Pelodryadinae	32.0
	<i>Litoria thesaurensis</i>	Pelodryadinae	50.0
	<i>Litoria sp.</i>	Pelodryadinae	
Argentina: Buenos Aires:	<i>Hypsiboas pulchella</i>		
Bahia Blanca	<i>pulchella</i>	Cophomantini	50.0
Argentina: Cordoba: Capilla	<i>Hypsiboas pulchella</i>		
de Monte	<i>cordoba</i>	Cophomantini	50.0
Argentina: Parque Nacional			
Chaco	<i>Hypsiboas raniceps</i>	Cophomantini	71.0
	<i>Dendropsophus nanus</i>	<i>Dendropsophus</i>	22.0

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		clade	
	<i>Trachycephalus</i>		
	<i>venulosus</i>	Lophiohylini	100.5
	<i>Phyllomedusa azurea</i>	Phyllomedusinae	41.7
	<i>Lysapsus limellus</i>	Pseudis clade	20.0
	<i>Pseudis paradoxus</i>		
	<i>platensis</i>	Pseudis clade	55.0
	<i>Scinax acuminatus</i>	Scinax clade	45.0
	<i>Scinax fuscovarius</i>	Scinax clade	47.1
	<i>Scinax nasicus</i>	Scinax clade	37.0
	<i>Scinax squalirostris</i>	Scinax clade	29.0
Brazil: Goias: Espora Power	<i>Hypsiboas</i>		
Plant	<i>albopunctatus</i>	Cophomantini	60.0
	<i>Hypsiboas lundii</i>	Cophomantini	76.0
	<i>Hypsiboas</i>		
	<i>multifasciatus</i>	Cophomantini	57.3
	<i>Hypsiboas raniceps</i>	Cophomantini	71.0
		<i>Dendropsophus</i>	
	<i>Dendropsophus cruzi</i>	clade	19.4
		<i>Dendropsophus</i>	
	<i>Dendropsophus jimi</i>	clade	20.9

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	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>minutus</i>	clade	23.0
		<i>Dendropsophus</i>	
	<i>Dendropsophus nanus</i>	clade	22.0
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>rubicundulus</i>	clade	23.8
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>soaresi</i>	clade	31.7
	<i>Trachycephalus</i>		
	<i>venulosus</i>	Lophiohylini	100.5
	<i>Phyllomedusa azurea</i>	Phyllomedusinae	41.7
	<i>Pseudis bolbodactylus</i>	<i>Pseudis</i> clade	45.0
	<i>Scinax</i>		
	<i>fuscumarginatus</i>	<i>Scinax</i> clade	24.0
	<i>Scinax fuscovarius</i>	<i>Scinax</i> clade	47.1
	<i>Scinax x-signatus</i>	<i>Scinax</i> clade	42.5
Brazil: Mato Grosso:			
Dardanelos Dam	<i>Hypsiboas boans</i>	Cophomantini	132.0
	<i>Hypsiboas calcaratus</i>	Cophomantini	47.5
	<i>Hypsiboas cinerascans</i>	Cophomantini	44.0
	<i>Hypsiboas cf.</i>	Cophomantini	62.0

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<i>geographicus</i>		
<i>Hypsiboas fasciatus</i>	Cophomantini	40.3
<i>Hypsiboas lanciformis</i>	Cophomantini	80.0
<i>Hypsiboas sp.</i>	Cophomantini	
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>minutus</i>	clade	23.0
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>parviceps</i>	clade	21.9
<i>Dendropsophus aff.</i>	<i>Dendropsophus</i>	
<i>microcephalus</i>	clade	24.5
	<i>Dendropsophus</i>	
<i>Dendropsophus sp.</i>	clade	
<i>Osteocephalus</i>		
<i>leprieurii</i>	Lophiohylini	48.0
<i>Osteocephalus</i>		
<i>taurnius</i>	Lophiohylini	85.0
<i>Phyllomedusa</i>		
<i>boliviana</i>	Phyllomedusinae	70.5
<i>Phyllomedusa</i>		
<i>vaillanti</i>	Phyllomedusinae	59.9
<i>Scinax boesemani</i>	<i>Scinax</i> clade	31.1

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	<i>Scinax catharinae</i>		
	group	<i>Scinax</i> clade	
	<i>Scinax garbei</i>	<i>Scinax</i> clade	42.2
	<i>Scinax nebulosus</i>	<i>Scinax</i> clade	30.0
	<i>Scinax ruber</i>	<i>Scinax</i> clade	41.2
	<i>Scinax ruber</i> group	<i>Scinax</i> clade	
Brazil: Rio Grande do Sul:			
Parque Nacional Aparados da Serra	<i>Aplastodiscus</i>		
	<i>peroviridis</i>	Cophomantini	46.1
	<i>Hypsiboas bischoffi</i>	Cophomantini	46.1
	<i>Hypsiboas faber</i>	Cophomantini	104.0
	<i>Hypsiboas</i>		
	<i>leptolineatus</i>	Cophomantini	31.6
	<i>Hypsiboas marginatus</i>	Cophomantini	51.1
	<i>Hypsiboas pulchellus</i>	Cophomantini	50.0
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>microps</i>	clade	26.0
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>minutus</i>	clade	23.0
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>sanborni</i>	clade	17.0

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	<i>Pseudis cardosoi</i>	Pseudis clade	45.9
	<i>Scinax berthae</i>	Scinax clade	22.2
	<i>Scinax catharinae</i>	Scinax clade	35.1
	<i>Scinax granulatus</i>	Scinax clade	36.0
	<i>Scinax pereracus</i>	Scinax clade	38.5
	<i>Scinax squalirostris</i>	Scinax clade	29.0
	<i>Scinax sp.</i>	Scinax clade	
Brazil: Tocantins: Lajeado,			
Luis Eduardo Magalhaes Dam	<i>Hypsiboas punctatus</i>	Cophomantini	40.0
	<i>Hypsiboas raniceps</i>	Cophomantini	71.0
	<i>Hypsiboas wavrini</i>	Cophomantini	113.0
	<i>Hypsiboas sp.</i>		
	( <i>pulchella</i> group)	Cophomantini	
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>anataliasiasi</i>	clade	21.8
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>minutus</i>	clade	23.0
	<i>Dendropsophus sp.</i>	<i>Dendropsophus</i>	
	( <i>nanus</i> group)	clade	
	<i>Dendropsophus sp.</i>	<i>Dendropsophus</i>	
	( <i>rubicundulus</i> group)	clade	

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	<i>Osteocephalus cf.</i>		
	<i>leprieuri</i>	Lophiohylini	48.0
	<i>Trachycephalus</i>		
	<i>venulosus</i>	Lophiohylini	100.5
	<i>Pseudis tocantins</i>	Pseudis clade	
	<i>Scinax</i>		
	<i>fuscumarginatus</i>	<i>Scinax</i> clade	24.0
	<i>Scinax fuscovaria</i>	<i>Scinax</i> clade	47.1
	<i>Scinax sp. 1 (rostratus</i>		
	<i>group)</i>	<i>Scinax</i> clade	
	<i>Scinax sp. 2</i>		
	<i>(catharinae group)</i>	<i>Scinax</i> clade	
	<i>Scinax sp. 3 (ruber</i>		
	<i>group)</i>	<i>Scinax</i> clade	
	<i>Scinax sp. 4 (ruber</i>		
	<i>group)</i>	<i>Scinax</i> clade	
Uruguay: Montevideo: Pajas			
Blancas	<i>Hypsiboas pulchella</i>	Cophomantini	50.0
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>sanborni</i>	clade	17.0
	<i>Scinax berthae</i>	<i>Scinax</i> clade	22.2

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	<i>Scinax granulatus</i>	<i>Scinax</i> clade	36.0
	<i>Scinax squalirostris</i>	<i>Scinax</i> clade	29.0
Uruguay: Salto: El Espinillar	<i>Hypsiboas pulchella</i>	Cophomantini	50.0
		<i>Dendropsophus</i>	
	<i>Dendropsophus nanus</i>	clade	22.0
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>sanborni</i>	clade	17.0
	<i>Lysapsus limellus</i>	Pseudis clade	20.0
	<i>Pseudis minuta</i>	<i>Pseudis</i> clade	40.0
	<i>Scinax fuscovarius</i>	<i>Scinax</i> clade	47.1
	<i>Scinax granulatus</i>	<i>Scinax</i> clade	36.0
	<i>Scinax nasicus</i>	<i>Scinax</i> clade	37.0
	<i>Scinax squalirostris</i>	<i>Scinax</i> clade	29.0
Ecuador: Los Rios: Rio			
Palenque	<i>Hypsiboas pellucens</i>	Cophomantini	61.6
	<i>Hypsiboas picturata</i>	Cophomantini	
	<i>Hypsiboas rosenbergi</i>	Cophomantini	90.0
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>gryllatus</i>	clade	25.5
	<i>Smilisca phaeota</i>	Hylini	65.5
	<i>Trachycephalus</i>	Lophiohylini	75.9

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<i>jordani</i>				
	<i>Agalychnis litodryas</i>	Phyllomedusinae	70.2	
	<i>Scinax quinquefasciata</i>	<i>Scinax</i> clade	30.0	
	<i>Scinax sugillatus</i>	<i>Scinax</i> clade	42.0	
	<i>Scinax</i> sp.	<i>Scinax</i> clade		
France: Provence-Alpes-Cote-				
d'Azur; 3 mi W				
	<i>Hyla arborea</i>	Hylini	50.0	
Georgia; ca. 10 km SSE				
	Borzhomi	<i>Hyla arborea</i>	Hylini	50.0
Germany: Rostock,				
Mecklenburg				
	<i>Hyla arborea</i>	Hylini	50.0	
Hungary: Budapest				
	<i>Hyla arborea</i>	Hylini	50.0	
Morocco: Settat; Ben-Slimane,				
5 km N of				
	<i>Hyla meridionalis</i>	Hylini	50.0	
Spain: Andalusia: Cadiz				
Prov.: Benalup de Sidonia				
	<i>Hyla meridionalis</i>	Hylini	50.0	
Ukraine: Kiev				
	<i>Hyla arborea</i>	Hylini	50.0	
Venezuela: Gran Sabana				
	<i>Hypsiboas boans</i>	Cophomantini	132.0	
	<i>Hypsiboas crepitans</i>	Cophomantini	63.0	
	<i>Hypsiboas lemai</i>	Cophomantini	30.2	
	<i>Hypsiboas</i>	Cophomantini	57.3	

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	<i>multifasciatus</i>		
	<i>Hypsiboas sibleszi</i>	Cophomantini	34.9
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>minutus</i>	clade	23.0
	<i>Tepuihyla rodriguezi</i>	Lophiohylini	34.7
	<i>Scinax exigua</i>	<i>Scinax</i> clade	20.8
Costa Rica: Cartago: Moravia	<i>Hyloscirtus colymba</i>	Cophomantini	37.0
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>ebraccatus</i>	clade	27.8
	<i>Anotheca spinosa</i>	Hylini	68.5
	<i>Duellmanohyla</i>		
	<i>rufioculis</i>	Hylini	30.0
	<i>Duellmanohyla</i>		
	<i>uranochroa</i>	Hylini	36.8
	<i>Isthmohyla lancasteri</i>	Hylini	33.6
	<i>Isthmohyla</i>		
	<i>pseudopuma</i>	Hylini	41.4
	<i>Smilisca phaeota</i>	Hylini	65.5
	<i>Tlalocohyla loquax</i>	Hylini	44.7
	<i>Agalychnis annae</i>	Phyllomedusinae	73.9
	<i>Hylomantis lemur</i>	Phyllomedusinae	40.8

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Costa Rica: Heredia: La Selva	<i>Hypsiboas rufitellus</i>	Cophomantini	49.2
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>ebzacattus</i>	clade	27.8
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>phlebodes</i>	clade	23.6
	<i>Smilisca baudinii</i>	Hylini	75.9
	<i>Smilisca phaeota</i>	Hylini	65.5
	<i>Smilisca puma</i>	Hylini	38.1
	<i>Tlalocohyla loquax</i>	Hylini	44.7
	<i>Cruziophyla calcarifer</i>	Phyllomedusinae	64.0
	<i>Agalychnis callidryas</i>	Phyllomedusinae	57.2
	<i>Agalychnis saltator</i>	Phyllomedusinae	46.7
	<i>Scinax boulengeri</i>	<i>Scinax</i> clade	48.7
	<i>Scinax elaeochrous</i>	<i>Scinax</i> clade	37.7
Costa Rica: Heredia: Volcan	<i>Isthmohyla</i>		
Barba	<i>angustilineata</i>	Hylini	34.2
	<i>Isthmohyla picadoi</i>	Hylini	32.8
	<i>Isthmohyla pictipes</i>	Hylini	39.0
	<i>Isthmohyla</i>		
	<i>pseudopuma</i>	Hylini	41.4
	<i>Isthmohyla rivularis</i>	Hylini	34.0

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Costa Rica: Las Canas: Finca	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
Taboga	<i>microcephalus</i>	clade	24.5
	<i>Smilisca baudinii</i>	Hylini	75.9
	<i>Smilisca sordida</i>	Hylini	44.6
	<i>Trachycephalus</i>		
	<i>venulosus</i>	Lophiohylini	100.5
	<i>Scinax boulengeri</i>	<i>Scinax</i> clade	48.7
	<i>Scinax staufferi</i>	<i>Scinax</i> clade	29.0
Costa Rica: Puntarenas: Las	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
Cruces	<i>ebraccatus</i>	clade	27.8
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>microcephalus</i>	clade	24.5
	<i>Anotheca spinosa</i>	Hylini	68.5
	<i>Duellmanohyla</i>		
	<i>rufiocularis</i>	Hylini	30.0
	<i>Ecnomiohyla miliaria</i>	Hylini	110.4
	<i>Isthmohyla lancasteri</i>	Hylini	33.6
	<i>Isthmohyla</i>		
	<i>pseudopuma</i>	Hylini	41.4
	<i>Ptychohyla legleri</i>	Hylini	36.7
	<i>Smilisca sordida</i>	Hylini	44.6

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	<i>Agalychnis annae</i>	Phyllomedusinae	73.9
Costa Rica: Puntarenas:			
Rincon de Osa	<i>Hypsiboas rosenbergi</i>	Cophomantini	90.0
	<i>Hypsiboas rufitelus</i>	Cophomantini	49.2
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>ebraccatus</i>	clade	27.8
	<i>Smilisca phaeota</i>	Hylini	65.5
	<i>Smilisca sila</i>	Hylini	44.8
	<i>Smilisca sordida</i>	Hylini	44.6
	<i>Trachycephalus</i>		
	<i>venulosus</i>	Lophiohylini	100.5
	<i>Agalychnis callidryas</i>	Phyllomedusinae	57.2
	<i>Agalychnis spurrelli</i>	Phyllomedusinae	92.8
	<i>Scinax boulengeri</i>	<i>Scinax</i> clade	48.7
	<i>Scinax elaeochrous</i>	<i>Scinax</i> clade	37.7
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
Guatemala: El Peten: Tikal	<i>ebraccatus</i>	clade	27.8
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>microcephalus</i>	clade	24.5
	<i>Tlalocohyla loquax</i>	Hylini	44.5
	<i>Tlalocohyla picta</i>	Hylini	21.4

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	<i>Smilisca baudinii</i>	Hylini	75.9
	<i>Tripurion petasatus</i>	Hylini	60.8
	<i>Agalychnis callidryas</i>	Phyllomedusinae	77.2
	<i>Scinax staufferi</i>	Scinax clade	29.0
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
Honduras: Atlantida: La Ceiba	<i>microcephalus</i>	clade	24.5
	<i>Smilisca baudinii</i>	Hylini	75.9
	<i>Tlalocohyla loquax</i>	Hylini	44.5
	<i>Tlalocohyla picta</i>	Hylini	21.4
	<i>Trachycephalus</i>		
	<i>venulosus</i>	Lophiohylini	100.5
	<i>Scinax staufferi</i>	Scinax clade	29.0
Honduras: Atlantida:	<i>Duellmanohyla</i>		
Quebrada de Oro	<i>salvavida</i>	Hylini	28.0
	<i>Plectrohyla</i>		
	<i>chrysopleura</i>	Hylini	65.6
	<i>Ptycholyla spinipollex</i>	Hylini	39.1
	<i>Smilisca baudinii</i>	Hylini	75.9
Honduras: Copan: Laguna de			
Cerro	<i>Duellmanohyla soralia</i>	Hylini	32.3
	<i>Plectrohyla matudai</i>	Hylini	46.0

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	<i>Ptychohyla</i>		
	<i>hypomykter</i>	Hylini	35.4
	<i>Smilisca baudinii</i>	Hylini	75.9
	<i>Tlalocohyla loquax</i>	Hylini	44.5
	<i>Tlalocohyla picta</i>	Hylini	21.4
	<i>Agalychnis callidryas</i>	Phyllomedusinae	57.2
	<i>Agalychnis moreletii</i>	Phyllomedusinae	65.7
Honduras: Copan: Quebrada Grande	<i>Bromeliohyla</i>		
	<i>bromeliacia</i>	Hylini	29.5
	<i>Duellmanohyla soralia</i>	Hylini	32.3
	<i>Ecnomiohyla salvaje</i>	Hylini	86.0
	<i>Plectrohyla</i>		
	<i>guatemalensis</i>	Hylini	59.5
	<i>Plectrohyla matudai</i>	Hylini	46.0
	<i>Ptychohyla</i>		
	<i>hypomykter</i>	Hylini	35.4
	<i>Smilisca baudinii</i>	Hylini	75.9
	<i>Agalychnis moreletii</i>	Phyllomedusinae	65.7
Honduras: Gracias de Dios: Barra Patuka	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>microcephalus</i>	clade	24.5
	<i>Smilisca baudinii</i>	Hylini	75.9

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	<i>Tlalocohyla loquax</i>	Hylini	44.5
	<i>Tlalocohyla picta</i>	Hylini	21.4
	<i>Trachycephalus</i>		
	<i>venulosus</i>	Lophiohylini	100.5
	<i>Agalychnis callidryas</i>	Phyllomedusinae	57.2
	<i>Scinax staufferi</i>	<i>Scinax</i> clade	29.0
Mexico: Chiapas: Rayon	<i>Charadrahyla</i>		
Mescalapan	<i>chaneque</i>	Hylini	60.7
	<i>Duellmanohyla</i>		
	<i>chamulae</i>	Hylini	30.5
	<i>Exerodonta bivocata</i>	Hylini	28.5
	<i>Plectrohyla acanthodes</i>	Hylini	63.2
	<i>Plectrohyla</i>		
	<i>guatemalensis</i>	Hylini	59.5
	<i>Plectrohyla ixil</i>	Hylini	41.6
Mexico: Distrito Federal: Lago			
Xochimilco	<i>Hyla eximia</i>	Hylini	34.9
Mexico: Durango: El Salto	<i>Hyla eximia</i>	Hylini	34.9
Mexico: Guerrero: Puerto del			
Gallo	<i>Charadrahyla trux</i>	Hylini	81.0
	<i>Exerodonta</i>	Hylini	29.9

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	<i>melanomma</i>		
	<i>Exerodonta pinorum</i>	Hylini	34.9
	<i>Plectrohyla hazelae</i>	Hylini	38.6
	<i>Plectrohyla mykter</i>	Hylini	46.0
	<i>Plectrohyla pentheter</i>	Hylini	52.1
	<i>Plectrohyla thorectes</i>	Hylini	34.2
	<i>Ptychohyla</i>		
	<i>leonardschultzei</i>	Hylini	35.1
Mexico: Hidalgo: El Chico	<i>Hyla eximia</i>	Hylini	34.9
	<i>Hyla plicata</i>	Hylini	44.0
	<i>Plectrohyla</i>		
	<i>robertsorum</i>	Hylini	47.9
	<i>Dendropsophus</i>		
Mexico: Jalisco: Chamela	<i>Dendropsophus sartori</i>	clade	26.0
	<i>Diaglena spatulata</i>	Hylini	85.9
	<i>Exerodonta</i>		
	<i>smaragdina</i>	Hylini	26.0
	<i>Smilisca baudinii</i>	Hylini	75.9
	<i>Smilisca fodiens</i>	Hylini	62.6
	<i>Tlalocohyla smithii</i>	Hylini	26.0
	<i>Trachycephalus</i>	Lophiohylini	100.5

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	<i>venulosus</i>		
	<i>Pachymedusa</i>		
	<i>dacnicolor</i>	Phyllomedusinae	82.6
Mexico: Michoachan: Nueva			
Italia	<i>Diaglena spatulata</i>	Hylini	85.9
	<i>Smilisca baudinii</i>	Hylini	75.9
	<i>Smilisca fodiens</i>	Hylini	62.6
	<i>Tlalocohyla smithii</i>	Hylini	26.0
	<i>Pachymedusa</i>		
	<i>dacnicolor</i>	Phyllomedusinae	82.6
Mexico: Oaxaca: Puerto		<i>Dendropsophus</i>	
Escondido	<i>Dendropsophus sartori</i>	clade	26.0
	<i>Diaglena spatulata</i>	Hylini	85.9
	<i>Tlalocohyla smithii</i>	Hylini	26.0
	<i>Trachycephalus</i>		
	<i>venulosus</i>	Lophiohylini	100.5
	<i>Pachymedusa</i>		
	<i>dacnicolor</i>	Phyllomedusinae	82.6
	<i>Scinax staufferi</i>	<i>Scinax</i> clade	29.0
Mexico: Oaxaca: San Gabriel	<i>Charadrahyla</i>		
Mixtepec, N	<i>altipotens</i>	Hylini	80.6

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	<i>Exerodonta juanita</i>	Hylini	35.8
	<i>Exerodonta</i>		
	<i>melanomma</i>	Hylini	29.9
	<i>Exerodonta</i>		
	<i>sumichrasti</i>	Hylini	27.7
	<i>Megastomatohyla</i>		
	<i>pellita</i>	Hylini	29.0
	<i>Plectrohyla pentheter</i>	Hylini	52.1
	<i>Plectrohyla thorectes</i>	Hylini	34.2
	<i>Ptychohyla</i>		
	<i>leonhardschultzii</i>	Hylini	35.1
Mexico: Oaxaca: Tehuantepec	<i>Smilisca baudinii</i>	Hylini	75.9
	<i>Tripurion petasatus</i>	Hylini	60.8
	<i>Pachymedusa</i>		
	<i>dacnicolor</i>	Phyllomedusinae	82.6
	<i>Scinax staufferi</i>	<i>Scinax</i> clade	29.0
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
Mexico: Oaxaca: Tuxtepec	<i>microcephalus</i>	clade	24.5
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>ebraccataus</i>	clade	27.8
	<i>Smilisca baudinii</i>	Hylini	75.9

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	<i>Tlalocohyla loquax</i>	Hylini	44.5
	<i>Tlalocohyla picta</i>	Hylini	21.4
	<i>Trachycephalus</i>		
	<i>venulosus</i>	Lophiohylini	100.5
	<i>Agalychnis callidryas</i>	Phyllomedusinae	57.2
	<i>Scinax staufferi</i>	<i>Scinax</i> clade	29.0
Mexico: Oaxaca: Vista			
Hermosa	<i>Anotheca spinosa</i>	Hylini	68.5
	<i>Bromeliohyla</i>		
	<i>dendroscarta</i>	Hylini	31.6
	<i>Charadrahyla nephila</i>	Hylini	70.9
	<i>Duellmanohyla</i>		
	<i>ignicolor</i>	Hylini	30.9
	<i>Ecnomiohyla echinata</i>	Hylini	57.0
	<i>Megastomatohyla</i>		
	<i>mixe</i>	Hylini	30.8
	<i>Plectrohyla</i>		
	<i>arborescandens</i>	Hylini	37.6
	<i>Ptychohyla acrochorda</i>	Hylini	36.3
	<i>Agalychnis moreleti</i>	Phyllomedusinae	65.7
Mexico: Puebla: 14.4 km W	<i>Ecnomiohyla</i>	Hylini	38.4

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Huachinango	<i>miotympanum</i>		
	<i>Hyla euphorbiaceae</i>	Hylini	39.6
	<i>Plectrohyla</i>		
	<i>arborescandens</i>	Hylini	37.6
	<i>Plectrohyla</i>		
	<i>charadricola</i>	Hylini	44.4
	<i>Pachymedusa</i>		
Mexico: Sinaloa: Mazatlan	<i>dacnicolor</i>	Phyllomedusinae	82.6
	<i>Diaglena spatulatua</i>	Hylini	85.9
	<i>Smilisca baudinii</i>	Hylini	75.9
	<i>Smilisca fodiens</i>	Hylini	62.6
	<i>Tlalocohyla smithii</i>	Hylini	26.0
Mexico: Sonora: Alamos	<i>Smilisca baudinii</i>	Hylini	75.9
	<i>Smilisca fodiens</i>	Hylini	62.6
	<i>Pachymedusa</i>		
	<i>dacnicolor</i>	Phyllomedusinae	82.6
Mexico: Veracruz: Acultzingo	<i>Ecnomiohyla</i>		
(1 km W)	<i>miotympanum</i>	Hylini	38.4
	<i>Hyla euphorbiaceae</i>	Hylini	39.6
	<i>Plectrohyla</i>		
	<i>arborescandens</i>	Hylini	37.6

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	<i>Plectrohyla bistincta</i>	Hylini	53.8
Mexico: Veracruz: Cuatlapan	<i>Anotheca spinosa</i>	Hylini	68.5
	<i>Bromeliohyla</i>		
	<i>dendroscarta</i>	Hylini	31.6
	<i>Ecnomiohyla</i>		
	<i>miotympanum</i>	Hylini	38.4
	<i>Hyla eximia</i>	Hylini	34.9
	<i>Smilisca baudini</i>	Hylini	75.9
	<i>Tlalocohyla picta</i>	Hylini	21.4
	<i>Trachycephalus</i>		
	<i>venulosus</i>	Lophiohylini	100.5
	<i>Agalychnis moreletii</i>	Phyllomedusinae	65.7
	<i>Scinax staufferi</i>	<i>Scinax</i> clade	29.0
Mexico: Veracruz: Estacion	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
Los Tuxtlas	<i>ebraccatus</i>	clade	27.8
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>microcephlus</i>	clade	24.5
	<i>Ecnomiohyla</i>		
	<i>valancifer</i>	Hylini	77.7
	<i>Smilisca baudini</i>	Hylini	75.9
	<i>Smilisca cyanosticta</i>	Hylini	57.8

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	<i>Tlalocohyla loquax</i>	Hylini	44.5
	<i>Tlalocohyla picta</i>	Hylini	21.4
	<i>Agalychnis callidryas</i>	Phyllomedusinae	57.2
Mexico: Veracruz: Huatusco (3k SW)	<i>Bromeliohyla</i>		
	<i>dendroscarta</i>	Hylini	31.6
	<i>Charadrahyla</i>		
	<i>taeniopus</i>	Hylini	65.9
	<i>Ecnomiohyla</i>		
	<i>miotympanum</i>	Hylini	38.4
	<i>Megastomatohyla</i>		
	<i>mixomaculata</i>	Hylini	29.1
	<i>Megastomatohyla</i>		
	<i>nubicola</i>	Hylini	36.7
	<i>Plectrohyla</i>		
	<i>arborescandens</i>	Hylini	37.6
	<i>Smilisca baudinii</i>	Hylini	75.9
Mexico: Veracruz: Mata de Oscura	<i>Smilisca baudini</i>	Hylini	75.9
	<i>Tlalocohyla godmani</i>	Hylini	38.0
	<i>Tlalocohyla picta</i>	Hylini	21.4
	<i>Scinax staufferi</i>	Scinax clade	29.0

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 Mexico: Veracruz: Volcan San

Martin	<i>Anothea spinosa</i>	Hylini	65.8
	<i>Charadrahyla nephila</i>	Hylini	70.9
	<i>Ecnomiohyla</i>		
	<i>miotympanum</i>	Hylini	38.4
	<i>Ecnomiohyla</i>		
	<i>valancifer</i>	Hylini	77.7
	<i>Smilisca cyanosticata</i>	Hylini	57.8
	<i>Agalychnis moreleti</i>	Phyllomedusinae	65.7

*Dendropsophus*      *Dendropsophus*

Mexico: Yucatan: Piste	<i>microcephalus</i>	clade	24.5
	<i>Smilisca baudinii</i>	Hylini	75.9
	<i>Tlalocohyla loquax</i>	Hylini	44.5
	<i>Tlalocohyla picta</i>	Hylini	21.4
	<i>Tripriion petasatus</i>	Hylini	60.8
	<i>Trachycephalus</i>		
	<i>venulosus</i>	Lophiohylini	100.5
	<i>Agalychnis callidryas</i>	Phyllomedusinae	57.2

Panama: BCI	<i>Hypsiboas rufitela</i>	Cophomantini	49.2
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>microcephalus</i>	clade	24.5

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	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>phlebodes</i>	clade	23.6
	<i>Smilisca phaeota</i>	Hylini	65.5
	<i>Smilisca sila</i>	Hylini	44.8
	<i>Trachycephalus</i>		
	<i>venulosus</i>	Lophiohylini	100.5
	<i>Cruzirohyla calcarifer</i>	Phyllomedusinae	64.0
	<i>Agalychnis callidryas</i>	Phyllomedusinae	57.2
	<i>Agalychnis spurrelli</i>	Phyllomedusinae	75.6
	<i>Scinax boulengeri</i>	<i>Scinax</i> clade	48.7
Panama: Cocle: El Valle	<i>Hypsiboas crepitans</i>	Cophomantini	63.0
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>ebraccatus</i>	clade	27.8
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>microcephalus</i>	clade	24.5
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>phlebodes</i>	clade	23.6
	<i>Anotheca spinosa</i>	Hylini	68.5
	<i>Ecnomiohyla miliaria</i>	Hylini	110.4
	<i>Smilisca sila</i>	Hylini	44.8
	<i>Hylomantis lemur</i>	Phyllomedusinae	40.8

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	<i>Scinax altae</i>	<i>Scinax</i> clade	26.0
Panama: Colon: Achiote	<i>Hypsiboas rufitelus</i>	Cophomantini	49.2
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>ebraccatus</i>	clade	27.8
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>phlebodes</i>	clade	23.6
	<i>Smilisca phaeota</i>	Hylini	65.5
	<i>Agalychnis callidryas</i>	Phyllomedusinae	57.2
	<i>Scinax boulengeri</i>	<i>Scinax</i> clade	48.7
	<i>Scinax rubra</i>	<i>Scinax</i> clade	41.2
Panama: Darien: Rio Tuirá at Rio Mono	<i>Hypsiboas boans</i>	Cophomantini	132.0
	<i>Hypsiboas rosenbergi</i>	Cophomantini	90.0
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>ebraccatus</i>	clade	27.8
	<i>Smilisca phaeota</i>	Hylini	65.5
	<i>Agalychnis callidryas</i>	Phyllomedusinae	57.2
	<i>Agalychnis litodryas</i>	Phyllomedusinae	70.2
	<i>Phyllomedusa venusta</i>	Phyllomedusinae	86.3
California: Contra Costa Co.; 1 mi NW Alamo	<i>Pseudacris regilla</i>	Hylini	48.0

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 California: San Diego Co.:

Spring, Pine Mountain	<i>Pseudacris cadaverina</i>	Hylini	48.0
	<i>Pseudacris regilla</i>	Hylini	35.9

## Colorado: Commanche

National Grassland	<i>Pseudacris triseriata</i>	Hylini	32.0
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## Florida: Everglades National

Park	<i>Hyla cinerea</i>	Hylini	59.0
	<i>Hyla squirella</i>	Hylini	36.0
	<i>Pseudacris nigrita</i>	Hylini	28.0
	<i>Pseudacris ocularis</i>	Hylini	15.5

## Florida: Timucuan Ecological

## and Historic Preserve

	<i>Hyla cinerea</i>	Hylini	59.0
	<i>Hyla femoralis</i>	Hylini	37.0
	<i>Hyla gratiosa</i>	Hylini	68.0
	<i>Hyla squirella</i>	Hylini	36.0
	<i>Acris gryllus</i>	Hylini	29.0
	<i>Pseudacris crucifer</i>	Hylini	29.0
	<i>Pseudacris ocularis</i>	Hylini	15.5

## Georgia: Okefenokee Swamp

	<i>Hyla chrysoscelis</i>	Hylini	51.0
	<i>Hyla cinerea</i>	Hylini	59.0
	<i>Hyla femoralis</i>	Hylini	37.0

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	<i>Hyla gratiosa</i>	Hylini	68.0
	<i>Hyla squirella</i>	Hylini	36.0
	<i>Acris gryllus</i>	Hylini	29.0
	<i>Pseudacris crucifer</i>	Hylini	29.0
	<i>Pseudacris nigrita</i>	Hylini	28.0
	<i>Pseudacris ocularis</i>	Hylini	15.5
	<i>Pseudacris ornata</i>	Hylini	35.0
Georgia: Okmulgee National			
Monument	<i>Acris crepitans</i>	Hylini	29.0
	<i>Acris gryllus</i>	Hylini	29.0
	<i>Pseudacris crucifer</i>	Hylini	29.0
	<i>Pseudacris nigrita</i>	Hylini	28.0
	<i>Pseudacris ornata</i>	Hylini	35.0
	<i>Hyla avivoca</i>	Hylini	39.0
	<i>Hyla chrysocelis</i>	Hylini	51.0
	<i>Hyla cinerea</i>	Hylini	59.0
	<i>Hyla squirella</i>	Hylini	36.0
North Carolina: Moores Creek			
National Battlefield	<i>Hyla chrysocelis</i>	Hylini	51.0
	<i>Hyla cinerea</i>	Hylini	59.0
	<i>Hyla femoralis</i>	Hylini	37.0

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	<i>Hyla gratiosa</i>	Hylini	68.0
	<i>Hyla squirella</i>	Hylini	36.0
	<i>Acris gryllus</i>	Hylini	29.0
	<i>Pseudacris crucifer</i>	Hylini	29.0
	<i>Pseudacris feriarum</i>	Hylini	30.0
	<i>Pseudacris ocularis</i>	Hylini	15.5
	<i>Pseudacris ornata</i>	Hylini	35.0
North Dakota: Badlands			
National Park	<i>Pseudacris triseriata</i>	Hylini	32.0
Ohio: Cuyahoga Valley			
National Park	<i>Hyla versicolor</i>	Hylini	51.0
	<i>Pseudacris crucifer</i>	Hylini	29.0
	<i>Pseudacris triseriata</i>	Hylini	32.0
South Carolina: Congaree			
Swamp National Monument	<i>Hyla chrysocelis</i>	Hylini	51.0
	<i>Hyla cinerea</i>	Hylini	59.0
	<i>Hyla femoralis</i>	Hylini	37.0
	<i>Hyla gratiosa</i>	Hylini	68.0
	<i>Hyla squirella</i>	Hylini	36.0
	<i>Acris crepitans</i>	Hylini	29.0
	<i>Acris gryllus</i>	Hylini	29.0

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	<i>Pseudacris brimleyi</i>	Hylini	28.0
	<i>Pseudacris crucifer</i>	Hylini	29.0
	<i>Pseudacris feriarum</i>	Hylini	30.0
	<i>Pseudacris nigrita</i>	Hylini	28.0
	<i>Pseudacris ornata</i>	Hylini	35.0
South Carolina: Savannah			
River Ecology Laboratory	<i>Hyla avivoca</i>	Hylini	39.0
	<i>Hyla chrysocelis</i>	Hylini	52.0
	<i>Hyla cinerea</i>	Hylini	59.0
	<i>Hyla femoralis</i>	Hylini	37.0
	<i>Hyla gratiosa</i>	Hylini	68.0
	<i>Hyla squirella</i>	Hylini	36.0
	<i>Acris crepitans</i>	Hylini	29.0
	<i>Acris gryllus</i>	Hylini	29.0
	<i>Pseudacris crucifer</i>	Hylini	29.0
	<i>Pseudacris nigrita</i>	Hylini	28.0
	<i>Pseudacris ocularis</i>	Hylini	15.5
	<i>Pseudacris ornata</i>	Hylini	35.0
South Dakota: Sand Lake			
National Wildlife Refuge	<i>Acris crepitans</i>	Hylini	29.0
	<i>Hyla chrysocelis</i>	Hylini	51.0

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	<i>Pseudacris triseriata</i>	Hylini	32.0
Tennessee: Great Smoky Mountains National Park	<i>Acris crepitans</i>	Hylini	29.0
	<i>Pseudacris crucifer</i>	Hylini	29.0
	<i>Pseudacris feriarum</i>	Hylini	30.0
	<i>Hyla chrysocelis</i>	Hylini	51.0
Texas: Padre Island National Seashore	<i>Hyla cinerea</i>	Hylini	59.0
	<i>Pseudacris clarkii</i>	Hylini	29.0
Utah: Zion National Park	<i>Hyla arenicolor</i>	Hylini	52.0
Virginia: Shenandoah National Park	<i>Acris crepitans</i>	Hylini	29.0
	<i>Hyla versicolor</i>	Hylini	51.0
	<i>Pseudacris crucifer</i>	Hylini	29.0
	<i>Pseudacris feriarum</i>	Hylini	30.0
Washington: Pierce Co. Parkland, Wake Lake	<i>Pseudacris regilla</i>	Hylini	48.0
Wyoming: Yellowstone National Park	<i>Pseudacris triseriata</i>	Hylini	32.0
Brazil: Bahia: Estacion VeraCruz	<i>Aplastodiscus gr.</i> <i>albosignata</i>	Cophomantini	

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<i>Hypsiboas</i>		
<i>albomarginatus</i>	Cophomantini	55.0
<i>Hypsiboas crepitans</i>	Cophomantini	63.0
<i>Hypsiboas faber</i>	Cophomantini	104.0
<i>Hypsiboas</i>		
<i>semilineatus</i>	Cophomantini	40.1
<i>Dendropsophus</i>		
<i>Dendropsophus anceps</i>	clade	40.0
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>bipunctatus</i>	clade	25.0
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>decipiens</i>	clade	20.0
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>elegans</i>	clade	29.6
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>microps</i>	clade	26.0
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>minutus</i>	clade	23.0
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>seniculus</i>	clade	37.7
<i>Aparasphenodon</i>	Lophiohylini	

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<i>brunoi</i>		
<i>Itapotihyla langsdorffii</i>	Lophiohylini	77.0
<i>Phyllodytes luteolus</i>	Lophiohylini	23.0
<i>Phyllodytes</i>		
<i>melanomystax</i>	Lophiohylini	25.4
<i>Trachycephalus</i>		
<i>mesopheus</i>	Lophiohylini	100.5
<i>Hylomantis aspera</i>	Phyllomedusinae	41.7
<i>Phyllomedusa</i>		
<i>burmeisteri</i>	Phyllomedusinae	79.0
<i>Scinax alter</i>	<i>Scinax</i> clade	30.0
<i>Scinax argyreornata</i>	<i>Scinax</i> clade	15.8
<i>Scinax cuspidatus</i>	<i>Scinax</i> clade	29.0
<i>Scinax euridyce</i>	<i>Scinax</i> clade	42.0
<i>Scinax catharinae</i>		
<i>group</i>	<i>Scinax</i> clade	
<i>Scinax ruber group</i>	<i>Scinax</i> clade	
<i>Sphaenorhynchus</i>		
<i>palustris</i>	<i>Scinax</i> clade	36.0
Undescribed species		
Undescribed species		

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Brazil: Bahia: Zumbidos	<i>Aplastodiscus</i> gr.		
Palmares	<i>albosignata</i>	Cophomantini	
	<i>Bokermannohyla</i> gr.		
	<i>circumdata</i>	Cophomantini	
	<i>Hypsiboas</i>		
	<i>albomarginatus</i>	Cophomantini	55.0
	<i>Hypsiboas atlantica</i>	Cophomantini	40.2
	<i>Hypsiboas crepitans</i>	Cophomantini	63.0
	<i>Hypsiboas faber</i>	Cophomantini	104.0
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>branneri</i>	clade	19.0
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>elegans</i>	clade	23.6
	<i>Dendropsophus</i> gr.	<i>Dendropsophus</i>	
	<i>marmoratus</i>	clade	
	<i>Dendropsophus</i>	<i>Dendropsophus</i>	
	<i>minutus</i>	clade	23.0
	<i>Phyllodytes luteolus</i>	Lophiohylini	23.0
	<i>Phyllodytes</i>		
	<i>melanomystax</i>	Lophiohylini	25.4
	<i>Trachycephalus</i>	Lophiohylini	100.5

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	<i>mesopheus</i>		
	<i>Hylomantis aspera</i>	Phyllomedusinae	41.7
	<i>Phyllomedusa</i>		
	<i>burmeisteri</i>	Phyllomedusinae	79.0
	<i>Scinax cuspidatus</i>	<i>Scinax</i> clade	29.0
	<i>Scinax euridyce</i>	<i>Scinax</i> clade	42.0
	<i>Scinax gr. ruber</i>	<i>Scinax</i> clade	
	Undescribed species		
	<i>Undescribed species</i>		
	<i>Aplastodiscus</i>		
Brazil: Boriacea	<i>albofrenatus</i>	Cophomantini	40.0
	<i>Aplastodiscus</i>		
	<i>leucopygius</i>	Cophomantini	45.1
	<i>Aplastodiscus</i>		
	<i>albosignatus</i>	Cophomantini	52.0
	<i>Bokermannohyla</i>		
	<i>astarteia</i>	Cophomantini	41.5
	<i>Bokermannohyla</i>		
	<i>circumdata</i>	Cophomantini	70.0
	<i>Bokermannohyla hylax</i>	Cophomantini	61.5
	<i>Hypsiboas</i>	Cophomantini	60.0

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<i>albopunctatus</i>		
<i>Hypsiboas bischoffi</i>		
( <i>multilineata</i> )	Cophomantini	46.1
<i>Hypsiboas faber</i>	Cophomantini	104.0
<i>Hypsiboas pardalis</i>	Cophomantini	69.0
<i>Hypsiboas polytaenius</i>	Cophomantini	31.4
<i>Hypsiboas prasinus</i>	Cophomantini	55.0
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>microps</i>	clade	26.0
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>minuta</i>	clade	23.0
<i>Dendropsophus</i>	<i>Dendropsophus</i>	
<i>senicula</i>	clade	37.7
<i>Itapotihyla langsdorfii</i>	Lophiohylini	77.0
<i>Phrynomedusa</i>		
<i>appendiculata</i>	Phyllomedusinae	37.4
<i>Phasmahyla cochranae</i>	Phyllomedusinae	33.9
<i>Scinax brieni</i>	<i>Scinax</i> clade	32.7
<i>Scinax crospedospilus</i>	<i>Scinax</i> clade	33.3
<i>Scinax flavoguttatus</i>	<i>Scinax</i> clade	29.3
<i>Scinax hayii</i>	<i>Scinax</i> clade	43.0

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	<i>Scinax obtriangulatus</i>	<i>Scinax</i> clade	28.0
	<i>Scinax perpusillus</i>	<i>Scinax</i> clade	18.3
	<i>Scinax x-signatus</i>	<i>Scinax</i> clade	42.5
	<i>Sphaenorhynchus</i>		
	<i>orophilus</i>	<i>Scinax</i> clade	32.0
Dominican Republic: Samana:	<i>Osteopilus</i>		
Laguna	<i>dominicensis</i>	Lophiohylini	66.0
	<i>Osteopilus</i>		
	<i>pulchrilineatus</i>	Lophiohylini	39.5
	<i>Osteopilus vastus</i>	Lophiohylini	108.8
Dominican Republic: Samana:	<i>Osteopilus</i>		
Rio San Juan	<i>dominicensis</i>	Lophiohylini	66.0
	<i>Osteopilus</i>		
	<i>pulchrilineatus</i>	Lophiohylini	39.5
	<i>Osteopilus vastus</i>	Lophiohylini	108.8
Haiti: Ouest: Furcy	<i>Hypsiboas heilprini</i>	Cophomantini	66.0
	<i>Osteopilus</i>		
	<i>dominicensis</i>	Lophiohylini	39.5
	<i>Osteopilus vastus</i>	Lophiohylini	108.8
Jamaica, Manchester Parish,			
Mandeville	<i>Osteopilus brunneus</i>	Lophiohylini	52.0

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	<i>Osteopilus crucialis</i>	Lophiohylini	100.0
	<i>Osteopilus wilderi</i>	Lophiohylini	27.3
Jamaica, Quick Step	<i>Osteopilus brunneus</i>	Lophiohylini	52.0
	<i>Osteopilus marianae</i>	Lophiohylini	40.0
	<i>Osteopilus wilderi</i>	Lophiohylini	27.3
Jamaica: Trelany Parish:			
Windsor	<i>Osteopilus brunneus</i>	Lophiohylini	52.0
	<i>Osteopilus marianae</i>	Lophiohylini	40.0
	<i>Osteopilus wilderi</i>	Lophiohylini	27.3

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836 Table S4. Average values of annual mean temperature (° C) and annual  
 837 precipitation (mm) for 337 hylid species. Sample size indicates the number of  
 838 georeferenced museum and literature localities from which climatic data were  
 839 obtained.

Species	Sample size	Annual mean temp.	Annual precipitation n
<b>Hylinae</b>			
<i>Acris crepitans</i>	59	15.8	1006.5
<i>Acris gryllus</i>	19	18.8	1470.7
<i>Anotheca spinosa</i>	15	21.4	2314.9
<i>Aparasphenodon brunoi</i>	3	23.8	1378.7
<i>Aplastodiscus albofrenatus</i>	4	20.8	2132.5
<i>Aplastodiscus leucopygius</i>	2	17.7	2214.5
<i>Aplastodiscus albosignatus</i>	5	18.2	2183
<i>Aplastodiscus arildae</i>	2	17.8	1848.5
<i>Aplastodiscus cochranæ</i>	1	16.1	1762
<i>Aplastodiscus perviridis</i>	3	17.1	1777.7
<i>Argenteohyla siemersi</i>	2	17.2	1208.5
<i>Bokermannohyla astartea</i>	1	16.9	2429
<i>Bokermannohyla circumdata</i>	6	18.8	2153.3

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<i>Bokermannohyla hylax</i>	3	18.8	2128.7
<i>Bromeliohyla bromeliacia</i>	7	19.8	2307
<i>Charadrahyla nephila</i>	2	20.3	3248
<i>Charadrahyla taeniopus</i>	9	16.9	1529.6
<i>Corythomantis greeningi</i>	1	26.1	1017
<i>Dendropsophus allenorum</i>	1	25.6	2479
<i>Dendropsophus anceps</i>	4	23.5	1514.3
<i>Dendropsophus aperomeus</i>	3	18.1	1230
<i>Dendropsophus berthaltutzae</i>	3	21.2	2458.7
<i>Dendropsophus bifurcus</i>	14	23.5	3561.1
<i>Dendropsophus bipunctatus</i>	3	23.4	1433.7
<i>Dendropsophus brevifrons</i>	6	24.8	2796
<i>Dendropsophus carnifex</i>	5	17.3	2264.5
<i>Dendropsophus ebraccatus</i>	10	23.9	3147.9
<i>Dendropsophus elegans</i>	6	23.4	1738.2
<i>Dendropsophus giesleri</i>	2	22.4	1564
<i>Dendropsophus koechlini</i>	3	25.3	2829.3
<i>Dendropsophus labialis</i>	1	15.3	1556
<i>Dendropsophus leali</i>	2	25.4	2755
<i>Dendropsophus leucophyllatus</i>	6	25.6	2844.2
<i>Dendropsophus marmoratus</i>	4	25.5	2913.3

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<i>Dendropsophus microcephalus</i>	13	25.3	2210.7
<i>Dendropsophus minusculus</i>	2	26.2	1496.5
<i>Dendropsophus minutus</i>	11	23.1	2230.2
<i>Dendropsophus miyatai</i>	2	26.2	3197.5
<i>Dendropsophus nanus</i>	9	24.6	1007.9
<i>Dendropsophus parviceps</i>	5	25.4	2936.8
<i>Dendropsophus pelidna</i>	2	11.3	1392.5
<i>Dendropsophus rhodopepla</i>	5	25.4	2867.4
<i>Dendropsophus riveroi</i>	4	25.3	2964.5
<i>Dendropsophus robertmertensi</i>	26	26.7	1956.8
<i>Dendropsophus rubicundulus</i>	7	21.5	1643.1
<i>Dendropsophus sanborni</i>	6	20.7	1495
<i>Dendropsophus sarayacuensis</i>	15	24.5	3137.3
<i>Dendropsophus sartori</i>	14	27.7	1149.4
<i>Dendropsophus schubarti</i>	2	26.3	2468.5
<i>Dendropsophus seniculus</i>	5	22.9	1657.8
<i>Dendropsophus triangulum</i>	15	25.4	3224.9
<i>Dendropsophus walfordi</i>	7	26.3	2460
<i>Diaglena spatulata</i>	15	25.7	862.33
<i>Duellmanohyla ruficolis</i>	2	21.2	3106.5
<i>Duellmanohyla soralia</i>	5	20.7	1700.8

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<i>Duellmanohyla uranochroa</i>	4	19.6	2793.3
<i>Ecnomiohyla miliaria</i>	5	22.9	3118.4
<i>Ecnomiohyla minera</i>	3	19.9	1949.7
<i>Ecnomiohyla miotympanum</i>	56	19.7	1585.6
<i>Exerodonta abdivita</i>	1	24.4	2894
<i>Exerodonta chimalapa</i>	2	21.8	1639.5
<i>Exerodonta melanomma</i>	7	22.5	1096.9
<i>Exerodonta perkinsi</i>	1	21.8	4292
<i>Exerodonta smaragdina</i>	14	24.0	1082.8
<i>Exerodonta sumichrasti</i>	11	21.5	1381.5
<i>Exerodonta xera</i>	1	18.9	373
<i>Hyla andersoni</i>	7	15.2	1428.4
<i>Hyla annectans</i>	13	15.7	1300.6
<i>Hyla arborea</i>	29	10.2	830.48
<i>Hyla arenicolor</i>	95	16.8	763.5
<i>Hyla avivoca</i>	15	16.2	1445.9
<i>Hyla chinensis</i>	9	17.7	1683.9
<i>Hyla chrysocelis</i>	130	13.3	1070.1
<i>Hyla cinerea</i>	35	18.2	1348.2
<i>Hyla euphorbiacea</i>	14	15.6	957.57
<i>Hyla eximia</i>	81	14.7	980.83

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<i>Hyla femoralis</i>	25	19.6	1424.4
<i>Hyla gratiosa</i>	19	17.8	1423.5
<i>Hyla japonica</i>	23	12.3	1632.7
<i>Hyla meridionalis</i>	12	14.6	669
<i>Hyla plicata</i>	30	12.3	1078.2
<i>Hyla savignyi</i>	13	18.4	543.46
<i>Hyla squirella</i>	32	19.3	1439
<i>Hyla versicolor</i>	27	12.5	1167.3
<i>Hyla walkeri</i>	7	13.8	1246.4
<i>Hyla wrightorum</i>	10	10.6	637
<i>Hyloscirtus armatus</i>	3	21.6	1771
<i>Hyloscirtus colymba</i>	2	21.8	2822.5
<i>Hyloscirtus lascinius</i>	1	17.2	1487
<i>Hyloscirtus lindae</i>	1	14.2	1837
<i>Hyloscirtus pacha</i>	1	16.0	2092
<i>Hyloscirtus palmeri</i>	1	23.7	806
<i>Hyloscirtus pantostictus</i>	1	11.5	1251
<i>Hyloscirtus phyllognathus</i>	10	18.9	2443.8
<i>Hyloscirtus simmonsii</i>	1	13.5	2221
<i>Hyloscirtus tapichalaca</i>	1	16.7	1487
<i>Hypsiboas albomarginatus</i>	10	21.3	1564.5

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<i>Hypsiboas albopunctatus</i>	17	21.6	1802.9
<i>Hypsiboas andinus</i>	5	13.3	871.2
<i>Hypsiboas balzani</i>	1	18.4	1814
<i>Hypsiboas benitezi</i>	4	20.8	2885.8
<i>Hypsiboas bischoffi</i>	9	18.1	2002.9
<i>Hypsiboas boans</i>	9	24.7	2568
<i>Hypsiboas caingua</i>	1	21.6	1862
<i>Hypsiboas calcaratus</i>	17	25.0	3009.8
<i>Hypsiboas cordobae</i>	1	14.9	729
<i>Hypsiboas crepitans</i>	18	23.6	1647.7
<i>Hypsiboas ericae</i>	1	21.1	1952
<i>Hypsiboas faber</i>	11	20.9	1882.2
<i>Hypsiboas fasciatus</i>	12	25.3	2850.2
<i>Hypsiboas geographicus</i>	19	24.5	2428.7
<i>Hypsiboas granosa</i>	17	25.5	2832.8
<i>Hypsiboas guentheri</i>	3	19.0	1704.7
<i>Hypsiboas heilprini</i>	8	22.2	1533.4
<i>Hypsiboas joaquini</i>	2	14.8	1755.5
<i>Hypsiboas lanciformis</i>	14	24.4	2856.5
<i>Hypsiboas latistriatus</i>	1	11.2	1959
<i>Hypsiboas lemai</i>	2	24.7	1942.5

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<i>Hypsiboas leptolineatus</i>	1	14.7	1888
<i>Hypsiboas lundii</i>	2	20.9	1855.5
<i>Hypsiboas marginatus</i>	1	14.7	1888
<i>Hypsiboas marianitae</i>	1	22.3	1177
<i>Hypsiboas microderma</i>	1	25.5	3641
<i>Hypsiboas multifasciatus</i>	3	25.2	2027.7
<i>Hypsiboas pardalis</i>	6	19.2	1978.8
<i>Hypsiboas pellucens</i>	8	24.3	3306.8
<i>Hypsiboas picturatus</i>	6	23.4	2883.2
<i>Hypsiboas polytaenia</i>	7	18.3	1868.1
<i>Hypsiboas prasinus</i>	4	17.8	2165.5
<i>Hypsiboas pulchellus</i>	4	16.9	1238
<i>Hypsiboas punctatus</i>	14	25.0	2317.3
<i>Hypsiboas raniceps</i>	9	24.8	1618.9
<i>Hypsiboas riojanus</i>	3	17.2	323
<i>Hypsiboas roraima</i>	1	20.4	2261
<i>Hypsiboas rosenbergi</i>	12	24.1	3141.5
<i>Hypsiboas rufitelus</i>	5	25.9	3630.4
<i>Hypsiboas semiguttatus</i>	2	17.0	1695.5
<i>Hypsiboas semilineatus</i>	1	24.0	1540
<i>Hypsiboas sibleszi</i>	2	20.7	2158.5

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<i>Isthmohyla pseudopuma</i>	8	17.8	3030.9
<i>Isthmohyla rivularis</i>	6	16.2	2931.8
<i>Isthmohyla tica</i>	6	19.2	2841.5
<i>Isthmohyla zeteki</i>	6	17.8	2779.7
<i>Itapotihyla langsdorffii</i>	3	20.2	2141.7
<i>Lysapsus caraya</i>	2	27.6	2132
<i>Lysapsus laevis</i>	1	27.0	1568
<i>Lysapsus limellus</i>	7	23.8	1783
<i>Lysapsus limellus bolivianus</i>	1	26.6	2983
<i>Megastomatohyla mixe</i>	1	20.6	3544
<i>Myersiophyla inparquesi</i>	1	12.4	3126
<i>Myersiophyla kanaima</i>	1	20.0	1619
<i>Nyctimantis rugiceps</i>	4	24.1	4400.8
<i>Osteocephalus alboguttatus</i>	4	24.2	3785
<i>Osteocephalus buckleyi</i>	11	25.3	3263.7
<i>Osteocephalus cabrerai</i>	1	26.2	3622
<i>Osteocephalus leprieurii</i>	7	24.4	2619
<i>Osteocephalus mutabor</i>	3	24.9	3620
<i>Osteocephalus oophagus</i>	2	26.5	3195
<i>Osteocephalus planiceps</i>	7	25.6	3474.3
<i>Osteocephalus taurinus</i>	8	26.0	2690.4

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<i>Osteocephalus verruciger</i>	7	19.6	2553.7
<i>Osteopilus brunneus</i>	3	23.0	1654.3
<i>Osteopilus crucialis</i>	1	22.0	1590
<i>Osteopilus dominicensis</i>	7	24.2	1433.3
<i>Osteopilus marianae</i>	2	23.6	1686.5
<i>Osteopilus pulchrilineata</i>	2	24.8	2083.5
<i>Osteopilus septentrionalis</i>	12	25.2	1322.8
<i>Osteopilus vastus</i>	3	23.2	1849
<i>Osteopilus wilderi</i>	3	23.0	1654.3
<i>Phyllodytes auratus</i>	1	21.5	2337
<i>Phyllodytes luteolus</i>	1	24.7	1151
<i>Plectrohyla ameibothalame</i>	2	14.5	792
<i>Plectrohyla arborescandens</i>	4	17.0	1850.8
<i>Plectrohyla bistincta</i>	10	16.9	1342.6
<i>Plectrohyla calthula</i>	1	15.7	2334
<i>Plectrohyla chrysopleura</i>	1	18.9	1735
<i>Plectrohyla cyclada</i>	4	16.7	1385.8
<i>Plectrohyla glandulosa</i>	10	14.5	3125.5
<i>Plectrohyla guatemalensis</i>	22	17.3	1960.4
<i>Plectrohyla matudi</i>	4	17.3	2078.5
<i>Plectrohyla pentheter</i>	3	18.2	1313.7

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<i>Plectrohyla siopela</i>	1	13.1	440
<i>Podonectes cardosoi</i>	1	14.7	1888
<i>Podonectes minutus</i>	12	18.2	1372.8
<i>Pseudacris brachyphona</i>	16	10.7	1201.7
<i>Pseudacris brimleyi</i>	26	14.7	1152
<i>Pseudacris cadaverina</i>	11	15.6	400.64
<i>Pseudacris clarkii</i>	32	16.1	771.72
<i>Pseudacris crucifer</i>	35	11.1	1227.6
<i>Pseudacris feriarum</i>	57	13.7	1104
<i>Pseudacris foquettei</i>	15	16.3	1403
<i>Pseudacris illinoensis</i>	4	13.2	1090.8
<i>Pseudacris kalmi</i>	6	13.4	1072.2
<i>Pseudacris maculata</i>	564	4.8	532.57
<i>Pseudacris nigrita</i>	16	19.4	1440.1
<i>Pseudacris ocularis</i>	14	21.1	1299.9
<i>Pseudacris ornata</i>	10	18.6	1277.7
<i>Pseudacris regilla</i>	88	12.0	588.22
<i>Pseudacris streckeri</i>	20	16.3	832.85
<i>Pseudacris triseriata</i>	5	8.9	936.2
<i>Pseudis bolbodactyla</i>	3	23.9	1505
<i>Pseudis fusca</i>	1	22.6	1088

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<i>Pseudis paradoxa</i>	10	25.1	1488.8
<i>Pseudis paradoxa platensis</i>	1	21.5	1336
<i>Pseudis tocantins</i>	1	19.6	1740
<i>Ptychohyala dendrophasma</i>	1	19.6	2829
<i>Ptychohyala euthysanota</i>	8	20.0	2770.4
<i>Ptychohyala hypomycter</i>	9	18.3	1534.4
<i>Ptychohyala leonhardschultzei</i>	3	20.5	1543.7
<i>Ptychohyala spinipollex</i>	6	19.8	1721
<i>Ptychohyala zophodes</i>	1	21.2	1566
<i>Scarthyla goinorum</i>	2	25.4	2755
<i>Scinax acuminatus</i>	2	21.8	1568.5
<i>Scinax berthae</i>	2	19.2	1455.5
<i>Scinax boesemani</i>	10	26.0	2633.6
<i>Scinax boulengeri</i>	23	25.9	3169.3
<i>Scinax catharinae</i>	4	18.1	1671.3
<i>Scinax chiquitanus</i>	2	25.3	2811
<i>Scinax crospedospilus</i>	3	17.1	2260
<i>Scinax cruentommus</i>	10	24.7	2830.5
<i>Scinax duartei</i>	3	15.5	1929.3
<i>Scinax elaeochrous</i>	26	24.9	3436.4
<i>Scinax funereus</i>	4	25.3	3231

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<i>Scinax fuscovarius</i>	6	20.0	1547.7
<i>Scinax garbei</i>	4	25.5	2926.5
<i>Scinax hayii</i>	14	19.3	1860.4
<i>Scinax ictericus</i>	2	25.3	2760.5
<i>Scinax nasicus</i>	2	20.0	1448.5
<i>Scinax obtriangulatus</i>	3	17.2	2066
<i>Scinax oreites</i>	3	14.5	1262
<i>Scinax pedromedinae</i>	3	25.4	2929.7
<i>Scinax quinquefasciatus</i>	7	24.8	2485
<i>Scinax ruber</i>	7	26.0	2647.4
<i>Scinax similis</i>	3	22.9	1451.3
<i>Scinax squalirostris</i>	15	17.9	1295.7
<i>Scinax staufferi</i>	10	25.0	1742.1
<i>Scinax sugillatus</i>	4	24.7	3317.8
<i>Scinax uruguayus</i>	6	17.6	1349
<i>Smilisca baudinii</i>	21	23.1	1972.3
<i>Smilisca cyanosticta</i>	10	23.3	2728.6
<i>Smilisca fodiens</i>	47	22.3	864.09
<i>Smilisca phaeota</i>	12	24.4	2975.3
<i>Smilisca puma</i>	11	25.5	3851.3
<i>Smilisca sila</i>	5	24.9	2948.8

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<i>Smilisca sordida</i>	53	24.2	3084.7
<i>Sphaenorhynchus dorisae</i>	5	26.0	2905.8
<i>Sphaenorhynchus lacteus</i>	6	26.0	2511.2
<i>Sphaenorhynchus orophilus</i>	3	18.4	2079.7
<i>Tepuihyla edelcae</i>	5	14.7	2969.6
<i>Tlalocohyla godmani</i>	3	22.3	1600
<i>Tlalocohyla loquax</i>	29	24.6	2118.9
<i>Tlalocohyla picta</i>	9	23.9	2059
<i>Tlalocohyla smithii</i>	71	25.0	1089.5
<i>Trachycephalus coriaceus</i>	12	25.6	2899.6
<i>Trachycephalus imitatrix</i>	3	18.2	1875.3
<i>Trachycephalus jordani</i>	3	23.9	2867.7
<i>Trachycephalus mesophaeus</i>	5	22.6	1746.6
<i>Trachycephalus nigromaculatus</i>	5	21.9	1728
<i>Trachycephalus resinifricrix</i>	5	26.0	2924
<i>Trachycephalus venulosus</i>	21	25.2	2176.3
<i>Tripurion petasatus</i>	24	25.3	1255.3
<i>Xenohyla truncata</i>	1	23.6	1515
<b>Pelodyadinae</b>			
<i>Cyclorana alboguttata</i>	9	22.9	949.22
<i>Cyclorana australis</i>	7	22.7	1125.1

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<i>Cyclorana brevipes</i>	14	20.8	662.14
<i>Cyclorana cryptotis</i>	11	28.0	949.46
<i>Cyclorana longipes</i>	14	27.3	1084.8
<i>Cyclorana manya</i>	3	26.1	1359.3
<i>Litoria amboinensis</i>	1	23.9	4678
<i>Litoria arfakiana</i>	12	21.6	4334.3
<i>Litoria aurea</i>	11	17.2	1123.6
<i>Litoria bicolor</i>	9	27.0	1642.7
<i>Litoria booroolongensis</i>	12	14.1	1020.7
<i>Litoria caerulea</i>	14	22.7	1875.6
<i>Litoria chloris</i>	15	17.0	1502.2
<i>Litoria coplandi</i>	12	27.4	1495.2
<i>Litoria cyclorhyncha</i>	5	16.5	472.6
<i>Litoria dahlii</i>	9	27.7	1792.6
<i>Litoria dentata</i>	11	16.9	1264.3
<i>Litoria eucnemis</i>	5	22.9	3452.6
<i>Litoria fallax</i>	11	19.4	1555.7
<i>Litoria freycyneti</i>	10	17.3	1153.8
<i>Litoria genimaculata</i>	9	22.1	2731.8
<i>Litoria gilleni</i>	6	21.4	341.5
<i>Litoria gracilentata</i>	8	21.5	2050

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<i>Litoria inermis</i>	12	26.1	1323.9
<i>Litoria infrafrenata</i>	9	25.2	2562.9
<i>Litoria lesueurii</i>	10	19.4	1665.6
<i>Litoria meiriana</i>	10	26.5	1514.7
<i>Litoria microbelos</i>	7	26.8	1841.1
<i>Litoria modica</i>	3	20.2	3690.3
<i>Litoria nannotis</i>	8	21.3	2442.1
<i>Litoria nasuta</i>	9	26.4	1676.6
<i>Litoria pallida</i>	10	27.0	1270.6
<i>Litoria peronii</i>	10	17.2	646.6
<i>Litoria phyllochroa</i>	10	16.3	1284.3
<i>Litoria rothii</i>	7	26.4	1500.7
<i>Litoria rubella</i>	11	24.1	1226
<i>Litoria splendida</i>	6	28.6	916.67
<i>Litoria subglandulosa</i>	6	14.8	1454
<i>Litoria thesaurensis</i>	1	21.4	3669
<i>Litoria tornieri</i>	11	27.7	1647.5
<i>Litoria watjulumensis</i>	9	27.8	1271
<i>Litoria wollastoni</i>	4	21.9	2915.3
<i>Litoria xanthomera</i>	8	20.8	2609.1
<i>Nyctimystes dayi</i>	6	23.3	2869.2

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<i>Nyctimystes foricula</i>	3	20.0	4173.3
<i>Nyctimystes kubori</i>	4	17.5	3090
<i>Nyctimystes narinosus</i>	1	18.5	2272
<i>Nyctimystes papua</i>	2	22.7	2532
<i>Nyctimystes pulcher</i>	4	21.8	3904.3
<b>Phyllomedusinae</b>			
<i>Agalychnis annae</i>	6	20.4	3069.8
<i>Agalychnis callidryas</i>	11	25.1	2767.2
<i>Agalychnis litodryas</i>	5	24.9	3178.6
<i>Agalychnis moreleti</i>	5	20.0	2323.2
<i>Agalychnis saltator</i>	7	25.7	3053.4
<i>Agalychnis spurrelli</i>	2	26.5	3630.5
<i>Cruzirohyla calcarifer</i>	2	26.4	3452
<i>Hylomantis granulosa</i>	1	23.1	1275
<i>Hylomantis lemur</i>	2	21.8	2822.5
<i>Pachymedusa dacnicolor</i>	127	25.3	990.22
<i>Phasmahyla cochranae</i>	3	18.4	2059.3
<i>Phasmahyla guttata</i>	2	23.1	1986
<i>Phrynomedusa marginata</i>	4	21.5	2011
<i>Phyllomedusa atelopoides</i>	2	25.4	2755
<i>Phyllomedusa azurea</i>	10	23.5	1378.6

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<i>Phyllomedusa bicolor</i>	9	26.9	2548.3
<i>Phyllomedusa boliviana</i>	4	24.6	1914.8
<i>Phyllomedusa duellmani</i>	2	17.6	1200.5
<i>Phyllomedusa hypochondrialis</i>	14	24.8	1872.1
<i>Phyllomedusa palliata</i>	5	25.4	3100.8
<i>Phyllomedusa perinesos</i>	3	19.7	3053
<i>Phyllomedusa tarsi</i>	5	26.0	2657
<i>Phyllomedusa tetraploidea</i>	2	19.1	1407.5
<i>Phyllomedusa tomopterna</i>	7	25.8	2685
<i>Phyllomedusa trinitatus</i>	3	26.3	1688
<i>Phyllomedusa vaillanti</i>	7	25.9	2827.6

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842 Table S5. Genera of hylid frogs, indicating the major clade to which they belong,  
 843 the average values of mean annual temperature (° C) and mean annual  
 844 precipitation (mm), based on averages for sampled species (Table S4) and  
 845 estimated rates of body-size evolution. Several genera lack estimates of rates of  
 846 body-size evolution, either because they contain less than 3 species or because  
 847 less than 3 species are included in our phylogeny.

Genera	Clade	Mean temp.	Mean	
			precip.	Size rate
<i>Aplastodiscus</i>	Cophomantini	17.900	1986.36	0.00066
<i>Bokermannohyla</i>	Cophomantini	18.150	2237.00	0.00275
<i>Hyloscirtus</i>	Cophomantini	17.500	1821.83	0.00080
<i>Hypsiboas</i>	Cophomantini	20.907	2071.14	0.00290
<i>Myersiohyla</i>	Cophomantini	16.200	2372.50	
<i>Acris</i>	Hylini	17.300	1238.60	
<i>Anotheca</i>	Hylini	21.413	2314.93	
<i>Bromeliohyla</i>	Hylini	19.777	2307.00	
<i>Charadrahyla</i>	Hylini	18.600	2388.78	
<i>Diaglena</i>	Hylini	25.652	862.33	
<i>Duellmanohyla</i>	Hylini	20.475	2533.52	
<i>Ecnomiohyla</i>	Hylini	20.800	2217.89	
<i>Exerodonta</i>	Hylini	22.100	1822.80	0.00506

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<i>Hyla</i>	Hylini	15.600	1126.14	0.00310
<i>Isthmohyla</i>	Hylini	17.700	2895.97	0.00192
<i>Megastomatohyla</i>	Hylini	20.600	3544.00	
<i>Plectrohyla</i>	Hylini	16.400	1668.92	0.00180
<i>Pseudacris</i>	Hylini	14.100	1020.70	0.00331
<i>Ptychohyla</i>	Hylini	19.600	2169.37	0.00018
<i>Smilisca</i>	Hylini	23.900	2632.16	0.00316
<i>Tlalocohyla</i>	Hylini	24.000	1716.85	0.00255
<i>Triprion</i>	Hylini	25.271	1255.33	
<i>Aparasphenodon</i>	Lophiohylini	23.800	1378.67	
<i>Argenteohyla</i>	Lophiohylini	17.200	1208.50	
<i>Corythomantis</i>	Lophiohylini	26.100	1017.00	
<i>Itapotihyla</i>	Lophiohylini	20.167	2141.67	
<i>Nyctimantis</i>	Lophiohylini	24.125	4400.75	
<i>Osteocephalus</i>	Lophiohylini	24.700	3202.57	0.00699
<i>Osteopilus</i>	Lophiohylini	23.600	1659.22	0.00820
<i>Phyllodytes</i>	Lophiohylini	23.100	1744.00	
<i>Tepuihyla</i>	Lophiohylini	14.700	2969.60	
<i>Trachycephalus</i>	Lophiohylini	23.300	2316.78	0.00379
	<i>Dendropsophus</i>			
<i>Dendropsophus</i>	clade	23.500	2252.41	0.00127

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<i>Dendropsophus</i>				
<i>Xenohyla</i>	clade	23.600	1515.00	
<i>Scinax</i>	<i>Scinax</i> clade	21.900	2234.08	0.00098
<i>Sphaenorhynchus</i>	<i>Scinax</i> clade	23.500	2498.88	
<i>Lysapsus</i>	<i>Pseudis</i> clade	26.100	1827.67	0.00057
<i>Podonectes</i>	<i>Pseudis</i> clade	16.500	1630.42	
<i>Pseudis</i>	<i>Pseudis</i> clade	22.500	1431.56	0.00143
<i>Scarthyla</i>	<i>Pseudis</i> clade	25.350	2755.00	
<i>Agalychnis</i>	Phyllomedusinae	23.800	3003.79	0.00198
<i>Cruziophyla</i>	Phyllomedusinae	26.400	3452.00	
<i>Hylomantis</i>	Phyllomedusinae	22.425	2048.75	
<i>Pachymedusa</i>	Phyllomedusinae	25.267	990.22	
<i>Phasmahyla</i>	Phyllomedusinae	20.700	2022.67	
<i>Phrynomedusa</i>	Phyllomedusinae	21.525	2011.00	
<i>Phyllomedusa</i>	Phyllomedusinae	23.900	2237.55	0.00265
Pelodryadinae	Pelodryadinae	22.300	1930.33	0.00444

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851 Table S6. Genera of hylid frogs, indicating the major clade to which they belong,  
 852 species richness, estimated age (based on the time-calibrated phylogeny), and  
 853 estimated diversification rate. Note that subfamily Pelodryadinae presently  
 854 consists of a single monophyletic genus (*Litoria*).

Genera	Clade	Species richness	Stem age	Div. rate (stem, $e = 0.45$ )
<i>Aplastodiscus</i>	Cophomantini	15	52.45	0.0412
<i>Bokermannohyla</i>	Cophomantini	27	56.69	0.0481
<i>Hyloscirtus</i>	Cophomantini	31	64.01	0.0447
<i>Hypsiboas</i>	Cophomantini	83	52.45	0.073
<i>Myersiohyla</i>	Cophomantini	4	71.18	0.0137
<i>Acris</i>	Hylini	3	44.4	0.017
<i>Anotheca</i>	Hylini	1	18.49	0.000
<i>Bromeliohyla</i>	Hylini	2	28.21	0.016
<i>Charadrahyla</i>	Hylini	5	39.34	0.030
<i>Diaglena</i>	Hylini	1	24.36	0.000
<i>Duellmanohyla</i>	Hylini	8	28.21	0.056
<i>Ecnomiohyla</i>	Hylini	11	43.42	0.043
<i>Exerodonta</i>	Hylini	11	33.62	0.056
<i>Hyla</i>	Hylini	36	42.05	0.0715

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<i>Isthmohyla</i>	Hylini	15	38.47	0.0562
<i>Megastomatohyla</i>	Hylini	4	39.34	0.0248
<i>Plectrohyla</i>	Hylini	42	33.62	0.094
<i>Pseudacris</i>	Hylini	16	44.4	0.0501
<i>Ptychohyla</i>	Hylini	13	28.21	0.0719
<i>Smilisca</i>	Hylini	8	32.64	0.0484
<i>Tlalocohyla</i>	Hylini	4	40.85	0.0239
<i>Tripurion</i>	Hylini	1	18.49	0
<i>Aparasphenodon</i>	Lophiohylini	4	22.8	0.0427
<i>Argenteohyla</i>	Lophiohylini	1	19.27	0
<i>Corythomantis</i>	Lophiohylini	1	35.43	0
<i>Itapotihyla</i>	Lophiohylini	1	39.01	0
<i>Nyctimantis</i>	Lophiohylini	1	19.27	0
<i>Osteocephalus</i>	Lophiohylini	21	30.01	0.0828
<i>Osteopilus</i>	Lophiohylini	8	39.28	0.0402
<i>Phyllodytes</i>	Lophiohylini	12	42.08	0.0464
<i>Tepuihyla</i>	Lophiohylini	8	30.01	0.0526
<i>Trachycephalus</i>	Lophiohylini	12	40.88	0.0478
	<i>Dendropsophus</i>			
<i>Dendropsophus</i>	clade	94	54.12	0.0731
<i>Xenohyla</i>	<i>Dendropsophus</i>	2	54.12	0.0081

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	clade			
<i>Scinax</i>	<i>Scinax</i> clade	96	68.31	0.0582
<i>Sphaenorhynchus</i>	<i>Scinax</i> clade	14	68.31	0.0307
<i>Lysapsus</i>	<i>Pseudis</i> clade	4	29.33	0.0332
<i>Podonectes</i>	<i>Pseudis</i> clade	2	29.33	0.0149
<i>Pseudis</i>	<i>Pseudis</i> clade	5	33.31	0.0349
<i>Scarthyla</i>	<i>Pseudis</i> clade	2	48.16	0.0091
<i>Agalychnis</i>	Phyllomedusinae	6	22.55	0.0586
<i>Cruzirohyla</i>	Phyllomedusinae	2	53.49	0.0082
<i>Hylomantis</i>	Phyllomedusinae	8	37.74	0.0418
<i>Pachymedusa</i>	Phyllomedusinae	1	22.55	0
<i>Phasmahyla</i>	Phyllomedusinae	7	43.32	0.0337
<i>Phrynomedusa</i>	Phyllomedusinae	5	45.48	0.0256
<i>Phyllomedusa</i>	Phyllomedusinae	32	43.32	0.0668
Pelodryadinae				
( <i>Litoria</i> )	Pelodryadinae	189	71.91	0.0646

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858 Table S7. Maximum male body size (SVL in mm) reported for hylid species  
 859 included in the phylogeny and/or local communities, indicating the literature  
 860 sources from which the size data are taken (see references listed immediately  
 861 below the table). Species are arranged alphabetically within clades and clades  
 862 are listed alphabetically as well.

	Max.		
Species	SVL	Literature source	Clade
<i>Aplastodiscus albofrenatus</i>	40	Lutz 1973	Cophomantini
<i>Aplastodiscus albosignatus</i>	52	Lutz 1973	Cophomantini
<i>Aplastodiscus arildae</i>	41.6	Heyer et al. 1990 Cruz and Peixoto	Cophomantini
<i>Aplastodiscus callipygius</i>	50.7	1984	Cophomantini
<i>Aplastodiscus cochranæ</i>	46.5	Garcia et al. 2001 Cruz and Peixoto	Cophomantini
<i>Aplastodiscus ehrhardti</i>	39.1	1985 Carvalho-e-Silva and Carvalho-e-	Cophomantini
<i>Aplastodiscus eugenioi</i>	36.1	Silva 2005 Cruz and Peixoto	Cophomantini
<i>Aplastodiscus flumineus</i>	50.4	1984	Cophomantini

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		Cruz, Pimenta, and	
<i>Aplastodiscus ibirapitanga</i>	41	Silvano 2003	Cophomantini
		Cruz and Peixoto	
<i>Aplastodiscus leucopygius</i>	45.1	1984	Cophomantini
<i>Aplastodiscus musicus</i>	50	Cochran 1955	Cophomantini
<i>Aplastodiscus perviridis</i>	46.1	Garcia et al. 2001	Cophomantini
		Cruz, Pimenta, and	
<i>Aplastodiscus sibilatus</i>	33.6	Silvano 2003	Cophomantini
		Napoli and	
<i>Bokermannohyla ahenea</i>	56.7	Caramaschi 2004	Cophomantini
<i>Bokermannohyla alvarengai</i>	80	Lutz 1973	Cophomantini
<i>Bokermannohyla astartea</i>	41.5	Heyer et al. 1990	Cophomantini
<i>Bokermannohyla caramaschii</i>	70	Napoli 2005	Cophomantini
<i>Bokermannohyla carvalhoi</i>	67	Peixoto 1981	Cophomantini
<i>Bokermannohyla circumdata</i>	70	Lutz 1973	Cophomantini
<i>Bokermannohyla claresignata</i>	42	Lutz 1973	Cophomantini
<i>Bokermannohyla clepsydra</i>	39	Cochran 1955	Cophomantini
		Napoli and Junca	
<i>Bokermannohyla diamantina</i>	51.7	2006	Cophomantini
		Napoli and	
<i>Bokermannohyla feioi</i>	40.2	Caramaschi 2004	Cophomantini

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		Pombal and	
<i>Bokermannohyla gouveai</i>	69	Haddad 1993	Cophomantini
<i>Bokermannohyla hylax</i>	61.5	Heyer et al. 1990	Cophomantini
<i>Bokermannohyla ibitiguara</i>	44.1	Cardoso 1983	Cophomantini
		Caramaschi and	
<i>Bokermannohyla ibitipoca</i>	42.7	Feio 1990	Cophomantini
		Jim and	
<i>Bokermannohyla izecksohni</i>	50.8	Caramaschi 1979	Cophomantini
<i>Bokermannohyla langei</i>	66	Lutz 1973	Cophomantini
		Napoli and Silva-	
<i>Bokermannohyla lucianae</i>	49.2	Pimenta 2003	Cophomantini
		Pombal and	
<i>Bokermannohyla luctuosa</i>	60.6	Haddad 1993	Cophomantini
<i>Bokermannohyla martinsi</i>	64	Lutz 1973	Cophomantini
		Bokermann and	
<i>Bokermannohyla nanuzae</i>	42	Sazima, 1973	Cophomantini
		Caramaschi,	
		Napoli, and	
<i>Bokermannohyla ravidia</i>	47.6	Bernardes, 2001	Cophomantini
<i>Bokermannohyla saxicola</i>	45	Lutz 1973	Cophomantini
<i>Bokermannohyla sazimai</i>	36.4	Cardoso and	Cophomantini

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		Andrade 1982	
		de Vasconcelos	
<i>Bokermannohyla vulcaniae</i>	50.2	and Giaretta 2003	Cophomantini
		Cochran and Goin	
<i>Hyloscirtus albopunctulatus</i>	41.5	1970	Cophomantini
<i>Hyloscirtus alytolylax</i>	37	Duellman 1972	Cophomantini
		Duellman et al.	
<i>Hyloscirtus armatus</i>	68.5	1997	Cophomantini
		Ruiz-Carranza and	
<i>Hyloscirtus bogotensis</i>	49.5	Lynch 1982	Cophomantini
		Ardila-Robayo et	
<i>Hyloscirtus caucanus</i>	61.9	al. 1993	Cophomantini
<i>Hyloscirtus charazani</i>	55	Vellard 1970	Cophomantini
<i>Hyloscirtus colymba</i>	37	Duellman 2001	Cophomantini
<i>Hyloscirtus denticulatus</i>	44.2	Duellman 1972	Cophomantini
<i>Hyloscirtus jahni</i>	34.5	Rivero 1961	Cophomantini
		Duellman and	
<i>Hyloscirtus larinopygion</i>	55.6	Berger 1982	Cophomantini
<i>Hyloscirtus lascinius</i>	38	Rivero 1969	Cophomantini
		Duellman and	
<i>Hyloscirtus lindae</i>	68.1	Altig 1978	Cophomantini

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		Ardila-Robayo and	
		Ruiz-Carranza	
<i>Hyloscirtus lynchi</i>	46.4	1991	Cophomantini
		Duellman and	
<i>Hyloscirtus pacha</i>	60.8	Hillis 1990	Cophomantini
<i>Hyloscirtus palmeri</i>	45	Duellman 2001	Cophomantini
		Duellman and	
<i>Hyloscirtus pantostictus</i>	63	Berger 1982	Cophomantini
<i>Hyloscirtus phyllognathus</i>	34	Duellman 1972	Cophomantini
		Ruiz-Carranza and	
<i>Hyloscirtus piceigularis</i>	36.9	Lynch 1982	Cophomantini
<i>Hyloscirtus platydactylus</i>	39.4	Duellman 1972	Cophomantini
		Duellman and	
<i>Hyloscirtus psarolaimus</i>	55.6	Hillis 1990	Cophomantini
		Duellman and	
<i>Hyloscirtus ptychodactylus</i>	67.5	Hillis 1990	Cophomantini
		Ruiz-Carranza and	
<i>Hyloscirtus sarampiona</i>	68.8	Lynch 1982	Cophomantini
<i>Hyloscirtus simmonsii</i>	37.8	Duellman 1989	Cophomantini
		Duellman and	
<i>Hyloscirtus staufferorum</i>	56.9	Coloma 1993	Cophomantini

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<i>Hyloscirtus tapichalaca</i>	63.8	Kizirian et al. 2003	Cophomantini
		Duellman and	
<i>Hyloscirtus torrenticola</i>	35.5	Altig 1978	Cophomantini
<i>Hypsiboas albomarginatus</i>	55	Lutz 1973	Cophomantini
		Duellman et al.	
<i>Hypsiboas alboniger</i>	56	1997	Cophomantini
<i>Hypsiboas albopunctatus</i>	60	Lutz 1973	Cophomantini
<i>Hypsiboas alemani</i>	30.5	Rivero 1964	Cophomantini
		Duellman et al.	
<i>Hypsiboas andinus</i>	57.6	1997	Cophomantini
		Caramaschi and	
<i>Hypsiboas atlanticus</i>	40.2	Velosa 1996	Cophomantini
		Duellman et al.	
<i>Hypsiboas balzani</i>	50.4	1997	Cophomantini
		Caramaschi and	
<i>Hypsiboas beckeri</i>	29	Cruz 2004	Cophomantini
<i>Hypsiboas benitezi</i>	37	Rivero 1961	Cophomantini
<i>Hypsiboas bischoffi</i>	46.1	Heyer et al. 1990	Cophomantini
<i>Hypsiboas boans</i>	132	Duellman 2001	Cophomantini
		Caramaschi and	
<i>Hypsiboas buriti</i>	31.9	Cruz 1999	Cophomantini

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		Lavilla and Cei	
<i>Hypsiboas caingua</i>	33.1	2001	Cophomantini
<i>Hypsiboas calcaratus</i>	47.5	Duellman 2005	Cophomantini
<i>Hypsiboas callipleura</i>	45	Boulenger 1902	Cophomantini
		Rodríguez and	
<i>Hypsiboas cinerascens</i>	44	Duellman 1994	Cophomantini
<i>Hypsiboas cipoensis</i>	33	Lutz 1973	Cophomantini
<i>Hypsiboas cordobae</i>	50	Lutz 1973	Cophomantini
<i>Hypsiboas crepitans</i>	63	Lutz 1973	Cophomantini
<i>Hypsiboas cymbalum</i>	49	Lutz 1973	Cophomantini
		Caramaschi and	
<i>Hypsiboas ericae</i>	34	Cruz 2000	Cophomantini
		Caramaschi and	
<i>Hypsiboas exastis</i>	99	Rodrigues 2003	Cophomantini
<i>Hypsiboas faber</i>	104	Heyer et al. 1990	Cophomantini
<i>Hypsiboas fasciatus</i>	40.3	Duellman 2005	Cophomantini
		Carnaval and	
<i>Hypsiboas freicanecae</i>	42.2	Peixoto 2004	Cophomantini
		Goin and Goin	
<i>Hypsiboas fuentei</i>	57	1968	Cophomantini
<i>Hypsiboas geographicus</i>	62	Rodríguez and	Cophomantini

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		Duellman 1994	
		Cruz and	
<i>Hypsiboas goianus</i>	37.6	Caramaschi 1998	Cophomantini
<i>Hypsiboas guentheri</i>	40	Lutz 1973	Cophomantini
		Trueb and Tyler	
<i>Hypsiboas heilprini</i>	54.3	1974	Cophomantini
		Cochran and Goin	
<i>Hypsiboas hobbsi</i>	42.5	1970	Cophomantini
		Pyburn and Hall	
<i>Hypsiboas hutchinsi</i>	47.1	1984	Cophomantini
<i>Hypsiboas joaquinii</i>	51.5	Lutz 1973	Cophomantini
		Rodríguez and	
<i>Hypsiboas lanciformis</i>	80	Duellman 1994	Cophomantini
		Caramaschi and	
<i>Hypsiboas latistriatus</i>	40.6	Cruz 2004	Cophomantini
<i>Hypsiboas lemai</i>	30.4	Duellman 1997	Cophomantini
		Cruz and	
<i>Hypsiboas leptolineatus</i>	31.6	Caramaschi 1998	Cophomantini
		Caramaschi and	
<i>Hypsiboas leucocheilus</i>	67.9	Niemeyer 2003	Cophomantini
<i>Hypsiboas lundii</i>	76	Bokermann and	Cophomantini

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		Sazima 1973	
		Caramaschi and	
<i>Hypsiboas marginatus</i>	51.1	Cruz 2000	Cophomantini
		Duellman et al.	
<i>Hypsiboas marianitae</i>	56.8	1997	Cophomantini
		Duellman et al.	
<i>Hypsiboas melanopleura</i>	43.6	1997	Cophomantini
		Rodríguez and	
<i>Hypsiboas microderma</i>	34	Duellman 1994	Cophomantini
<i>Hypsiboas multifasciatus</i>	57.3	Duellman 1997	Cophomantini
		Faivovich et al.	
<i>Hypsiboas nympa</i>	31.2	2006	Cophomantini
<i>Hypsiboas ornatissimus</i>	31	Lutz 1973	Cophomantini
		Duellman et al.	
<i>Hypsiboas palaestes</i>	50.4	1997	Cophomantini
<i>Hypsiboas pardalis</i>	69	Lutz 1973	Cophomantini
		Cochran and Goin	
<i>Hypsiboas pellucens</i>	61.6	1970	Cophomantini
		Caramaschi and	
<i>Hypsiboas phaeopleura</i>	35.2	Cruz 2000	Cophomantini
<i>Hypsiboas polytaenius</i>	31.4	Cruz and	Cophomantini

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		Caramaschi 1998	
		Caramaschi et al.	
<i>Hypsiboas pombali</i>	55.6	2004	Cophomantini
<i>Hypsiboas prasinus</i>	55	Cochran 1955	Cophomantini
<i>Hypsiboas pugnax</i>	77.9	Duellman 2001	Cophomantini
<i>Hypsiboas pulchellus</i>	50	Lutz 1973	Cophomantini
<i>Hypsiboas pulidoi</i>	23.2	Rivero 1968	Cophomantini
		Rodriguez and	
<i>Hypsiboas punctatus</i>	40	Duellman 1994	Cophomantini
		Caramaschi and	
<i>Hypsiboas raniceps</i>	71	Niemeyer 2003	Cophomantini
		Senaris and	
<i>Hypsiboas rhythmicus</i>	34.2	Ayarzaguena 2002	Cophomantini
<i>Hypsiboas riojanus</i>	56	Cei 1980	Cophomantini
		Duellman and	
<i>Hypsiboas roraima</i>	45.5	Hoogmoed 1992	Cophomantini
<i>Hypsiboas rosenbergi</i>	90	Duellman 2001	Cophomantini
		Cochran and Goin	
<i>Hypsiboas rubracylus</i>	50.4	1970	Cophomantini
<i>Hypsiboas rufitelus</i>	49.2	Duellman 2001	Cophomantini
<i>Hypsiboas secedens</i>	57	Lutz 1973	Cophomantini

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		Caramaschi and	
<i>Hypsiboas semiguttatus</i>	41.6	Cruz 2000	Cophomantini
<i>Hypsiboas sibleszi</i>	34.9	Duellman 1997	Cophomantini
		Caramaschi and	
<i>Hypsiboas stenocephalus</i>	30.4	Cruz 1999	Cophomantini
		Lavilla and Cei	
<i>Hypsiboas varelae</i>	52.9	2001	Cophomantini
<i>Hypsiboas wavrini</i>	113	Hoogmoed 1990	Cophomantini
			<i>Dendropsophus</i>
<i>Dendropsophus acreanus</i>	35	Lutz 1973	clade
			<i>Dendropsophus</i>
<i>Dendropsophus allenorum</i>	21.4	Duellman 2005	clade
		Mijares-Urrutia	<i>Dendropsophus</i>
<i>Dendropsophus amicornum</i>	22.6	1998	clade
		Napoli and	<i>Dendropsophus</i>
<i>Dendropsophus anataliasiasi</i>	21.8	Caramaschi 1999	clade
			<i>Dendropsophus</i>
<i>Dendropsophus anceps</i>	40	Lutz 1973	clade
			<i>Dendropsophus</i>
<i>Dendropsophus aperomeus</i>	21.3	Duellman 1982	clade
<i>Dendropsophus araguaya</i>	20.5	Napoli and	<i>Dendropsophus</i>

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		Caramaschi 1998	clade
			<i>Dendropsophus</i>
<i>Dendropsophus baileyi</i>	21.5	Cochran 1952	clade
			<i>Dendropsophus</i>
<i>Dendropsophus battersbyi</i>	33	Rivero 1961	clade
			<i>Dendropsophus</i>
<i>Dendropsophus berthallutzae</i>	21	Lutz 1973	clade
			<i>Dendropsophus</i>
<i>Dendropsophus bifurcus</i>	28	Duellman 1978	clade
			<i>Dendropsophus</i>
<i>Dendropsophus bipunctatus</i>	25	Lutz 1973	clade
		Cochran and Goin	<i>Dendropsophus</i>
<i>Dendropsophus bogerti</i>	33.3	1970	clade
			<i>Dendropsophus</i>
<i>Dendropsophus bokermanni</i>	24	Duellman 1978	clade
			<i>Dendropsophus</i>
<i>Dendropsophus branneri</i>	19	Lutz 1973	clade
			<i>Dendropsophus</i>
<i>Dendropsophus brevifrons</i>	22	Duellman 1978	clade
		Napoli and	<i>Dendropsophus</i>
<i>Dendropsophus cachimbo</i>	21	Caramaschi 1999	clade

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			<i>Dendropsophus</i>
<i>Dendropsophus carnifex</i>	27.7	Duellman 1969	clade
		Napoli and	<i>Dendropsophus</i>
<i>Dendropsophus cerradensis</i>	19.3	Caramaschi 1998	clade
			<i>Dendropsophus</i>
<i>Dendropsophus coffeus</i>	21.2	Koehler et al. 2005	clade
		Duellman and	<i>Dendropsophus</i>
<i>Dendropsophus columbianus</i>	29.3	Trueb 1983	clade
		Pombal and Bastos	<i>Dendropsophus</i>
<i>Dendropsophus cruzi</i>	19.4	1998	clade
			<i>Dendropsophus</i>
<i>Dendropsophus decipiens</i>	20	Lutz 1973	clade
		Koehler and	<i>Dendropsophus</i>
<i>Dendropsophus delarivai</i>	19.4	Loetters 2001	clade
		Gomes and	<i>Dendropsophus</i>
<i>Dendropsophus dutrai</i>	34.2	Peixoto 1996	clade
			<i>Dendropsophus</i>
<i>Dendropsophus ebraccatus</i>	27.8	Duellman 2001	clade
			<i>Dendropsophus</i>
<i>Dendropsophus elegans</i>	29.6	<b>Lutz 1973</b>	clade
<i>Dendropsophus elianeae</i>	25.5	Napoli and	<i>Dendropsophus</i>

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		Caramaschi 2000	clade
		Lescure and Marty	<i>Dendropsophus</i>
<i>Dendropsophus gaucheri</i>	19.2	2000	clade
		Weygoldt and	<i>Dendropsophus</i>
<i>Dendropsophus giesleri</i>	25	Peixoto 1987	clade
			<i>Dendropsophus</i>
<i>Dendropsophus grandisonae</i>	20.8	Goin 1966	clade
			<i>Dendropsophus</i>
<i>Dendropsophus gryllatus</i>	25.5	Duellman 1973	clade
		Bastos and Pombal	<i>Dendropsophus</i>
<i>Dendropsophus haddadi</i>	19.45	1996	clade
<i>Dendropsophus</i>		Rodríguez and	<i>Dendropsophus</i>
<i>haraldschultzi</i>	22	Duellman 1994	clade
		Napoli and	<i>Dendropsophus</i>
<i>Dendropsophus jimi</i>	20.9	Caramaschi 1999	clade
		Koehler and	<i>Dendropsophus</i>
<i>Dendropsophus joannae</i>	18.6	Loetters 2001	clade
			<i>Dendropsophus</i>
<i>Dendropsophus koechlini</i>	24	Duellman 2005	clade
			<i>Dendropsophus</i>
<i>Dendropsophus labialis</i>	43	Amezquita 1999	clade

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			<i>Dendropsophus</i>
<i>Dendropsophus leali</i>	23	Duellman 2005	clade
<i>Dendropsophus leucophyllatus</i>	36	Rodríguez and Duellman 1994	<i>Dendropsophus</i> clade
			<i>Dendropsophus</i>
<i>Dendropsophus limai</i>	19	Bokermann 1962	clade
			<i>Dendropsophus</i>
<i>Dendropsophus luteocellatus</i>	31	Rivero 1961	clade
		Rodríguez and	<i>Dendropsophus</i>
<i>Dendropsophus marmoratus</i>	44	Duellman 1994	clade
		Cochran and Goin	<i>Dendropsophus</i>
<i>Dendropsophus mathiassoni</i>	21.4	1970	clade
<i>Dendropsophus melanargyreus</i>	34	Lutz 1973	<i>Dendropsophus</i> clade
			<i>Dendropsophus</i>
<i>Dendropsophus meridianus</i>	19	Lutz 1973	clade
<i>Dendropsophus microcephalus</i>	24.5	Duellman 2001	<i>Dendropsophus</i> clade
			<i>Dendropsophus</i>
<i>Dendropsophus microps</i>	26	Lutz 1973	clade
<i>Dendropsophus minusculus</i>	20.6	Duellman 1997	<i>Dendropsophus</i>

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			clade
			<i>Dendropsophus</i>
<i>Dendropsophus minutus</i>	23	Duellman 1997	clade
		Vigle and	
		Goberdhan-Vigle	<i>Dendropsophus</i>
<i>Dendropsophus miyatai</i>	18.1	1990	clade
			<i>Dendropsophus</i>
<i>Dendropsophus nahdereri</i>	45	Lutz 1973	clade
			<i>Dendropsophus</i>
<i>Dendropsophus nanus</i>	22	Lutz 1973	clade
		Carvalho-e-Silva et	<i>Dendropsophus</i>
<i>Dendropsophus oliveirai</i>	17	al. 2003	clade
		Kaplan and Ruiz	<i>Dendropsophus</i>
<i>Dendropsophus padreluna</i>	30.4	1997	clade
			<i>Dendropsophus</i>
<i>Dendropsophus parviceps</i>	21.9	Duellman 2005	clade
			<i>Dendropsophus</i>
<i>Dendropsophus pauiniensis</i>	20.3	Heyer 1977	clade
			<i>Dendropsophus</i>
<i>Dendropsophus pelidna</i>	36.9	Duellman 1989	clade
<i>Dendropsophus phlebodes</i>	23.6	Duellman 2001	<i>Dendropsophus</i>

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			clade
		Duellman and	<i>Dendropsophus</i>
<i>Dendropsophus praestans</i>	31.5	Trueb 1983	clade
<i>Dendropsophus</i>		Cruz, Caramaschi,	<i>Dendropsophus</i>
<i>pseudomeridianus</i>	19.9	and Dias 2000	clade
		Napoli and	<i>Dendropsophus</i>
<i>Dendropsophus rhea</i>	20.7	Caramaschi 1999	clade
			<i>Dendropsophus</i>
<i>Dendropsophus rhodopeplus</i>	24.2	Duellman 2005	clade
		Rodríguez and	<i>Dendropsophus</i>
<i>Dendropsophus riveroi</i>	20	Duellman 1994	clade
<i>Dendropsophus</i>			<i>Dendropsophus</i>
<i>robertmertensi</i>	26.4	Duellman 2001	clade
		Rodríguez and	<i>Dendropsophus</i>
<i>Dendropsophus rossalleni</i>	20	Duellman 1994	clade
		Napoli and	<i>Dendropsophus</i>
<i>Dendropsophus rubicundulus</i>	23.8	Caramaschi 1999	clade
		Weygoldt and	<i>Dendropsophus</i>
<i>Dendropsophus ruschii</i>	27.9	Peixoto 1987	clade
			<i>Dendropsophus</i>
<i>Dendropsophus sanborni</i>	17	Lutz 1973	clade

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		Rodríguez and	<i>Dendropsophus</i>
<i>Dendropsophus sarayacuensis</i>	29	Duellman 1994	clade
			<i>Dendropsophus</i>
<i>Dendropsophus sartori</i>	26	Duellman 2001	clade
			<i>Dendropsophus</i>
<i>Dendropsophus schubarti</i>	19.5	Duellman 2005	clade
			<i>Dendropsophus</i>
<i>Dendropsophus seniculus</i>	37.7	Heyer et al. 1990	clade
		Caramaschi and	<i>Dendropsophus</i>
<i>Dendropsophus soaresi</i>	31.7	Jim 1983	clade
			<i>Dendropsophus</i>
<i>Dendropsophus stingi</i>	24.3	Kaplan 1994	clade
		Carvalho-e-Silva et	<i>Dendropsophus</i>
<i>Dendropsophus studerae</i>	24.8	al. 2003	clade
		Duellman and	<i>Dendropsophus</i>
<i>Dendropsophus subocularis</i>	23.1	Crump 1974	clade
		Martins and	<i>Dendropsophus</i>
<i>Dendropsophus timbeba</i>	21.9	Cardoso 1987	clade
<i>Dendropsophus</i>			<i>Dendropsophus</i>
<i>tintinnabulum</i>	20	Lutz 1973	clade
<i>Dendropsophus triangulum</i>	28	Rodríguez and	<i>Dendropsophus</i>

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		Duellman 1994	clade
			<i>Dendropsophus</i>
<i>Dendropsophus tritaeniatus</i>	20.5	Bokermann 1965	clade
		Kaplan and Ruiz	<i>Dendropsophus</i>
<i>Dendropsophus virolinensis</i>	26.7	1997	clade
			<i>Dendropsophus</i>
<i>Dendropsophus walfordi</i>	19.5	Bokermann 1962	clade
			<i>Dendropsophus</i>
<i>Dendropsophus weneri</i>	20	Lutz 1973	clade
		Martins and	<i>Dendropsophus</i>
<i>Dendropsophus xapuriensis</i>	18.4	Cardoso 1987	clade
		Mijares-Urrutia	<i>Dendropsophus</i>
<i>Dendropsophus yaracuyanus</i>	30.4	and River 2000	clade
			<i>Dendropsophus</i>
<i>Xenohyla eugenioi</i>	31.5	Caramaschi 1998	clade
<i>Acris crepitans</i>	29	Duellman 2001	Hylini
<i>Acris gryllus</i>	29	Lannoo 2005	Hylini
<i>Anothea spinosa</i>	68.5	Duellman 2001	Hylini
<i>Bromeliohyla bromeliacea</i>	29.5	Duellman 2001	Hylini
<i>Bromeliohyla dendroscarta</i>	31.6	Duellman 2001	Hylini
<i>Charadrahyla altipotens</i>	80.6	Duellman 2001	Hylini

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<i>Charadrahyla chaneque</i>	60.7	Duellman 2001	Hylini
<i>Charadrahyla nephila</i>	70.9	Duellman 2001	Hylini
<i>Charadrahyla taeniopus</i>	65.9	Duellman 2001	Hylini
<i>Charadrahyla trux</i>	81	Duellman 2001	Hylini
<i>Duellmanohyla chamulae</i>	30.5	Duellman 2001	Hylini
<i>Duellmanohyla ignicolor</i>	30.9	Duellman 2001	Hylini
<i>Duellmanohyla lythrodes</i>	32.6	Duellman 2001	Hylini
<i>Duellmanohyla rufiocularis</i>	30	Duellman 2001	Hylini
<i>Duellmanohyla salvavida</i>	28	Duellman 2001	Hylini
<i>Duellmanohyla schmidtorum</i>	32.8	Duellman 2001	Hylini
<i>Duellmanohyla soralia</i>	32.3	Duellman 2001	Hylini
<i>Duellmanohyla uranochroa</i>	36.8	Duellman 2001	Hylini
<i>Ecnomiohyla echinata</i>	57	Duellman 2001	Hylini
<i>Ecnomiohyla miliaria</i>	110.4	Duellman 2001	Hylini
<i>Ecnomiohyla minera</i>	83.1	Duellman 2001	Hylini
<i>Ecnomiohyla miotympanum</i>	38.4	Duellman 2001	Hylini
		Cochran and Goin	
<i>Ecnomiohyla phantasmagoria</i>	109.7	1970	Hylini
<i>Ecnomiohyla salvaje</i>	86	Duellman 2001	Hylini

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		Rodríguez and	
<i>Ecnomiohyla tuberculosa</i>	90	Duellman 1994	Hylini
<i>Ecnomiohyla valancifer</i>	77.7	Duellman 2001	Hylini
<i>Exerodonta abdivita</i>	27.5	Duellman 2001	Hylini
<i>Exerodonta bivocata</i>	28.5	Duellman 2001	Hylini
<i>Exerodonta catracha</i>	28.7	Duellman 2001	Hylini
<i>Exerodonta chimalapa</i>	24.9	Duellman 2001	Hylini
<i>Exerodonta juanita</i>	35.8	Duellman 2001	Hylini
<i>Exerodonta melanomma</i>	29.9	Duellman 2001	Hylini
<i>Exerodonta pinorum</i>	34.5	Duellman 2001	Hylini
<i>Exerodonta smaragdina</i>	26	Duellman 2001	Hylini
<i>Exerodonta sumichrasti</i>	27.7	Duellman 2001	Hylini
<i>Exerodonta xera</i>	27.9	Duellman 2001	Hylini
		Conant and Collins	
<i>Hyla andersonii</i>	51	1998	Hylini
<i>Hyla annectans</i>	35	Fei et al. 1999	Hylini
<i>Hyla arborea</i>	50	Arnold 2002	Hylini
<i>Hyla arenicolor</i>	52	Lannoo 2005	Hylini
<i>Hyla avivoca</i>	39	Lannoo 2005	Hylini
<i>Hyla bocourti</i>	34.6	Duellman 2001	Hylini
<i>Hyla chinensis</i>	32	Fei et al. 1999	Hylini

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		Wright and Wright	
<i>Hyla chrysoscelis</i>	51	1949	Hylini
		Wright and Wright	
<i>Hyla cinerea</i>	59	1949	Hylini
<i>Hyla euphorbiacea</i>	39.6	Duellman 2001	Hylini
<i>Hyla eximia</i>	34.9	Duellman 2001	Hylini
		Wright and Wright	
<i>Hyla femoralis</i>	37	1949	Hylini
		Wright and Wright	
<i>Hyla gratiosa</i>	68	1949	Hylini
		Goris and Maeda	
<i>Hyla hallowellii</i>	37	2003	Hylini
<i>Hyla immaculata</i>	31.4	Fei et al. 1999	Hylini
<i>Hyla intermedia</i>	50	Arnold 2002	Hylini
		Goris and Maeda	
<i>Hyla japonica</i>	39	2003	Hylini
<i>Hyla meridionalis</i>	50	Arnold 2002	Hylini
<i>Hyla plicata</i>	44	Duellman 2001	Hylini
<i>Hyla sanchiangensis</i>	32.7	Fei et al. 1999	Hylini
<i>Hyla sarda</i>	50	Arnold 2002	Hylini
<i>Hyla savignyi</i>	40	Tarkhnishvili and	Hylini

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Gokhelaashvili 1999			
<i>Hyla simplex</i>	34	Pope 1931	Hylini
<i>Hyla simplex</i>	37	Fei et al. 1999	Hylini
Wright and Wright			
<i>Hyla squirella</i>	36	1949	Hylini
<i>Hyla tsinglingensis</i>	41	Fei et al. 1999	Hylini
<i>Hyla ussuriensis</i>	40	Fei et al. 1999	Hylini
Wright and Wright			
<i>Hyla versicolor</i>	51	1949	Hylini
<i>Hyla walkeri</i>	35.9	Duellman 2001	Hylini
Degenhardt et al.			
<i>Hyla wrightorum</i>	44	1996	Hylini
<i>Hyla zhaopingensis</i>	30.1	Fei et al. 1999	Hylini
<i>Isthmohyla angustilineata</i>	34.2	Duellman 2001	Hylini
<i>Isthmohyla calypsa</i>	34	Duellman 2001	Hylini
<i>Isthmohyla debilis</i>	29.5	Duellman 2001	Hylini
<i>Isthmohyla graceae</i>	38.7	Duellman 2001	Hylini
<i>Isthmohyla infucata</i>	42.9	Duellman 2001	Hylini
<i>Isthmohyla insolita</i>	36	Duellman 2001	Hylini
<i>Isthmohyla lancasteri</i>	33.6	Duellman 2001	Hylini
<i>Isthmohyla picadoi</i>	32.8	Duellman 2001	Hylini

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<i>Isthmohyla pictipes</i>	39	Duellman 2001	Hylini
<i>Isthmohyla pseudopuma</i>	41.4	Duellman 2001	Hylini
<i>Isthmohyla rivularis</i>	34	Duellman 2001	Hylini
<i>Isthmohyla tica</i>	34.1	Duellman 2001	Hylini
<i>Isthmohyla zeteki</i>	23.5	Duellman 2001	Hylini
<i>Megastomatohyla mixe</i>	30.8	Duellman 2001	Hylini
<i>Megastomatohyla</i>			
<i>mixomaculata</i>	29.1	Duellman 2001	Hylini
<i>Megastomatohyla nubicola</i>	36.7	Duellman 2001	Hylini
<i>Megastomatohyla pellita</i>	29	Duellman 2001	Hylini
<i>Plectrohyla aconthodes</i>	63.2	Duellman 2001	Hylini
		Canseco-Márquez	
<i>Plectrohyla ameibothalame</i>	43.1	et al. 2002	Hylini
<i>Plectrohyla arborescandens</i>	37.6	Duellman 2001	Hylini
<i>Plectrohyla avia</i>	90.4	Duellman 2001	Hylini
<i>Plectrohyla bistincta</i>	53.8	Duellman 2001	Hylini
<i>Plectrohyla calthula</i>	56.1	Duellman 2001	Hylini
<i>Plectrohyla calvicollina</i>	37.6	Duellman 2001	Hylini
<i>Plectrohyla celata</i>	46.2	Duellman 2001	Hylini
<i>Plectrohyla cembra</i>	37	Duellman 2001	Hylini
<i>Plectrohyla charadricola</i>	44.4	Duellman 2001	Hylini

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<i>Plectrohyla chryses</i>	41.1	Duellman 2001	Hylini
<i>Plectrohyla chrysopleura</i>	65.6	Duellman 2001	Hylini
<i>Plectrohyla crassa</i>	59.6	Duellman 2001	Hylini
<i>Plectrohyla cyanomma</i>	64.5	Duellman 2001	Hylini
<i>Plectrohyla cyclada</i>	39.5	Duellman 2001	Hylini
<i>Plectrohyla dasypus</i>	44	Duellman 2001	Hylini
<i>Plectrohyla exquisita</i>	80.7	Duellman 2001	Hylini
<i>Plectrohyla glandulosa</i>	49.1	Duellman 2001	Hylini
<i>Plectrohyla guatemalensis</i>	59.5	Duellman 2001	Hylini
<i>Plectrohyla hartwegi</i>	75.8	Duellman 2001	Hylini
<i>Plectrohyla hazelae</i>	38.6	Duellman 2001	Hylini
<i>Plectrohyla ixil</i>	41.6	Duellman 2001	Hylini
<i>Plectrohyla lacertosa</i>	47.8	Duellman 2001	Hylini
<i>Plectrohyla matudai</i>	46	Duellman 2001	Hylini
<i>Plectrohyla mykter</i>	42.3	Duellman 2001	Hylini
<i>Plectrohyla pachyderma</i>	39.9	Duellman 2001	Hylini
<i>Plectrohyla pentheter</i>	52.1	Duellman 2001	Hylini
<i>Plectrohyla pokomchi</i>	55.2	Duellman 2001	Hylini
<i>Plectrohyla psarosema</i>	31.4	Duellman 2001	Hylini
<i>Plectrohyla psiloderma</i>	48.5	Duellman 2001	Hylini
<i>Plectrohyla pycnochila</i>	60.5	Duellman 2001	Hylini

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<i>Plectrohyla quecchi</i>	43.8	Duellman 2001	Hylini
<i>Plectrohyla robertsororum</i>	47.9	Duellman 2001	Hylini
<i>Plectrohyla sabrina</i>	30.2	Duellman 2001	Hylini
<i>Plectrohyla sagorum</i>	45.5	Duellman 2001	Hylini
<i>Plectrohyla siopela</i>	46.2	Duellman 2001	Hylini
<i>Plectrohyla tecunumani</i>	61.6	Duellman 2001	Hylini
<i>Plectrohyla teuchestes</i>	76.1	Duellman 2001	Hylini
<i>Plectrohyla thorectes</i>	34.2	Duellman 2001	Hylini
		Wright and Wright	
<i>Pseudacris brachyphona</i>	32	1949	Hylini
		Wright and Wright	
<i>Pseudacris brimleyi</i>	28	1949	Hylini
<i>Pseudacris cadaverina</i>	35.9	Duellman 2001	Hylini
<i>Pseudacris clarkii</i>	29	Duellman 2001	Hylini
		Wright and Wright	
<i>Pseudacris crucifer</i>	29	1949	Hylini
		Wright and Wright	
<i>Pseudacris nigrita</i>	28	1949	Hylini
		Wright and Wright	
<i>Pseudacris ocularis</i>	15.5	1949	Hylini
<i>Pseudacris ornata</i>	39	Lannoo 2005	Hylini

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		Wright and Wright	
<i>Pseudacris regilla</i>	48	1949	Hylini
		Wright and Wright	
<i>Pseudacris streckeri</i>	41	1949	Hylini
		Wright and Wright	
<i>Pseudacris triseriata</i>	32	1949	Hylini
<i>Ptychohyla acrochorda</i>	36.3	Duellman 2001	Hylini
<i>Ptychohyla euthysanota</i>	37.3	Duellman 2001	Hylini
<i>Ptychohyla hypomyketer</i>	35.4	Duellman 2001	Hylini
<i>Ptychohyla legleri</i>	36.7	Duellman 2001	Hylini
<i>Ptychohyla leonhardschultzei</i>	35.1	Duellman 2001	Hylini
<i>Ptychohyla macrotympanum</i>	38.8	Duellman 2001	Hylini
<i>Ptychohyla panchoi</i>	34.2	Duellman 2001	Hylini
<i>Ptychohyla salvadorensis</i>	34.2	Duellman 2001	Hylini
<i>Ptychohyla sanctaecrucis</i>	32.5	Duellman 2001	Hylini
<i>Ptychohyla spinipollex</i>	39.1	Duellman 2001	Hylini
<i>Ptychohyla zophodes</i>	37.4	Duellman 2001	Hylini
<i>Smilisca baudinii</i>	75.9	Duellman 2001	Hylini
<i>Smilisca cyanosticta</i>	57.8	Duellman 2001	Hylini
<i>Smilisca dentata</i>	62.1	Duellman 2001	Hylini
<i>Smilisca fodiens</i>	62.6	Duellman 2001	Hylini

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<i>Smilisca phaeto</i>	65.5	Duellman 2001	Hylini
<i>Smilisca puma</i>	38.1	Duellman 2001	Hylini
<i>Smilisca sila</i>	44.8	Duellman 2001	Hylini
<i>Smilisca sordida</i>	44.6	Duellman 2001	Hylini
<i>Tlalocohyla godmani</i>	38	Duellman 2001	Hylini
<i>Tlalocohyla loquax</i>	44.7	Duellman 2001	Hylini
<i>Tlalocohyla picta</i>	21.4	Duellman 2001	Hylini
<i>Tlalocohyla smithii</i>	26	Duellman 2001	Hylini
<i>Tripurion petasatus</i>	60.8	Duellman 2001	Hylini
<i>Tripurion spatulatus</i>	85.9	Duellman 2001	Hylini
<i>Aparasphenodon bokermanni</i>	71.1	Pombal 1993	Lophiohylini
<i>Aparasphenodon venezolanus</i>	58	Rivero 1961	Lophiohylini
<i>Argenteohyla siemersi</i>	70	Cei 1980	Lophiohylini
<i>Corythomantis greeningi</i>	73	Jared et al. 1999	Lophiohylini
		personal	
<i>Itapotihyla langsdorffii</i>	77	measurements	Lophiohylini
		Duellman and	
<i>Nyctimantis rugiceps</i>	67.5	Trueb 1976	Lophiohylini
<i>Osteocephalus alboguttatus</i>	34	Duellman 1978	Lophiohylini
		Cochran and Goin	
<i>Osteocephalus buckleyi</i>	64.1	1970	Lophiohylini

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<i>Osteocephalus deridens</i>	34.9	Jungfer et al. 2000	Lophiohylini
<i>Osteocephalus</i>			
<i>elkejungingerae</i>	22	Henle 1981 Smith and Noonan	Lophiohylini
<i>Osteocephalus exophthalmus</i>	32.7	2001	Lophiohylini
<i>Osteocephalus fuscifacies</i>	45.6	Jungfer et al. 2000	Lophiohylini
<i>Osteocephalus heyeri</i>	36.1	Lynch 2002 Jungfer and Lehr	Lophiohylini
<i>Osteocephalus leoniae</i>	40.1	2001 Rodríguez and	Lophiohylini
<i>Osteocephalus leprieurii</i>	48	Duellman 1994 Jungfer and Hodl	Lophiohylini
<i>Osteocephalus mutabor</i>	50.3	2003 Jungfer and	Lophiohylini
<i>Osteocephalus oophagus</i>	47.2	Schiesari 1995 Trueb and	Lophiohylini
<i>Osteocephalus pearsoni</i>	46.2	Duellman 1971 Duellman and	Lophiohylini
<i>Osteocephalus planiceps</i>	65.9	Mendelson 1995 Martins and	Lophiohylini
<i>Osteocephalus subtilis</i>	38.8	Cardoso 1987	Lophiohylini

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		Trueb and	
<i>Osteocephalus taurinus</i>	85	Duellman 1971 personal	Lophiohylini
<i>Osteocephalus verruciger</i>	58.6	measurements Ron and Pramuk	Lophiohylini
<i>Osteocephalus yasuni</i>	55.7	1999 <a href="http://evo.bio.psu.edu/caribherp/">http://evo.bio.psu.edu/caribherp/</a>	Lophiohylini
<i>Osteopilus brunneus</i>	52	(male) <a href="http://evo.bio.psu.edu/caribherp/">http://evo.bio.psu.edu/caribherp/</a>	Lophiohylini
<i>Osteopilus crucialis</i>	100	(male) <a href="http://evo.bio.psu.edu/caribherp/">http://evo.bio.psu.edu/caribherp/</a>	Lophiohylini
<i>Osteopilus dominicensis</i>	66	(male) <a href="http://evo.bio.psu.edu/caribherp/">http://evo.bio.psu.edu/caribherp/</a>	Lophiohylini
<i>Osteopilus marianae</i>	40	(male) Trueb and Tyler	Lophiohylini
<i>Osteopilus pulchrilineatus</i>	39.5	1974	Lophiohylini
<i>Osteopilus septentrionalis</i>	89	<a href="http://evo.bio.psu.edu/caribherp/">http://evo.bio.psu.edu/caribherp/</a>	Lophiohylini

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		.edu/caribherp/	
		(male)-	
		Trueb and Tyler	
<i>Osteopilus vastus</i>	108.8	1974	Lophiohylini
		Trueb and Tyler	
<i>Osteopilus wilderi</i>	27.3	1974	Lophiohylini
<i>Phyllodytes acuminatus</i>	24.5	Bokermann 1966	Lophiohylini
<i>Phyllodytes auratus</i>	29	Murphy 1997	Lophiohylini
<i>Phyllodytes edelmoi</i>	28.7	Peixoto et al. 2003	Lophiohylini
<i>Phyllodytes gyrinaethes</i>	27.9	Peixoto et al. 2003	Lophiohylini
<i>Phyllodytes luteolus</i>	23	Bokermann 1966	Lophiohylini
		Caramaschi et al.	
<i>Phyllodytes melanomystax</i>	25.4	1992	Lophiohylini
		Caramaschi and	
<i>Phyllodytes punctatus</i>	22.8	Peixoto 2004	Lophiohylini
<i>Phyllodytes tuberculosus</i>	24	Bokermann 1966	Lophiohylini
		Caramaschi et al.	
<i>Phyllodytes wuchereri</i>	26	2004	Lophiohylini
		Mijares-Urrutia et	
<i>Tepuihyla aecii</i>	34	al. 1999	Lophiohylini
<i>Tepuihyla celsae</i>	46.5	Mijares-Urrutia et	Lophiohylini

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		al. 1999	
		Mijares-Urrutia et	
<i>Tepuihyla edelcae</i>	40.2	al. 1999	Lophiohylini
		Mijares-Urrutia et	
<i>Tepuihyla galani</i>	40	al. 1999	Lophiohylini
		Ayarzaguena et al.	
<i>Tepuihyla luteolabris</i>	42.8	1992	Lophiohylini
		Mijares-Urrutia et	
<i>Tepuihyla rimarum</i>	31.8	al. 1999	Lophiohylini
		Duellman and	
<i>Tepuihyla rodriguezi</i>	34.7	Hoogmoed 1992	Lophiohylini
		Duellman and	
<i>Tepuihyla talbergae</i>	32.7	Yoshpa 1996	Lophiohylini
<i>Trachycephalus atlas</i>	98	Bokermann 1966	Lophiohylini
		Rodríguez and	
<i>Trachycephalus coriaceus</i>	63	Duellman 1994	Lophiohylini
		Lescure and Marty	
<i>Trachycephalus hadroceps</i>	60	2000	Lophiohylini
<i>Trachycephalus imitatrix</i>	57.1	Lutz 1973	Lophiohylini
		personal	
<i>Trachycephalus jordani</i>	75.9	measurements	Lophiohylini

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Pombal et al. 2003			
<i>Trachycephalus lepidus</i>	49.1	Copeia	Lophiohylini
<i>Trachycephalus mesophaeus</i>	85	Lutz 1973	Lophiohylini
<i>Trachycephalus</i>			
<i>nigromaculatus</i>	86	Cochran 1955	Lophiohylini
		Rodríguez and	
<i>Trachycephalus resinifictrix</i>	76	Duellman 1994	Lophiohylini
<i>Trachycephalus venulosus</i>	100.5	Duellman 2001	Lophiohylini
<i>Lysapsus caraya</i>	16.5	Gallardo 1964	<i>Pseudis</i> clade
<i>Lysapsus laevis</i>	21	Parker 1935	<i>Pseudis</i> clade
<i>Lysapsus limellum</i>	20	Gallardo 1964	<i>Pseudis</i> clade
		Caramaschi and	
<i>Pseudis bolbodactyla</i>	45	Cruz 1998	<i>Pseudis</i> clade
<i>Pseudis cardosoi</i>	45.9	Kwet 2000	<i>Pseudis</i> clade
		Caramaschi and	
<i>Pseudis fusca</i>	40.8	Cruz 1998	<i>Pseudis</i> clade
<i>Pseudis minuta</i>	40	Cei 1980	<i>Pseudis</i> clade
<i>Pseudis paradoxa</i>	55	Gallardo 1964	<i>Pseudis</i> clade
<i>Scarthylla goinorum</i>	21	Duellman 2005	<i>Pseudis</i> clade
<i>Scarthylla vigilans</i>	17.4	Solano 1971	<i>Pseudis</i> clade
<i>Scinax acuminatus</i>	45	Lutz 1973	<i>Scinax</i> clade

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<i>Scinax agilis</i>	19.5	Faivovich 2005	<i>Scinax</i> clade
<i>Scinax albicans</i>	31.2	Faivovich 2005	<i>Scinax</i> clade
		Brasileiro et al.	
<i>Scinax alcatraz</i>	24.4	2007	<i>Scinax</i> clade
<i>Scinax altae</i>	26	León 1969	<i>Scinax</i> clade
<i>Scinax alter</i>	30	Lutz 1973	<i>Scinax</i> clade
<i>Scinax angrensis</i>	32	Lutz 1973	<i>Scinax</i> clade
<i>Scinax arduous</i>	19.5	Peixoto 2002	<i>Scinax</i> clade
<i>Scinax argyreornatus</i>	15.8	Faivovich 2005	<i>Scinax</i> clade
<i>Scinax ariadne</i>	36.5	Lutz 1973	<i>Scinax</i> clade
<i>Scinax aromothyella</i>	24.8	Faivovich 2005	<i>Scinax</i> clade
<i>Scinax atratus</i>	19.2	Peixoto 1988	<i>Scinax</i> clade
<i>Scinax auratus</i>	23	Lutz 1973	<i>Scinax</i> clade
<i>Scinax baumgardneri</i>	29	Rivero 1961	<i>Scinax</i> clade
<i>Scinax berthae</i>	22.2	Faivovich 2005	<i>Scinax</i> clade
		Fouquette and	
<i>Scinax blairi</i>	30.1	Pyburn 1972	<i>Scinax</i> clade
<i>Scinax boesemani</i>	31.1	Duellman 1997	<i>Scinax</i> clade
<i>Scinax boulengeri</i>	48.7	Duellman 2001	<i>Scinax</i> clade
<i>Scinax brienii</i>	32.7	Heyer et al. 1990	<i>Scinax</i> clade
<i>Scinax caldarum</i>	25	Lutz 1973	<i>Scinax</i> clade

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		Caramaschi and	
<i>Scinax camposseabrai</i>	33.5	Cardoso 2006	<i>Scinax</i> clade
		Cardoso and	
<i>Scinax canastrensis</i>	28	Haddad 1982	<i>Scinax</i> clade
		Caramaschi and	
<i>Scinax carnevallii</i>	25	Kisteumacher 1989	<i>Scinax</i> clade
<i>Scinax castroviejoii</i>	45	De la Riva 1993	<i>Scinax</i> clade
<i>Scinax catharinae</i>	35.1	Faivovich 2005	<i>Scinax</i> clade
		Pombal and Bastos	
<i>Scinax centralis</i>	21.2	1996	<i>Scinax</i> clade
<i>Scinax chiquitanus</i>	33.3	Duellman 2005	<i>Scinax</i> clade
		Lima, Bastos, and	
<i>Scinax constrictus</i>	29.4	Giaretta 2004	<i>Scinax</i> clade
<i>Scinax crospedospilus</i>	33.3	Heyer et al. 1990	<i>Scinax</i> clade
<i>Scinax cruentommus</i>	28	Duellman 1978	<i>Scinax</i> clade
<i>Scinax curicica</i>	30.2	Pugliese et al. 2004	<i>Scinax</i> clade
<i>Scinax cuspidatus</i>	29	Lutz 1973	<i>Scinax</i> clade
<i>Scinax danae</i>	27.4	Duellman 1986	<i>Scinax</i> clade
<i>Scinax duartei</i>	35	Lutz 1973	<i>Scinax</i> clade
<i>Scinax elaeochrous</i>	37.7	Duellman 2001	<i>Scinax</i> clade
<i>Scinax eurydice</i>	42	Lutz 1973	<i>Scinax</i> clade

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<i>Scinax exiguus</i>	20.8	Duellman 1986	<i>Scinax</i> clade
<i>Scinax flavidus</i>	30.5	La Marca 2004	<i>Scinax</i> clade
<i>Scinax flavoguttatus</i>	29.3	Heyer et al. 1990	<i>Scinax</i> clade
		Rodriguez and	
<i>Scinax funereus</i>	37	Duellman 1994	<i>Scinax</i> clade
<i>Scinax fuscomarginatus</i>	24	Cochran 1955	<i>Scinax</i> clade
<i>Scinax fuscovarius</i>	>47.1	De la Riva 1993	<i>Scinax</i> clade
		Duellman and	
<i>Scinax garbei</i>	42.2	Wiens 1993	<i>Scinax</i> clade
<i>Scinax granulatus</i>	36	Lutz 1973	<i>Scinax</i> clade
<i>Scinax hayii</i>	43	Heyer et al. 1990	<i>Scinax</i> clade
		Peixoto and	
<i>Scinax heyeri</i>	35.6	Weygoldt 1987	<i>Scinax</i> clade
		Haddad and	
<i>Scinax hiemalis</i>	28.2	Pombal 1987	<i>Scinax</i> clade
<i>Scinax humilis</i>	28	Lutz 1973	<i>Scinax</i> clade
<i>Scinax ictericus</i>	31.8	Duellman 2005	<i>Scinax</i> clade
		Lescure and Marty	
<i>Scinax jolyi</i>	39.1	2000	<i>Scinax</i> clade
		Pombal and Gordo	
<i>Scinax jureia</i>	30	1991	<i>Scinax</i> clade

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<i>Scinax karenanneae</i>	28.9	Pyburn 1993	<i>Scinax</i> clade
<i>Scinax kennedyi</i>	35.7	Pyburn 1973	<i>Scinax</i> clade
<i>Scinax lindsayi</i>	25.4	Pyburn 1992	<i>Scinax</i> clade
		Pombal and Gordo	
<i>Scinax littoralis</i>	29.6	1991	<i>Scinax</i> clade
<i>Scinax littoreus</i>	19.3	Peixoto 1988	<i>Scinax</i> clade
		Caramaschi and	
<i>Scinax luizotavioi</i>	24.3	Kisteumacher 1989	<i>Scinax</i> clade
		Bokermann and	
<i>Scinax machadoi</i>	21	Sazima 1973	<i>Scinax</i> clade
		Barrio-Amoros et	
<i>Scinax manriquei</i>	30	al. 2004	<i>Scinax</i> clade
		Cardoso and	
<i>Scinax maracaya</i>	28	Sazima 1980	<i>Scinax</i> clade
<i>Scinax melloi</i>	17	Peixoto 1988	<i>Scinax</i> clade
<i>Scinax nasicus</i>	37	Lutz 1973	<i>Scinax</i> clade
<i>Scinax nebulosus</i>	30	Pyburn 1973	<i>Scinax</i> clade
<i>Scinax obtriangulatus</i>	28	Lutz 1973	<i>Scinax</i> clade
		Duellman and	
<i>Scinax oreites</i>	33.5	Wiens 1993	<i>Scinax</i> clade
<i>Scinax pachycrus</i>	33	Lutz 1973	<i>Scinax</i> clade

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<i>Scinax parkeri</i>	23.9	Duellman 1986	<i>Scinax</i> clade
<i>Scinax pedromedinae</i>	29.4	Duellman 2005	<i>Scinax</i> clade
<i>Scinax perereca</i>	38.5	Pombal et al. 1995	<i>Scinax</i> clade
<i>Scinax perpusillus</i>	18.3	Heyer et al. 1990	<i>Scinax</i> clade
<i>Scinax pinima</i>	25	Bokermann and Sazima 1973 Lima, Bastos, and	<i>Scinax</i> clade
<i>Scinax proboscideus</i>	39.8	Giaretta 2004	<i>Scinax</i> clade
<i>Scinax quinquefasciatus</i>	30	Fowler 1913 Andrade and	<i>Scinax</i> clade
<i>Scinax ranki</i>	23.3	Cardoso 1987	<i>Scinax</i> clade
<i>Scinax rizibilis</i>	27	Lutz 1973	<i>Scinax</i> clade
<i>Scinax rostratus</i>	45.7	Duellman 2001 Duellman and	<i>Scinax</i> clade
<i>Scinax ruber</i>	41.2	Wiens 1993	<i>Scinax</i> clade
<i>Scinax similis</i>	41	Lutz 1973	<i>Scinax</i> clade
<i>Scinax squalirostris</i>	29	Lutz 1973	<i>Scinax</i> clade
<i>Scinax staufferi</i>	29	Duellman 2001	<i>Scinax</i> clade
<i>Scinax strigilatus</i>	28	<b>Cochran 1959</b>	<i>Scinax</i> clade
<i>Scinax sugillatus</i>	42	Duellman 1973	<i>Scinax</i> clade
<i>Scinax trapicheiroi</i>	30	Lutz 1973	<i>Scinax</i> clade

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		Hoogmoed and	
<i>Scinax trilineatus</i>	22.5	Gorzula 1979	<i>Scinax</i> clade
<i>Scinax uruguayus</i>	25.8	Langone 1990	<i>Scinax</i> clade
		Pyburn and	
<i>Scinax wandae</i>	26.9	Fouquette 1971	<i>Scinax</i> clade
<i>Scinax x-signatus</i>	42.5	Heyer et al. 1990	<i>Scinax</i> clade
		Brasileiro et al.	
<i>Scinax peixotoi</i>	20.7	2007	<i>Scinax</i> clade
<i>Sphaenorhynchus bromelicola</i>	30	Bokermann 1966	<i>Scinax</i> clade
<i>Sphaenorhynchus carneus</i>	20	Duellman 1978	<i>Scinax</i> clade
		Rodríguez and	
<i>Sphaenorhynchus dorisae</i>	29	Duellman 1994	<i>Scinax</i> clade
<i>Sphaenorhynchus lacteus</i>	41.5	Duellman 2005	<i>Scinax</i> clade
<i>Sphaenorhynchus orophilus</i>	32	Heyer et al. 1990	<i>Scinax</i> clade
<i>Sphaenorhynchus palustris</i>	36	Bokermann 1966	<i>Scinax</i> clade
<i>Sphaenorhynchus pauloalvini</i>	20	Bokermann 1973	<i>Scinax</i> clade
<i>Sphaenorhynchus planicola</i>	24	Cochran 1955	<i>Scinax</i> clade
<i>Sphaenorhynchus</i>			
<i>platycephalus</i>	33	Harding 1991	<i>Scinax</i> clade
<i>Sphaenorhynchus prasinus</i>	31	Bokermann 1973	<i>Scinax</i> clade
<i>Sphaenorhynchus surdus</i>	28	Cochran 1953	<i>Scinax</i> clade

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<i>Cyclorana australis</i>	79	Barker et al. 1995	Pelodryadinae
<i>Cyclorana brevipes</i>	45	Barker et al. 1995	Pelodryadinae
<i>Cyclorana cryptotis</i>	44	Barker et al. 1995	Pelodryadinae
<i>Cyclorana cultripes</i>	41	Barker et al. 1995	Pelodryadinae
<i>Cyclorana longipes</i>	46	Barker et al. 1995	Pelodryadinae
<i>Cyclorana maculosus</i>	55	Barker et al. 1995	Pelodryadinae
<i>Cyclorana maini</i>	46	Barker et al. 1995	Pelodryadinae
<i>Cyclorana manya</i>	30	Barker et al. 1995	Pelodryadinae
<i>Cyclorana novaehollandiae</i>	81	Barker et al. 1995	Pelodryadinae
<i>Cyclorana platycephala</i>	64	Barker et al. 1995	Pelodryadinae
<i>Cyclorana vagitus</i>	48	Barker et al. 1995	Pelodryadinae
<i>Cyclorana verrucosus</i>	45	Barker et al. 1995	Pelodryadinae
<i>Litoria adelaidensis</i>	45	Barker et al. 1995	Pelodryadinae
<i>Litoria alboguttata</i>	67	Barker et al. 1995	Pelodryadinae
<i>Litoria amboiensis</i>	54	Menzies 2006	Pelodryadinae
<i>Litoria angiana</i>	65	Menzies 2006	Pelodryadinae
<i>Litoria arfakiana</i>	45	Menzies 2006	Pelodryadinae
<i>Litoria auae</i>	32	Menzies 2006	Pelodryadinae
<i>Litoria aurea</i>	69	Barker et al. 1995	Pelodryadinae
<i>Litoria becki</i>	38	Menzies 2006	Pelodryadinae
<i>Litoria bibonius</i>	27	Menzies 2006	Pelodryadinae

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<i>Litoria bicolor</i>	27	Barker et al. 1995	Pelodryadinae
<i>Litoria booroolongensis</i>	42	Barker et al. 1995	Pelodryadinae
<i>Litoria brevipalmata</i>	43	Barker et al. 1995	Pelodryadinae
<i>Litoria brongersmai</i>	24	Menzies 2006	Pelodryadinae
<i>Litoria bulmeri</i>	34	Menzies 2006	Pelodryadinae
<i>Litoria burrowsae</i>	53	Barker et al. 1995	Pelodryadinae
<i>Litoria caerulea</i>	80	Barker et al. 1995	Pelodryadinae
<i>Litoria capitula</i>	34	Menzies 2006	Pelodryadinae
<i>Litoria cavernicola</i>	51	Barker et al. 1995	Pelodryadinae
<i>Litoria chloris</i>	62	Barker et al. 1995	Pelodryadinae
<i>Litoria chloronata</i>	30	Menzies 2006	Pelodryadinae
<i>Litoria citropa</i>	57	Barker et al. 1995	Pelodryadinae
<i>Litoria congenita</i>	35	Menzies 2006	Pelodryadinae
<i>Litoria contrastens</i>	30	Menzies 2006	Pelodryadinae
<i>Litoria cooloolensis</i>	26	Barker et al. 1995	Pelodryadinae
<i>Litoria coplandi</i>	36	Barker et al. 1995	Pelodryadinae
<i>Litoria cyclorhyncha</i>	66	Barker et al. 1995	Pelodryadinae
<i>Litoria dahlii</i>	63	Barker et al. 1995	Pelodryadinae
<i>Litoria darlingtoni</i>	50	Menzies 2006	Pelodryadinae
<i>Litoria dentata</i>	40	Barker et al. 1995	Pelodryadinae
<i>Litoria dorsalis</i>	24	Menzies 2006	Pelodryadinae

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<i>Litoria dorsivena</i>	30	Menzies 2006	Pelodryadinae
<i>Litoria electrica</i>	38	Barker et al. 1995	Pelodryadinae
<i>Litoria elkeae</i>	29	Menzies 2006	Pelodryadinae
<i>Litoria eucnemis</i>	48	Barker et al. 1995	Pelodryadinae
<i>Litoria ewingii</i>	40	Barker et al. 1995	Pelodryadinae
<i>Litoria exophthalmia</i>	39	Menzies 2006	Pelodryadinae
<i>Litoria fallax</i>	26	Barker et al. 1995	Pelodryadinae
<i>Litoria flavescens</i>	50	Menzies 2006	Pelodryadinae
<i>Litoria flavipunctata</i>	73	Barker et al. 1995	Pelodryadinae
<i>Litoria freycineti</i>	39	Barker et al. 1995	Pelodryadinae
<i>Litoria genimaculata</i>	41	Barker et al. 1995	Pelodryadinae
<i>Litoria gilleni</i>	62	Barker et al. 1995	Pelodryadinae
<i>Litoria gracilentata</i>	42	Barker et al. 1995	Pelodryadinae
<i>Litoria graminea</i>	70	Menzies 2006	Pelodryadinae
<i>Litoria havina</i>	37	Menzies 2006	Pelodryadinae
<i>Litoria impura</i>	50	Menzies 2006	Pelodryadinae
<i>Litoria inermis</i>	33	Barker et al. 1995	Pelodryadinae
<i>Litoria infrafronata</i>	102	Barker et al. 1995	Pelodryadinae
<i>Litoria iris</i>	36	Menzies 2006	Pelodryadinae
<i>Litoria jervisiensis</i>	37	Barker et al. 1995	Pelodryadinae
<i>Litoria jeudii</i>	33	Menzies 2006	Pelodryadinae

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<i>Litoria kumae</i>	27	Menzies 2006	Pelodryadinae
<i>Litoria latopalmata</i>	39	Barker et al. 1995	Pelodryadinae
<i>Litoria lesueuri</i>	43	Barker et al. 1995	Pelodryadinae
<i>Litoria leucova</i>	35	Menzies 2006	Pelodryadinae
<i>Litoria littlejohni</i>	56	Barker et al. 1995	Pelodryadinae
<i>Litoria longicrus</i>	27	Menzies 2006	Pelodryadinae
<i>Litoria longirostris</i>	27	Barker et al. 1995	Pelodryadinae
<i>Litoria lorica</i>	32	Barker et al. 1995	Pelodryadinae
<i>Litoria louisianensis</i>	29	Menzies 2006	Pelodryadinae
<i>Litoria lutea</i>	65	Menzies 2006	Pelodryadinae
<i>Litoria macki</i>	46	Menzies 2006	Pelodryadinae
<i>Litoria majikthise</i>	35	Menzies 2006	Pelodryadinae
<i>Litoria meiriana</i>	20	Barker et al. 1995	Pelodryadinae
<i>Litoria michaeltyleri</i>	78	Menzies 2006	Pelodryadinae
<i>Litoria microbelos</i>	16	Barker et al. 1995	Pelodryadinae
<i>Litoria micromembrana</i>	40	Menzies 2006	Pelodryadinae
<i>Litoria modica</i>	30	Menzies 2006	Pelodryadinae
<i>Litoria moorei</i>	64	Barker et al. 1995	Pelodryadinae
<i>Litoria mucro</i>	31	Menzies 2006	Pelodryadinae
<i>Litoria multicolor</i>	55	Menzies 2006	Pelodryadinae
<i>Litoria multiplica</i>	42	Menzies 2006	Pelodryadinae

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<i>Litoria mystax</i>	30	Menzies 2006	Pelodryadinae
<i>Litoria nannotis</i>	48	Barker et al. 1995	Pelodryadinae
<i>Litoria napaea</i>	23	Menzies 2006	Pelodryadinae
<i>Litoria nasuta</i>	45	Menzies 2006	Pelodryadinae
<i>Litoria nigrofrenata</i>	42	Barker et al. 1995	Pelodryadinae
<i>Litoria nigropunctata</i>	32	Menzies 1977	Pelodryadinae
<i>Litoria nyakalensis</i>	33	Barker et al. 1995	Pelodryadinae
<i>Litoria oenicolon</i>	50	Menzies 1977	Pelodryadinae
<i>Litoria ollauro</i>	34	Menzies 2006	Pelodryadinae
<i>Litoria olongburensis</i>	25	Barker et al. 1995	Pelodryadinae
<i>Litoria pallida</i>	34	Barker et al. 1995	Pelodryadinae
<i>Litoria paraewingi</i>	28	Barker et al. 1995	Pelodryadinae
<i>Litoria pearsoniana</i>	29	Barker et al. 1995	Pelodryadinae
<i>Litoria peronii</i>	53	Barker et al. 1995	Pelodryadinae
<i>Litoria personata</i>	29	Barker et al. 1995	Pelodryadinae
<i>Litoria phyllochroa</i>	32	Barker et al. 1995	Pelodryadinae
<i>Litoria piperata</i>	27	Barker et al. 1995	Pelodryadinae
<i>Litoria pratti</i>	31	Menzies 2006	Pelodryadinae
<i>Litoria pronimia</i>	34	Menzies 2006	Pelodryadinae
<i>Litoria prora</i>	42	Menzies 2006	Pelodryadinae
<i>Litoria pygmaea</i>	30	Menzies 2006	Pelodryadinae

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<i>Litoria quadrilineata</i>	30	Menzies 2006	Pelodryadinae
<i>Litoria raniformis</i>	65	Barker et al. 1995	Pelodryadinae
<i>Litoria revelata</i>	28	Barker et al. 1995	Pelodryadinae
<i>Litoria rheocola</i>	32	Barker et al. 1995	Pelodryadinae
<i>Litoria rothii</i>	48	Barker et al. 1995	Pelodryadinae
<i>Litoria rubella</i>	37	Barker et al. 1995	Pelodryadinae
<i>Litoria rubrops</i>	25	Menzies 2006	Pelodryadinae
<i>Litoria sanguinolenta</i>	40	Menzies 2006	Pelodryadinae
<i>Litoria spenceri</i>	41	Barker et al. 1995	Pelodryadinae
<i>Litoria spinifera</i>	42	Menzies 2006	Pelodryadinae
<i>Litoria splendida</i>	104	Barker et al. 1995	Pelodryadinae
<i>Litoria subglandulosa</i>	40	Barker et al. 1995	Pelodryadinae
<i>Litoria thesaurensis</i>	50	Menzies 2006	Pelodryadinae
<i>Litoria timida</i>	24	Menzies 2006	Pelodryadinae
<i>Litoria tornieri</i>	36	Barker et al. 1995	Pelodryadinae
<i>Litoria tyleri</i>	48	Barker et al. 1995	Pelodryadinae
<i>Litoria umarensis</i>	30	Menzies 2006	Pelodryadinae
<i>Litoria umbonata</i>	36	Menzies 2006	Pelodryadinae
<i>Litoria verae</i>	35	Menzies 2006	Pelodryadinae
<i>Litoria verreauxii</i>	36	Barker et al. 1995	Pelodryadinae
<i>Litoria vociovincens</i>	27	Menzies 2006	Pelodryadinae

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<i>Litoria watjulumensis</i>	38	Barker et al. 1995	Pelodryadinae
<i>Litoria wisselensis</i>	32	Menzies 2006	Pelodryadinae
<i>Litoria wollastoni</i>	50	Menzies 1977	Pelodryadinae
<i>Litoria xanthomera</i>	56	Barker et al. 1995	Pelodryadinae
<i>Nyctimystes avocalis</i>	35	Menzies 2006	Pelodryadinae
<i>Nyctimystes cheesemanae</i>	60	Menzies 2006	Pelodryadinae
<i>Nyctimystes dayi</i>	42	Barker et al. 1995	Pelodryadinae
<i>Nyctimystes daymani</i>	60	Menzies 2006	Pelodryadinae
<i>Nyctimystes disrupta</i>	60	Menzies 2006	Pelodryadinae
<i>Nyctimystes foricula</i>	40	Menzies 2006	Pelodryadinae
<i>Nyctimystes granti</i>	80	Menzies 2006	Pelodryadinae
<i>Nyctimystes gularis</i>	37	Menzies 2006	Pelodryadinae
<i>Nyctimystes humeralis</i>	100	Menzies 2006	Pelodryadinae
<i>Nyctimystes kubori</i>	45	Menzies 2006	Pelodryadinae
<i>Nyctimystes narinosa</i>	59	Menzies 2006	Pelodryadinae
<i>Nyctimystes obsoleta</i>	35	Menzies 2006	Pelodryadinae
<i>Nyctimystes oktediensis</i>	67	Menzies 2006	Pelodryadinae
<i>Nyctimystes papua</i>	67	Menzies 2006	Pelodryadinae
<i>Nyctimystes perimetri</i>	52	Menzies 2006	Pelodryadinae
<i>Nyctimystes persimilis</i>	40	Menzies 2006	Pelodryadinae
<i>Nyctimystes pulchra</i>	60	Menzies 2006	Pelodryadinae

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<i>Nyctimystes semipalmata</i>	65	Menzies 2006	Pelodryadinae
<i>Nyctimystes trachydermis</i>	46	Menzies 2006	Pelodryadinae
<i>Nyctimystes zweifeli</i>	80	Menzies 2006	Pelodryadinae
<i>Agalychnis annae</i>	73.9	Duellman 2001	Phyllomedusinae
<i>Agalychnis callidryas</i>	57.2	Duellman 2001	Phyllomedusinae
<i>Agalychnis litodryas</i>	70.2	Duellman 2001	Phyllomedusinae
<i>Agalychnis moreletii</i>	65.7	Duellman 2001	Phyllomedusinae
<i>Agalychnis saltator</i>	46.7	Duellman 2001	Phyllomedusinae
<i>Agalychnis spurrelli</i>	75.6	Duellman 2001	Phyllomedusinae
<i>Cruziophyla calcarifer</i>	64	Duellman 2001	Phyllomedusinae
<i>Cruziophyla craspedopus</i>	57	Duellman 2005	Phyllomedusinae
<i>Hylomantis aspera</i>	41.7	Cruz 1988	Phyllomedusinae
<i>Hylomantis buckleyi</i>	44.5	Cannatella 1980	Phyllomedusinae
		Ruiz-Carranza et	
<i>Hylomantis danieli</i>	80.8	al. 1988	Phyllomedusinae
<i>Hylomantis granulosa</i>	37.4	Cruz 1988	Phyllomedusinae
		Duellman and	
<i>Hylomantis hulli</i>	37.1	Mendelson 1995	Phyllomedusinae
<i>Hylomantis lemur</i>	40.8	Duellman 2001	Phyllomedusinae
<i>Hylomantis medinai</i>	43.2	Cannatella 1980	Phyllomedusinae
<i>Hylomantis psilopygion</i>	42	Cannatella 1980	Phyllomedusinae

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<i>Pachymedusa dactylicolor</i>	82.6	Duellman 2001	Phyllomedusinae
<i>Phasmahyla cochranae</i>	33.9	Heyer et al. 1990	Phyllomedusinae
<i>Phasmahyla exilis</i>	34.5	Cruz 1980	Phyllomedusinae
<i>Phasmahyla guttata</i>	35	Cochran 1955	Phyllomedusinae
		Bokermann and	
<i>Phasmahyla jandaia</i>	31	Sazima 1978	Phyllomedusinae
<i>Phrynomedusa appendiculata</i>	33.4	Cruz 1985	Phyllomedusinae
<i>Phrynomedusa bokermanni</i>	46	Cruz 1991	Phyllomedusinae
		Izeckson and Cruz	
<i>Phrynomedusa marginata</i>	31	1976	Phyllomedusinae
<i>Phrynomedusa vanzolinii</i>	36.5	Cruz 1991	Phyllomedusinae
<i>Phyllomedusa atelopoides</i>	37.4	Duellman 2005	Phyllomedusinae
<i>Phyllomedusa azurea</i>	43.3	Caramaschi 2006	Phyllomedusinae
		Pombal and	
<i>Phyllomedusa bahiana</i>	74.5	Haddad 1992	Phyllomedusinae
<i>Phyllomedusa baltea</i>	45.2	Cannatella 1982	Phyllomedusinae
<i>Phyllomedusa bicolor</i>	115.3	Duellman 1978	Phyllomedusinae
<i>Phyllomedusa boliviana</i>	70.5	Vaira 2001	Phyllomedusinae
<i>Phyllomedusa burmeisteri</i>	79	Cochran 1955	Phyllomedusinae
<i>Phyllomedusa camba</i>	70	Duellman 2005	Phyllomedusinae
<i>Phyllomedusa centralis</i>	42	Bokermann 1965	Phyllomedusinae

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		Duellman and	
<i>Phyllomedusa coelestis</i>	64.8	Mendelson 1995	Phyllomedusinae
		Pombal and	
<i>Phyllomedusa distincta</i>	66	Haddad 1992	Phyllomedusinae
<i>Phyllomedusa duellmani</i>	54.2	Cannatella 1982	Phyllomedusinae
<i>Phyllomedusa ecuatoriana</i>	55.4	Cannatella 1982	Phyllomedusinae
<i>Phyllomedusa</i>			
<i>hypochondrialis</i>	37.7	Duellman 1997	Phyllomedusinae
<i>Phyllomedusa iheringii</i>	65	Cei 1980	Phyllomedusinae
		Caramaschi et al.	
<i>Phyllomedusa itacolomi</i>	42.6	2006	Phyllomedusinae
<i>Phyllomedusa megacephala</i>	43.2	Caramaschi 2006	Phyllomedusinae
		Barrio-Amorós	
<i>Phyllomedusa neildi</i>	63.8	2006	Phyllomedusinae
<i>Phyllomedusa nordestina</i>	42.1	Caramaschi 2006	Phyllomedusinae
<i>Phyllomedusa oreades</i>	42.64	Brandão 2002	Phyllomedusinae
<i>Phyllomedusa palliata</i>	49.1	Duellman 2005	Phyllomedusinae
<i>Phyllomedusa perinesos</i>	51.5	Cannatella 1982	Phyllomedusinae
<i>Phyllomedusa rohdei</i>	36	Cochran 1955	Phyllomedusinae
<i>Phyllomedusa sauvagii</i>	70	Cei 1980	Phyllomedusinae
<i>Phyllomedusa tarsi</i>	97	Duellman 1978	Phyllomedusinae

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		Pombal and	
<i>Phyllomedusa tetraploidea</i>	69.4	Haddad 1992	Phyllomedusinae
		Rodríguez and	
<i>Phyllomedusa tomopterna</i>	48	Duellman 1994	Phyllomedusinae
		Barrio-Amorós	
<i>Phyllomedusa trinitatis</i>	81	2006	Phyllomedusinae
		Duellman and	
<i>Phyllomedusa vaillantii</i>	59.9	Mendelson 1995	Phyllomedusinae
<i>Phyllomedusa venusta</i>	86.3	Duellman 2001	Phyllomedusinae

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- 1321
- 1322

1323

1324 Table S8. The 123 local sites considered in this study, indicating the size (in mm)  
 1325 of the species with the smallest maximum male body size (minimum size), the  
 1326 largest maximum male body size (maximum size), and the range of maximum  
 1327 male body sizes among species in the community.

1328

Locality	Minimum size	Maximum size	Range size
Brazil: Manaus, INPA	23	115	92
Brazil: Para: Belem	22	115	93
Ecuador: Santa Cecilia	20	132	112
Peru: Huanoco: Panguana	20	132	112
Peru: Loreto: Teniente Lopez	20	132	112
Peru: Madre de Dios: Manu National Park (Cocha Cashu)	20	132	112
Peru: Madre de Dios: Cuzco Amazonico Reserve	20	132	112
Venezuela: Bolivar: El Manteco	21	100	79
Venezuela: Gaurico: Hato Masaguaral	25	100	75

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Venezuela: La Escalera Region	21	132	111
Venezuela: Sierra de Lema	23	132	109
Argentina: La Rioja: Chilecito	56	56	0
Colombia: Boyaca: Parama de la Rusia	43	43	0
Ecuador: Napo: Rio Salado	22	59	37
Ecuador: Pichincha: Quebrada Zapadores	37	37	0
Peru: Cuzco: Rio Cosnipata	34	69	35
China: Fujian Prov.: Kuliang, near Fuzhou	32	32	0
China: Guangdong Prov.: Tai- Yong, E Kwantun	32	32	0
China: Guangdong Prov: Yim- Na-San	32	32	0
China: Guangdong: Dinghushan: Cha Chang	32	32	0
China: Jiangxi: Hong-San, SE Kiangsi Prov.	32	32	0
China: Yunnan Province, Nu Jiang Prefecture	35	35	0

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China: Yunnan, Baoshan			
Prefecture, Qushi	35	35	0
Japan: Kanagawa: Odawara			
City	39	39	0
Japan: Miyagi Pref.: Honshu:			
Takayama	39	39	0
Korea: Kyonggi Prov.; Mt.			
Buckak, near Seoul	39	39	0
Myanmar: Chin State: Falam			
Township, Lon Pi	35	35	0
Australia: New South Wales:			
Mutawintji National Park	37	80	43
Australia: New South Wales:			
Northern Yengo National Park	26	80	54
Australia: Northern Territory:			
Kakadu National Park	16	80	64
Australia: Northern Territory:			
Nitmiluk National Park	16	80	64
Australia: Queensland:			
Lamington National Park	26	80	54
Australia: Queensland:	26	81	55

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Townsville			
Australia: Queensland: Cairns	16	102	86
Australia: Queensland:			
Kuranda	16	102	86
Australia: Queensland: Dinden			
Forest Reserve	16	80	64
Australia: Queensland:			
Wooroonooran National Park	32	102	70
New Guinea: Lakekamu Basin,			
Ivimka Research	24	102	78
New Guinea: Utai	32	102	70
Argentina: Buenos Aires:			
Bahia Blanca	50	50	0
Argentina: Cordoba: Capilla			
de Monte	50	50	0
Argentina: Parque Nacional			
Chaco	20	101	81
Brazil: Goias: Espora Power			
Plant	19	101	82
Brazil: Mato Grosso:			
Dardanelos Dam	22	132	110

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Brazil: Rio Grande do Sul:			
Parque Nacional Aparados da Serra			
Serra	17	104	87
Brazil: Tocantins: Lajeado,			
Luis Eduardo Magalhaes Dam	22	113	91
Uruguay: Montevideo: Pajas Blancas			
Blancas	17	50	33
Uruguay: Salto: El Espinillar			
El Espinillar	17	50	33
Ecuador: Los Rios: Rio Palenque Biological Station			
Palenque Biological Station	26	90	64
France: Provence-Alpes-Cote-d'Azur; 3 mi W			
3 mi W	50	50	0
Georgia; ca. 10 km SSE			
Borzhomi	50	50	0
Germany: Rostock, Mecklenburg			
Mecklenburg	50	50	0
Hungary: Budapest			
Budapest	50	50	0
Morocco: Settat; Ben-Slimane, 5 km N of			
5 km N of	50	50	0
Spain: Andalusia: Cadiz Prov.:			
Benalup de Sidonia	50	50	0

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Ukraine: Kiev	50	50	0
Venezuela: Gran Sabana	21	132	111
Costa Rica: Cartago: Moravia	28	74	46
Costa Rica: Heredia: La Selva	24	76	52
Costa Rica: Heredia: Volcan Barba	33	41	8
Costa Rica: Las Canas: Finca Taboga	25	101	76
Costa Rica: Puntarenas: Las Cruces	28	110	82
Costa Rica: Puntarenas: Rincon de Osa	28	101	73
Guatemala: El Peten: Tikal	21	77	56
Honduras: Atlantida: La Ceiba	25	101	76
Honduras: Atlantida: Quebrada de Oro	28	76	48
Honduras: Copan: Laguna de Cerro	21	76	55
Honduras: Copan: Quebrada Grande	30	86	56
Honduras: Gracias de Dios:	21	100	79

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 Barra Patuka

Mexico: Chiapas: Rayon

Mescalapan	29	63	34
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Mexico: Distrito Federal: Lago

Xochimilco	35	35	0
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Mexico: Durango: El Salto	35	35	0
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Mexico: Guerrero: Puerto del

Gallo	30	81	51
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Mexico: Hidalgo: El Chico	35	48	13
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Mexico: Jalisco: Chamela	26	101	75
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Mexico: Michoachan: Nueva

Italia	26	86	60
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Mexico: Oaxaca: Puerto

Escondido	26	101	75
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Mexico: Oaxaca: San Gabriel

Mixtepec	29	81	52
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Mexico: Oaxaca: Tehuantepec	29	83	54
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Mexico: Oaxaca: Tuxtepec	21	101	80
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Mexico: Oaxaca: Vista

Hermosa	31	71	40
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Mexico: Puebla: 14.4 km W	38	44	6
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Huachinango			
Mexico: Sinaloa: Mazatlan	26	86	60
Mexico: Sonora: Alamos	63	83	20
Mexico: Veracruz: Acultzingo			
(1 km W)	38	54	16
Mexico: Veracruz: Cuatlapan	21	101	80
Mexico: Veracruz: Estacion			
Los Tuxtlas	21	78	57
Mexico: Veracruz: Huatusco			
(3k SW)	29	76	47
Mexico: Veracruz: Mata de			
Oscura	21	76	55
Mexico: Veracruz: Volcan San			
Martin	38	78	40
Mexico: Yucatan: Piste	21	101	80
Panama: Barro Colorado			
Island	24	101	77
Panama: Cocle: El Valle	24	110	86
Panama: Colon: Achioté	24	66	42
Panama: Darien: Rio Tuirá at			
Rio Mono	28	132	104

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California: Contra Costa Co.; 1			
mi NW Alamo	48	48	0
California: San Diego Co.:			
Spring, Pine Mountain	36	48	12
Colorado: Commanche			
National Grassland	32	32	0
Florida: Everglades National			
Park	16	59	43
Florida: Timucuan Ecological			
and Historic Preser	29	68	39
Georgia: Okeefenokee Swamp	16	68	52
Georgia: Okmulgee National			
Monument	29	59	30
North Carolina: Moores Creek			
National Battlefield	16	68	52
North Dakota: Badlands			
National Park	32	32	0
Ohio: Cuyahoga Valley			
National Park	29	51	22
South Carolina: Congaree			
Swamp National Monument	28	68	40

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South Carolina: Savannah			
River Ecology Laboratory	16	68	52
South Dakota: Sand Lake			
National Wildlife Refuge	29	51	22
Tennessee: Great Smoky			
Mountains National Park	29	51	22
Texas: Padre Island National			
Seashore	29	59	30
Utah: Zion National Park	52	52	0
Virginia: Shenandoah National			
Park	29	51	22
Washington: Pierce Co.			
Parkland, Wake Lake	48	48	0
Wyoming: Yellowstone			
National Park	32	32	0
Brazil: Bahia: Estacion			
VeraCruz	16	104	88
Brazil: Bahia: Zumbidos			
Palmares	19	104	85
Brazil: Boriacea	18	104	86
Dominican Republic: Samana:	40	109	69

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Laguna

Dominican Republic: Samana:

Rio San Juan 40 109 69

Haiti: Ouest: Furcy 54 109 55

Jamaica, Manchester Parish,

Mandeville 27 100 73

Jamaica, Quick Step 27 52 25

Jamaica: Trelany Parish:

Windsor 27 52 25

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1330

1331 Table S9. Values of the gamma statistic for the eight major clades of hylid frogs,  
 1332 including the observed gamma and associated *P*-value (indicating significant  
 1333 support for decreasing diversification over time within a clade), simulated  
 1334 critical values based on 5,000 simulation replicates (and associated *P*-value), and  
 1335 the difference between the observed and simulated critical gamma values (more  
 1336 negative values indicate support for a slowdown after accounting for incomplete  
 1337 taxon sampling). Asterisked clades are partially or entirely allopatric relative to  
 1338 other hylid clades.

Clade	Observed gamma	<i>P</i> -value	Simulated critical value	Observed - simulated	<i>P</i> -value	Species in tree (% of total richness)
Pelodyridinae*	-3.2256	0.0006	-3.992	0.767	0.2327	51 (27%)
Phyllomedusinae	-2.5790	0.0050	-2.6073	0.028	0.0526	26 (43%)
Cophomantini	-3.1912	0.0007	-3.3079	0.117	0.0614	77 (48%)
<i>Dendropsophus</i> clade	-4.0309	2.7780e-05	-3.0890	-0.942	0.0030	35 (36%)
<i>Scinax</i> clade	-3.8414	6.1167e-05	-3.3395	-0.502	0.0122	29 (26%)
<i>Pseudis</i> clade	0.2583	0.6019	NA	0.258	NA	13 (100%)



Lophiohylini	-2.8915	0.0019	-2.5749	-0.317	0.0228	35 (51%)
Hylini*	-2.7314	0.0032	-3.2942	0.563	0.1444	95 (52%)

1339

1340 Table S10. Eigenvectors for PC1 from the PCA of 19 climatic variables, based on  
1341 analysis of 123 local sites.

1342

Climatic variable	Eigenvector	Description of variable
Bio1	0.28975	Annual mean temperature
Bio2	-0.15425	Mean diurnal temperature range [mean of monthly (maximum temperature ) minimum temperature)
Bio3	0.25826	Isothermality (Bio2/Bio7 · 100)
Bio4	-0.28808	Temperature seasonality (standard deviation of monthly temperature)
Bio5	0.07982	Minimum temperature of the coldest month
Bio6	0.31530	Maximum temperature of the warmest month
Bio7	-0.29840	Temperature range (Bio6 - Bio5)
Bio8	0.18895	Mean temperature of the wettest quarter
Bio9	0.28156	Mean temperature of the driest quarter
Bio10	0.15957	Mean temperature of the warmest quarter
Bio11	0.30757	Mean temperature of the coldest quarter

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Bio12	0.26747	Annual precipitation
Bio13	0.24597	Precipitation of the wettest month
Bio14	0.15441	Precipitation of the driest month
Bio15	0.02171	Precipitation seasonality (standard deviation of monthly precipitation)
Bio16	0.24808	Precipitation of the wettest quarter
Bio17	0.16023	Precipitation of the driest quarter
Bio18	0.17491	Precipitation of the warmest quarter
Bio19	0.19629	Precipitation of the coldest quarter

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1344

1345 Table S11. Eigenvectors for PC1 from the PCA of 19 climatic variables, based on  
 1346 analysis of mean climatic values (averaged across localities within a species for  
 1347 each variable) for 337 hylid species.  
 1348

Climatic variable	Eigenvector	Description of variable
Bio1	0.28190	Annual mean temperature
Bio2	-0.0133	Mean diurnal temperature range [mean of monthly (maximum temperature ) minimum temperature)
Bio3	0.11373	Isothermality (Bio2/Bio7 · 100)
Bio4	-0.28259	Temperature seasonality (standard deviation of monthly temperature)
Bio5	0.07620	Minimum temperature of the coldest month
Bio6	0.31746	Maximum temperature of the warmest month
Bio7	-0.28332	Temperature range (Bio6 - Bio5)
Bio8	0.18611	Mean temperature of the wettest quarter
Bio9	0.30190	Mean temperature of the driest quarter
Bio10	0.14588	Mean temperature of the warmest quarter
Bio11	0.29517	Mean temperature of the coldest quarter
Bio12	0.29303	Annual precipitation
Bio13	0.27884	Precipitation of the wettest month

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Bio14	0.18097	Precipitation of the driest month
Bio15	0.02554	Precipitation seasonality (standard deviation of monthly precipitation)
Bio16	0.28635	Precipitation of the wettest quarter
Bio17	0.18773	Precipitation of the driest quarter
Bio18	0.21063	Precipitation of the warmest quarter
Bio19	0.21886	Precipitation of the coldest quarter

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1350

1351 Table S12. Estimates of mean local richness and estimated time of colonizaiton  
 1352 (in millions of years before present) for 14 bins representing the range of  
 1353 variation in PC1 among sites.  
 1354

Range of PC1	Number of sites	Mean local richness	Estimated time
4-5	7	12.714	60.05
3-4	9	12.778	126.79
2-3	18	10.611	226.29
1-2	13	10.308	314.92
0-1	18	8.440	310.23
-1-0	9	4.667	197.02
-2-(-1)	12	5.333	151.43
-3-(-2)	13	4.308	94.29
-4-(-3)	10	1.500	87.29
-5-(-4)	3	1.000	48.29
-6-(-5)	5	2.000	38.41
-7-(-6)	1	1.000	34.52
-8-(-7)	1	3.000	2.44
-9-(-8)	1	1.000	2.44

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1356

1357 Table S13. Local species richness, regional species richness, summed clade ages  
 1358 (millions of years), annual mean temperature, annual precipitation, and PC1  
 1359 (from a principal components analysis of 19 climatic variables (Table S10) across  
 1360 123 local site) for each of 12 local sites, each representing the 12 major regions  
 1361 (see Fig. 1). In this table, the site representing each region has local richness  
 1362 equal (or close to) the mean local richness for the region, as opposed to the  
 1363 maximum local richness (see instead Table 1). Estimates for the oldest  
 1364 colonization of hylids in each region are given in Table 1.  
 1365

Local site	Region	Local richness	Regional richness	Summed clade ages	Temp. (°C)	Precip. (mm)	PC1
Peru: Panguana	Amazon	21	142	333.601	25.7	2179	2.678
Brazil: Boriacea	Atlantic rainforests	26	166	134.873	17.2	2468	0.606
Venezuela: Gran Sabana	Guyana highlands	8	23	58.922	18.4	2162	1.183
Ecuador:	Chocó	10	31	76.692	23.8	3463	4.398

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Palenque							
Argentina: Chaco	Cerrado	10	127	160.882	21.5	1336	-
National Park							0.824
Peru: Rio	Andes	3	82	103.811	18.4	1814	.315
Cosnipata							
China: Cha	Asia	1	10	26.575	19.8	1674	-.458
Chang							
Australia:	Australi	12	189	58.41	24.4	1333	.612
Townsend	a						
France:	Europe	1	8	22.113	14.3	900	-
Provence-Alpes-							2.676
Cote-d'Azur							
Mexico: Piste	Middle	7	159	163.157	25.4	1234	0.979
America							
U.S.A.: Great	North	4	29	44.397	12.5	1384	-
Smoky	America						3.663
Mountains							
Jamaica: Quick	West	3	10	49.983	23.0	1742	1.895
Step	Indies						

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1370

1371 Table S14. Local species richness, time of first colonization by hylids (in millions  
 1372 of years), summed clade ages (millions of years), annual mean temperature,  
 1373 annual precipitation, and PC1 (based on a principal components analysis of 19  
 1374 climatic variables (Table S10) across 123 local sites) for 19 local sites representing  
 1375 19 regions, after subdividing several of the original 12 regions (e.g., Amazon,  
 1376 Australia, Cerrado, Middle America, North America). Note that the time of first  
 1377 colonization and summed clade ages are the same for subdivided regions,  
 1378 because of the limited number of regions that can be analyzed with existing  
 1379 likelihood methods for biogeographic reconstructions.

1380

Local site	Region	Local richness	Time first colonization	Summed clade ages	Temp. (° C)	Precip. (mm)	PC1
Ecuador: Santa Cecilia	Amazon-Western	36	77.605	333.60	25.4	3670	5.360
Brazil: Para: Belem	Amazon-Eastern	23	77.605	333.60	26.9	2728	4.363
Venezuela: Gaurico: Hato	Amazon-Llanos	5	77.605	333.60	27.5	1336	2.234

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Masaguaral							
Brazil: Estacion	Atlantic	28	38.438	134.87	24.0	1540	1.723
VeraCruz	rainforest			3			
Venezuela: Gran	Guyana	8	35.905	58.922	18.4	2162	1.183
Sabana	highland						
	s						
Ecuador:	Chocó	10	20.498	76.692	23.8	3463	4.398
Palenque							
Brazil:	Cerrado-	21	37.974	160.88	24.4	2256	1.312
Dardanelos Dam	Cerrado			2			
Argentina: Chaco	Cerrado-	10	37.974	160.88	21.5	1336	-0.824
National Park	Chaco			2			
Uruguay: Salto: El	Cerrado-	9	37.974	160.88	19.6	1434	-1.080
Espinillar	Monte			2			
Ecuador: Rio	Andes	7	52.203	103.81	19.8	2742	2.740
Salado				1			
China: Cha Chang	Asia	1	26.575	26.575	19.8	1674	-0.458
Australia:	Australia	17	58.410	58.41	26.7	1417	0.545
Nitmiluk	-						
National Park	Australia						

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Australia: New Guinea:	Australia	9	58.410	58.41	26.7	1835	2.717
Lakekamu Basin, Ivimka Research	Guinea						
France: Provence-Alpes-Cote-d'Azur	Europe	1	22.113	22.113	14.3	900	-0.302
Costa Rica: La Selva	Middle America-Central America	12	55.220	163.15	26.1	4147	5.537
				7			
Mexico: Veracruz: Cuatlapan	Middle America-Mexico	9	55.220	163.15	20.4	1757	0.307
				7			
U.S.A.: Savannah River Ecology Lab	North America-Eastern	12	44.397	44.397	16.3	1290	-2.182
U.S.A.: California: Pine Mountain	North America-Western	2	44.397	44.397	13.8	508	-0.396
Jamaica: Quick	West	3	31.017	49.983	23.0	1742	1.895

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Step

Indies

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1381

1382

1383 **Appendix S2.A.** Results of likelihood-based biogeographic analyses (using  
 1384 LAGRANGE), treating South America as a single region. Regions are: A = South  
 1385 America; B = Middle America; C = Australia; D = North America; E = Europe; F  
 1386 = West Indies; G = Asia.

1387

1388

1389 lagrange: likelihood analysis of geographic range evolution

1390 Version 2 released February 2008

1391 This is development snapshot 20090327

1392 Authors: Richard Ree <rree@fieldmuseum.org>

1393 Stephen Smith <sasmith@nescent.org>

1394 <http://lagrange.googlecode.com>

1395 Newick tree with interior nodes labeled:

1396

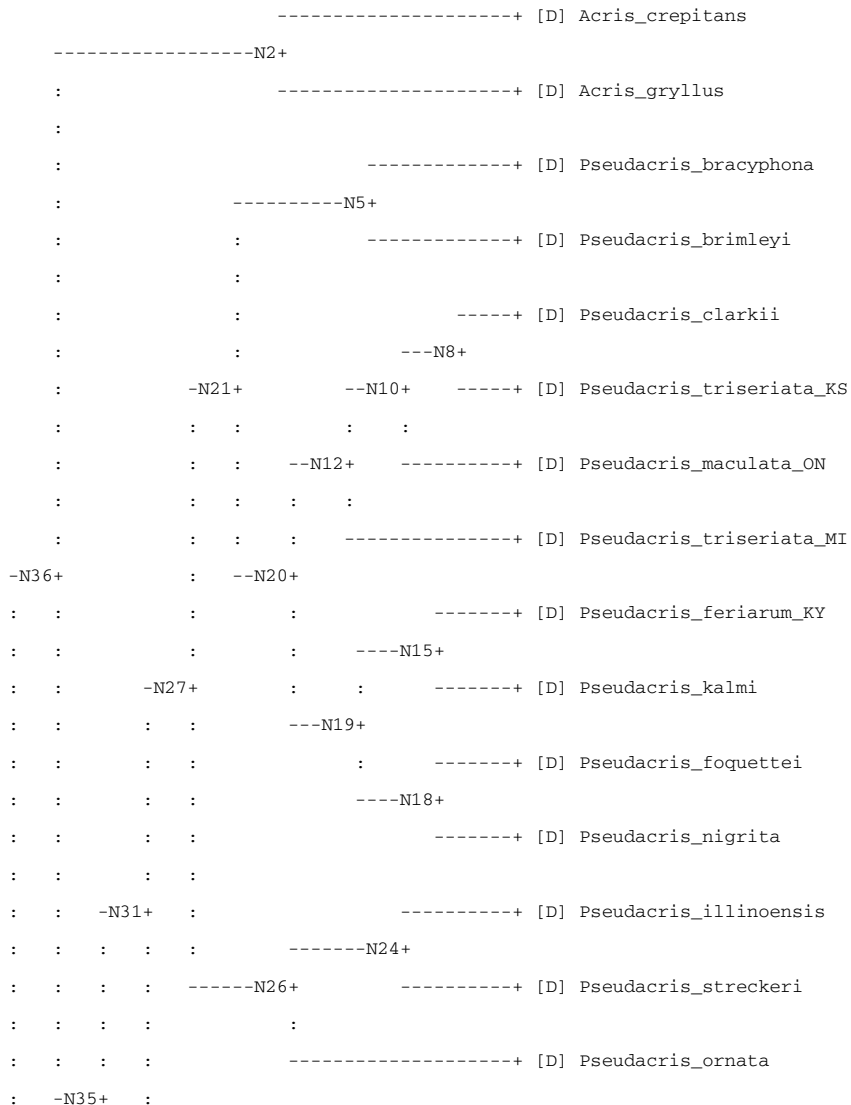
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1566 3.02,(Litoria\_inermis:7.81,Litoria\_pallida:7.81)N658:5.21)N659:2.9,Litor  
1567 ia\_nasuta:15.92)N661:3.11)N662:2.76,Litoria\_tornieri:21.79)N664:17.9)N66  
1568 5:3.88)N666:1.46,Litoria\_meiriana:45.03)N668:13.38)N669:13.5,(((((((Ag  
1569 alychnis\_anna:9.15,Agalychnis\_moreleti:9.15)N672:3.68,Agalychnis\_callid  
1570 ryas:12.82)N674:4.42,((Agalychnis\_litodryas:1.83,Agalychnis\_spurrelli:1.  
1571 83)N677:12.14,Agalychnis\_saltator:13.98)N679:3.27)N680:5.3,Pachymedusa\_d  
1572 acnicolor:22.55)N682:10.55,Hylomantis\_granulosa:33.1)N684:4.64,Hylomanti  
1573 s\_lemur:37.74)N686:7.74,Phrynomedusa\_marginata:45.48)N688:2.24,((Phasmah  
1574 yla\_cochranae:13.67,Phasmahyla\_guttata:13.67)N691:29.65,(((Phyllomedusa  
1575 \_atelopoides:21.35,(Phyllomedusa\_duellmani:15.65,Phyllomedusa\_perinesos:  
1576 15.65)N695:5.7)N696:5.16,((Phyllomedusa\_azurea:12.47,Phyllomedusa\_hypoch  
1577 ondrialis:12.47)N699:5.13,Phyllomedusa\_palliata:17.6)N701:8.91)N702:3.04  
1578 ,Phyllomedusa\_tomopterna:29.55)N704:6.43,((Phyllomedusa\_bicolor:17.37,Ph  
1579 yllomedusa\_vaillantii:17.37)N707:10.77,(Phyllomedusa\_boliviana:23.15,(Ph  
1580 yllomedusa\_tarsius:3.98,Phyllomedusa\_trinitatus:3.98)N711:13.86,Phyllome  
1581 dusa\_tetraploidea:17.84)N713:5.31)N714:4.98)N715:7.84)N716:7.34)N717:4.4  
1582 )N718:5.77,Cruziophyla\_calcarifer:53.49)N720:18.42)N721:11.06)N722;  
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Cladogram (branch lengths not to scale):



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: : : -----+ [D] Pseudacris_crucifer
: : -----N30+
: : -----+ [D] Pseudacris_ocularis
: : -----+ [D] Pseudacris_cadaverina
: -----N34+
: -----+ [D] Pseudacris_regilla
: -----+ [B] Anotheca_spinosa
: -----N39+
: -----N41+ -----+ [B] Triprion_petasatus
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: : -----+ [B] Diaglena_spatulata
: :
: N55+ -----+ [BD] Smilisca_baudinii
: : :
: : : -----+ [B] Smilisca_cyanosticta
: : : -N47+
: : -N54+ : : ----+ [AB] Smilisca_phaeota
: : : -N49+ -N46+
: : : : ----+ [B] Smilisca_puma
: : : :
: N63+ -N53+ -----+ [BD] Smilisca_fodiens
: : :
: : : -----+ [AB] Smilisca_sila
: : : -----N52+
: : : -----+ [B] Smilisca_sordida
: : :
: : : -----+ [B] Isthmohyla_pseudopuma
: : : -----N58+
: N71+ : : -----+ [B] Isthmohyla_zeteki
: : : -----N62+
: : : : -----+ [B] Isthmohyla_rivularis
: : : -----N61+
: : : -----+ [B] Isthmohyla_tica
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: : : -----+ [B] Tlalocohyla_godmani
: : : -----N66+
: : : : -----+ [B] Tlalocohyla_loquax
: : : -----N70+
: : : : -----+ [B] Tlalocohyla_picta
: : : -----N69+
: : : -----+ [B] Tlalocohyla_smithi
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: : : -----+ [D] Hyla_andersoni
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1742 : : : : -----+ [D] Hyla_avivoca
1743 : : : : -N80+ --N75+
1744 : : : : --N77+ -----+ [D] Hyla_chrysocelis
1745
1746 : : : : : :
1747 N188+ : : : : : :
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1749 : : : : --N79+ -----+ [D] Hyla_versicolor
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1751 : : N115+ : :
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1754 : : : : : : -----+ [D] Hyla_femorialis
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1756 : : : : : :
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1758 : : : : N96+ -----+ [BD] Hyla_arenicolor
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1762 : : : : : : : -----+ [B] Hyla_euphorbiacea
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1764 : : : : : : : -N84+
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1766 : : : : : : N91+ -N86+ -----+ [B] Hyla_plicata
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1768 : : : : : : : : : :
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1770 : : : : : : : : N88+ -----+ [B] Hyla_eximia
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1772 : : : : : : : : : :
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1774 : : : : : : N95+ N90+ -----+ [B] Hyla_walkerii
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1776 : : : : N102+ : :
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1778 : : : : : : : : : : -----+ [BD] Hyla_wrightorum
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1780 : : : : : : : :
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1782 : : : : : : : : -----+ [G] Hyla_immaculata
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1784 : : : : : : : : -----N94+
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1786 : : : : : : : : -----+ [G] Hyla_japonica
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1788 : : : : : : : :
1789
1790 : : N121+ : : : : -----+ [D] Hyla_cinerea
1791
1792 : : : : : : : : -----N99+
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1794 : : : : N114+ -----N101+ -----+ [D] Hyla_gratiosa
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1798 : : : : : : : : -----+ [D] Hyla_squirella
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1802 : : : : : : : : -----+ [G] Hyla_annectans
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1804 : : : : : : : : -----N105+
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1812 : : : : ---N113+
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1814 : : : : : : : : -----+ [E] Hyla_arborea
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1816 : : : : : : : : -----N110+
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1818 : : : : : : ---N112+ -----+ [E] Hyla_sauvignyii
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1826 : : : : : : : : -----+ [B] Charadrahyla_nephila
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1830 : : : : -----N120+ -----+ [B] Charadrahyla_taeniopus
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: : N149+ -----+ [B] Megastomatohyala_mixe
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: : : : :
: : : : ---N128+ -----+ [B] Duellmanohyla_rufiocolis
: : : : : : ---N125+
: : : : : ---N127+ -----+ [B] Duellmanohyla_uranochroa
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: : : : : -----+ [B] Duellmanohyla_soralia
: : : : :
: : : : N142+ -----+ [B] Ptychohyala_dendrophasma
: : : : : :
: : : : : : -----+ [B] Ptychohyala_euthysanota
: : : : : : N139+ :
: : : : : : : N136+ ----+ [B] Ptychohyala_leonhardtschultei
: : : : : : : : N133+
: : : : : : : : N135+ ----+ [B] Ptychohyala_zophodes
: : : : N144+ N141+ N138+ :
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: : : : : : -----+ [B] Ptychohyala_hypomycter
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: N187+ N148+ : -----+ [B] Ptychohyala_spinipollex
: : : :
: : : : -----+ [B] Ecnomiohyala_miotympanum
N258+ : :
: : : : -----+ [B] Ecnomiohyala_miliara
: : : : -----N147+
: : : : -----+ [B] Ecnomiohyala_minera
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: : : : -----+ [B] Exerodonta_abdivita
: : : : -----N152+
: : : : ----N154+ -----+ [B] Exerodonta_perkinsi
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: : : : : -----+ [B] Exerodonta_melanoma
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: : : : ---N162+ -----+ [B] Exerodonta_chimalapa
: : : : : : ---N157+
: : : : : : ---N159+ -----+ [B] Exerodonta_smaragdina
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: : : : : ---N161+ -----+ [B] Exerodonta_xera
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: : : : : -----+ [B] Exerodonta_sumichrasti
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: : : : : -----+ [B] Plectrohyla_ameibothalamae
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: : : : : -----+ [B] Plectrohyla_bistincta
: : : : : --N168+
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: : : --N177+ -----+ [B] Plectrohyla_pentheter
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: : : : : --N172+
: : : : : --N174+ -----+ [B] Plectrohyla_cyclada
: : --N185+ : : :
: : : --N176+ -----+ [B] Plectrohyla_siopela
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: : : -----+ [B] Plectrohyla_chrysopleura
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: : : -----+ [B] Plectrohyla_glandulosa
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: : : -----+ [B] Plectrohyla_guatemalensis
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: : -----+ [B] Plectrohyla_matudai
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: : -----+ [A] Aparasphenodon_brunoi
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: : : : : --N199+
: : : : -N208+ --N201+ -----+ [A] Trachycephalus_resinifrixtix
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: : : : : --N205+ -----+ [AB] Trachycephalus_venulosus
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: : : -N210+ -N207+ -----N204+
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: : : : ---N263+
: : : : -----+ [A] Dendropsophus_minusculus
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: : : : N273+ -----+ [A] Dendropsophus_nanus
: : : : : N266+
: : : : : N268+ -----+ [A] Dendropsophus_walfordi
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: : : : N288+ : N272+ -----+ [A] Dendropsophus_riveroi
: : : : : N275+ :
: : : : : : -----+ [A] Dendropsophus_rubicundula
: : : : : : --N271+
: : : : : : -----+ [A] Dendropsophus_sanborni
: : : : : N285+ :
: : : : : : : -----+ [A] Dendropsophus_bipunctata
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: : : : : : : -----+ [A] Dendropsophus_leali
: : : : : : : -N284+ -----+ [AB] Dendropsophus_microcephalus
: : : : : N287+ : -N279+
: : : : : : : -----+ [A] Dendropsophus_rhodopepla
: : : N325+ : : -N283+
: : : : : : : -----+ [B] Dendropsophus_robertmertensi
: : : : : N304+ : -N282+
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: : : : : N318+ : : -----+ [A] Dendropsophus_ebraccatus
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2202 : : N327+ : : : -----+ [A] Dendropsophus_schubarti
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2216 : : : : : --N317+
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2218 : : : : : : -----+ [A] Dendropsophus_carnifex
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2220 : : : : : : ---N314+
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2235
2236 : : : : -----N323+
2237
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2239
2240 : : : : -----N322+
2241
2242 : : N355+ : : -----+ [A] Dendropsophus_senicula
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2252 : : : : -----N332+
2253
2254 : : : : : : -----+ [A] Lysapsus_limellus
2255
2256 : : : : : -----N331+
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2258 : : : : ---N336+ -----+ [A] Lysapsus_limellus_bolivianus
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2263
2264 : : : : : -----N335+
2265
2266 : : : : : -----+ [A] Lysapsus_sp.
2267
2268 : : : : -N352+
2269
2270 : : : : : : -----+ [A] Podonectes_cardosoi
2271
2272 : : : : : : -----N339+
2273
2274 : : : : : : -----+ [A] Podonectes_minutus
2275
2276 : : : : : :
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2278 : : : : : : -----+ [A] Pseudis_bolbodactyla
2279
2280 : : : : : -N351+ :
2281
2282 : : : : : : --N346+ -----+ [A] Pseudis_paradoxa
2283
2284 : : : -N354+ : : : --N343+
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2286 : : : : : : --N345+ -----+ [A] Pseudis_par_platensis
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2288 : : : : : --N350+ :
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2290 : : : : : -----+ [A] Pseudis_par_occidentalis
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2292 : : : : :

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2295	:	:	:	:	:	-----N349+	
2296	:	:	:	:	:		
2297	:	:	:	:	:	-----+	[A] Pseudis_tocantins
2298	:	:	:	:	:		
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2312	:	:	:	:	:	--N359+	
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2317	:	:	:	:	:	--N366+      --N361+      -----	[A] Scinax_sugillatus
2318	:	:	:	:	:		
2320	:	:	:	:	:	--N363+      -----	[A] Scinax_garbei
2322	:	N413+	:	:	:		
2323	:	:	:	:	:	--N365+      -----	[A] Scinax_pedromedinai
2324	:	:	:	:	:		
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2332	:	:	:	:	:	--N373+	
2334	:	:	:	:	:		
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2336	:	:	:	:	:	--N372+	
2337	:	:	:	:	:		
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2340	:	:	:	:	:	--N371+	
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2348	:	:	:	:	:		
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2352	:	:	:	:	:		
2354	:	:	:	:	:	N380+      -----	[A] Scinax_oreites
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2362	:	:	:	:	:	N393+      N384+      -----	[A] Scinax_hayi
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2365	:	:	:	:	:		
2366	:	:	:	:	:	N386+      -----	[A] Scinax_similis
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2370	:	:	:	:	:	N400+      -----	[A] Scinax_duartei
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2372	:	:	:	:	:		
2374	:	:	:	:	:	N395+      -----	[A] Scinax_cruentomma
2377	:	:	:	:	:	-----N390+	
2378	:	:	:	:	:	-----N392+      -----	[B] Scinax_elaeochroa
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2382	:	:	:	:	:	N397+      -----	[B] Scinax_staufferi
2384	:	:	:	:	:	N406+      -----	



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2401 : N568+ : : : -----+ [A] Scinax\_catharinae  
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2403 : : : : : : -----N404+  
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2488	:	:	:	:	:		--N446+			
2490	:	:	:		--N451+	--N448+	-----+	[A]	Hypsiboas_sp2	
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2495	:	:	:	:	:	:				
2496	:	:	:	:	:	:				
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2508	:	:	:	:	:		---N456+			
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2546	:	:		N524+	:	:	N475+	-----+	[A]	Hypsiboas_marginata
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2553	:	:	:	:	:					
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2557	:	:	:	:	:					
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2562	:	:	:	:	:		N481+	----+	[A]	Hypsiboas_pulchella
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2568	:	:	:	:	:					

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2571 : : : : : : : : : : -N487+
2572 : : : : : : : : : : N489+ -----+ [A] Hypsiboas_aff_semiguttata_sp7
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2575 : : N541+ : : : : : : : -----+ [A] Hypsiboas_semiguttata
2576 : : : : : : : : : : N495+
2577 : : : : : : : : : : : : -----+ [A] Hypsiboas_latistriata
2578 : : : : : : : : : : : : -N492+
2579 : : : : : : : : : : N494+ -----+ [A] Hypsiboas_polytaenia
2580 : : : : : : : : : : : :
2581 : : : : : : : : : : -----+ [A] Hypsiboas_leptolineata
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2586 : : : : : : : : : : -----N502+
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2598 : : : : : : : : : : -N516+ :
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2600 : : : : : : : : : : : :
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2604 : : : : : : : : : : -----N522+
2605 : : N563+ : : : : : : -----+ [A] Hypsiboas_geographica
2606 : : : : : : : : : : -----N521+
2607 : : : : : : : : : : -----+ [A] Hypsiboas_semilineata
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2614 : : : : : : : : : : : : --N530+
2615 : : : : : : : : : : --N536+ --N532+ -----+ [A] Bokermannohyla_sp3
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2754 : : : -----+ [C] Litoria_dahliei
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2756 : N587+ -----N582+
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2758 : : : -----+ [C] Litoria_eucnemis
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2762 : : : -----+ [C] Litoria_aurea
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2764 : : -----N586+
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2766 : N595+ -----+ [C] Litoria_cyclorhyncha
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2772 : : : -----N590+
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2780 : N607+ -----N593+
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2788 : : : -----N600+
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2790 : : : : : -----+ [C] Litoria_gilleni
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2792 : : : : -----N599+
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2794 : : -----N606+ -----+ [C] Litoria_splendida
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2796 : N611+ :
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2812 : N627+ -----N610+
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2824 : : : : --N619+
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2828 : : --N626+ : ---N618+
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2830 : : : : : -----+ [C] Nyctimystes_humeralis
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2832 : : : --N623+ ---N617+
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2840 : : : -----N622+
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2842 : : : : -----+ [C] Nyctimystes_narinosus
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:      :      -----+ [C] Nyctimystes_papua
:      :
:      :      -----+ [C] Litoria_amboinensis
:      :      -----N630+
:      :      -----N632+      -----+ [C] Litoria_peronii
:      :      :      :
:  N669+  -----N636+  -----+ [C] Litoria_rothii
:      :      :      :
:      :      :      :      -----+ [C] Litoria_dentata
:      :      :      :      -----N635+
:      :      :      :      -----+ [C] Litoria_rubella
:      :      :      :
:      :      :      :      -----+ [C] Litoria_arfakiana
:      :      :      :      ---N639+
:      :      :      :      ---N641+      -----+ [C] Litoria_thesaurensis
:      :      :      :      :      :
:      :      :  -N666+      :      -----+ [C] Litoria_modica
:      :      :      :      ---N647+
:      :      :      :      :      :      -----+ [C] Litoria_bicolor
:      :      :      :      :      :      ---N644+
:      :      :      :      :  --N649+  ---N646+      -----+ [C] Litoria_booroolongensis
:      :      :      :      :      :      :
:      :      :      :      :      :      -----+ [C] Litoria_fallax
:      :      :      :      :      :
:      :      :      :      :      -----+ [C] Litoria_microbelos
:      :      :      :      :
:      :      :      :      :      -----+ [C] Litoria_coplandi
:      :      :      :  -N665+      ----N654+
:      :      :      :      :      :      -----+ [C] Litoria_watjulumensis
:      :  -N668+      :      :      ----N653+
:      :      :      :      :      -----+ [C] Litoria_wollastoni
:      :      :      :      :  -N662+
:      :      :      :      :      :      -----+ [C] Litoria_frecyneti
:      :      :      :      :      :      --N659+
:      :      :      :      :      :      :      :      -----+ [C] Litoria_inermis
:      :      :      :  -N664+  --N661+  --N658+
:      :      :      :      :      :      -----+ [C] Litoria_pallida
:      :      :      :      :      :
:      :      :      :      :      -----+ [C] Litoria_nasuta
:      :      :      :      :
:      :      :      :      -----+ [C] Litoria_tornieri
:      :      :
N721+  -----+ [C] Litoria_meiriana
:
:      :      -----+ [B] Agalychnis_anae
:      :
--N672+

```

```

2037
2038 : --N674+ -----+ [B] Agalychnis_moreletii
2039 : : :
2040 : : :
2041 : : :
2042 : : :
2043 : : :
2044 : : :
2045 : : :
2046 : : :
2047 : : :
2048 : : :
2049 : : :
2050 : : :
2051 : : :
2052 : : :
2053 : : :
2054 : : :
2055 : : :
2056 : : :
2057 : : :
2058 : : :
2059 : : :
2060 : : :
2061 : : :
2062 : : :
2063 : : :
2064 : : :
2065 : : :
2066 : : :
2067 : : :
2068 : : :
2069 : : :
2070 : : :
2071 : : :
2072 : : :
2073 : : :
2074 : : :
2075 : : :
2076 : -N718+ -----N691+
2077 : : : :
2078 : : : :
2079 : : : :
2080 : : : :
2081 : : : :
2082 : : : :
2083 : : : :
2084 : : : :
2085 : : : :
2086 : : : :
2087 : : : :
2088 : : : :
2089 : : : :
2090 : : : :
2091 : : : :
2092 : : : :
2093 : : : :
2094 : : : :
2095 : : : :
2096 : : : :
2097 : : : :
2098 : : : :
2099 : : : :
3020 : : : :
3021 : : : :
3022 : : : :
3023 : : : :
3024 : : : :
3025 : : : :
3026 : : : :
3027 : : : :
3028 : : : :

```

```

3029
3030 : -----+ [A] Phyllomedusa_tetraploidea
3031 :
3032 :
3033 :
3034 -----+ [AB] Cruziohyala_calcarifer
3035
3036
3037
3038
3038
3040
3041
3042 Global ML at root node:
3043 -lnL = 177.2
3044 dispersal = 0.001134
3045 extinction = 4.285e-009
3046
3047 Ancestral range subdivision/inheritance scenarios ('splits') at
3048 internal nodes.
3049
3050 * Split format: [left|right], where 'left' and 'right' are the ranges
3051 inherited by each descendant branch (on the printed tree, 'left' is
3052 the upper branch, and 'right' the lower branch).
3053
3054 * Only splits within 2 log-likelihood units of the maximum for each
3055 node are shown. 'Rel.Prob' is the relative probability (fraction of
3056 the global likelihood) of a split.
3057
3058 At node N722:
3059 split lnL Rel.Prob
3060 [A|AC] -177.4 0.8697
3061
3062 At node N568:
3063 split lnL Rel.Prob
3064 [A|A] -177.3 0.9155
3065
3066 At node N414:
3067 split lnL Rel.Prob
3068 [A|A] -177.5 0.7922
3069 [AB|A] -178.8 0.2067
3070
3071 At node N258:
3072 split lnL Rel.Prob
3073 [B|A] -177.2 0.9994
3074
3075 At node N188:
3076 split lnL Rel.Prob
3077 [D|B] -177.7 0.6089
3078 [D|BD] -178.2 0.3911
3079
3080 At node N36:
3081 split lnL Rel.Prob
3082 [D|D] -177.2 1
3083
3084 At node N2:
3085 split lnL Rel.Prob
3086 [D|D] -177.2 1
3087
3088 At node N35:
3089 split lnL Rel.Prob
3090 [D|D] -177.2 1
3091
3092 At node N187:
3093 split lnL Rel.Prob
3094 [B|B] -177.7 0.6076
3095 [BD|B] -178.2 0.3916
3096

```



```

3097 At node N149:
3098 split lnL Rel.Prob
3099 [B|B] -177.7 0.6031
3100 [BD|B] -178.1 0.3958
3101
3102 At node N121:
3103 split lnL Rel.Prob
3104 [B|B] -177.8 0.567
3105 [BD|B] -178.1 0.4299
3106
3107 At node N115:
3108 split lnL Rel.Prob
3109 [B|D] -177.3 0.9707
3110
3111 At node N71:
3112 split lnL Rel.Prob
3113 [B|B] -177.3 0.9706
3114
3115 At node N63:
3116 split lnL Rel.Prob
3117 [B|B] -177.3 0.9702
3118
3119 At node N55:
3120 split lnL Rel.Prob
3121 [B|B] -177.3 0.9631
3122
3123 At node N41:
3124 split lnL Rel.Prob
3125 [B|B] -177.2 1
3126
3127 At node N54:
3128 split lnL Rel.Prob
3129 [B|B] -177.3 0.9213
3130
3131 At node N53:
3132 split lnL Rel.Prob
3133 [B|B] -177.2 0.9807
3134
3135 At node N49:
3136 split lnL Rel.Prob
3137 [B|B] -177.3 0.9565
3138
3139 At node N47:
3140 split lnL Rel.Prob
3141 [B|B] -177.2 0.9841
3142
3143 At node N46:
3144 split lnL Rel.Prob
3145 [B|B] -177.3 0.9285
3146
3147 At node N52:
3148 split lnL Rel.Prob
3149 [B|B] -177.3 0.9138
3150
3151 At node N62:
3152 split lnL Rel.Prob
3153 [B|B] -177.2 1
3154
3155 At node N70:
3156 split lnL Rel.Prob
3157 [B|B] -177.2 1
3158
3159 At node N114:
3160 split lnL Rel.Prob

```

3161	[DG G]	-177.4	0.8044
3162			
3163	At node N102:		
3164	split lnL		Rel.Prob
3165	[DG D]	-177.4	0.8183
3166	[D D]	-179	0.1763
3167			
3168	At node N96:		
3169	split lnL		Rel.Prob
3170	[D DG]	-177.3	0.8815
3171			
3172	At node N80:		
3173	split lnL		Rel.Prob
3174	[D D]	-177.2	1
3175			
3176	At node N79:		
3177	split lnL		Rel.Prob
3178	[D D]	-177.2	1
3179			
3180	At node N77:		
3181	split lnL		Rel.Prob
3182	[D D]	-177.2	1
3183			
3184	At node N95:		
3185	split lnL		Rel.Prob
3186	[D G]	-177.2	1
3187			
3188	At node N91:		
3189	split lnL		Rel.Prob
3190	[D D]	-177.8	0.5736
3191	[BD B]	-178.7	0.2269
3192			
3193	At node N90:		
3194	split lnL		Rel.Prob
3195	[B BD]	-177.4	0.8106
3196	[B B]	-178.9	0.1844
3197			
3198	At node N88:		
3199	split lnL		Rel.Prob
3200	[B B]	-177.2	1
3201			
3202	At node N86:		
3203	split lnL		Rel.Prob
3204	[B B]	-177.2	1
3205			
3206	At node N84:		
3207	split lnL		Rel.Prob
3208	[B B]	-177.2	1
3209			
3210	At node N94:		
3211	split lnL		Rel.Prob
3212	[G G]	-177.2	1
3213			
3214	At node N101:		
3215	split lnL		Rel.Prob
3216	[D D]	-177.2	1
3217			
3218	At node N99:		
3219	split lnL		Rel.Prob
3220	[D D]	-177.2	1
3221			
3222	At node N113:		
3223	split lnL		Rel.Prob
3224	[G E]	-177.2	1

3225  
3226 At node N107:  
3227 split lnL Rel.Prob  
3228 [G|G] -177.2 1  
3229  
3230 At node N105:  
3231 split lnL Rel.Prob  
3232 [G|G] -177.2 1  
3233  
3234 At node N112:  
3235 split lnL Rel.Prob  
3236 [E|E] -177.2 1  
3237  
3238 At node N120:  
3239 split lnL Rel.Prob  
3240 [B|B] -177.2 1  
3241  
3242  
3243 At node N148:  
3244 split lnL Rel.Prob  
3245 [B|B] -177.2 1  
3246  
3247 At node N186:  
3248 split lnL Rel.Prob  
3249 [B|B] -177.2 1  
3250  
3251 At node N257:  
3252 split lnL Rel.Prob  
3253 [A|A] -177.3 0.9615  
3254  
3255 At node N413:  
3256 split lnL Rel.Prob  
3257 [A|A] -177.2 1  
3258  
3259 At node N355:  
3260 split lnL Rel.Prob  
3261 [A|A] -177.2 1  
3262  
3263 At node N327:  
3264 split lnL Rel.Prob  
3265 [A|A] -177.2 1  
3266  
3267 At node N354:  
3268 split lnL Rel.Prob  
3269 [A|A] -177.2 1  
3270  
3271 At node N412:  
3272 split lnL Rel.Prob  
3273 [A|A] -177.2 1  
3274  
3275 At node N406:  
3276 split lnL Rel.Prob  
3277 [A|A] -177.2 1  
3278  
3279 At node N411:  
3280 split lnL Rel.Prob  
3281 [A|A] -177.2 1  
3282  
3283 At node N567:  
3284 split lnL Rel.Prob  
3285 [A|A] -177.2 1  
3286  
3287 At node N721:  
3288 split lnL Rel.Prob

3289 [C|A] -177.2 1  
 3290  
 3291 At node N669:  
 3292 split lnL Rel.Prob  
 3293 [C|C] -177.2 1  
 3294  
 3295 At node N720:  
 3296 split lnL Rel.Prob  
 3297 [A|A] -177.3 0.9151  
 3298  
 3299 At node N718:  
 3300 split lnL Rel.Prob  
 3301 [A|A] -177.3 0.9574  
 3302  
 3303 At node N688:  
 3304 split lnL Rel.Prob  
 3305 [A|A] -177.3 0.911  
 3306  
 3307  
 3308 At node N686:  
 3309 split lnL Rel.Prob  
 3310 [AB|B] -177.2 0.977  
 3311  
 3312 At node N684:  
 3313 split lnL Rel.Prob  
 3314 [B|A] -177.5 0.7617  
 3315 [AB|A] -178.7 0.223  
 3316  
 3317 At node N682:  
 3318 split lnL Rel.Prob  
 3319 [B|B] -177.5 0.759  
 3320 [AB|B] -178.6 0.2395  
 3321  
 3322 At node N680:  
 3323 split lnL Rel.Prob  
 3324 [B|B] -177.5 0.7514  
 3325 [B|AB] -178.6 0.2477  
 3326  
 3327 At node N674:  
 3328 split lnL Rel.Prob  
 3329 [B|B] -177.2 1  
 3330  
 3331 At node N672:  
 3332 split lnL Rel.Prob  
 3333 [B|B] -177.2 1  
 3334  
 3335 At node N679:  
 3336 split lnL Rel.Prob  
 3337 [B|B] -177.5 0.7236  
 3338 [AB|B] -178.5 0.2719  
 3339  
 3340 At node N677:  
 3341 split lnL Rel.Prob  
 3342 [AB|B] -178.7 0.2169  
 3343 [B|AB] -178.7 0.2169  
 3344 [AB|A] -178.7 0.2169  
 3345 [A|AB] -178.7 0.2169  
 3346 [B|B] -179.3 0.1308  
 3347  
 3348 At node N717:  
 3349 split lnL Rel.Prob  
 3350 [A|A] -177.2 1  
 3351  
 3352 At node N691:

```
3353 split lnL Rel.Prob
3354 [A|A] -177.2 1
3355
3356 At node N716:
3357 split lnL Rel.Prob
3358 [A|A] -177.2 1
3359
3360
3361
3362
3363
3364
```

3365  
 3366  
 3367 **Appendix S2.B.** Results of likelihood-based biogeographic analyses (using  
 3368 LAGRANGE), treating South America as multiple regions. A = Northern  
 3369 Hemisphere regions (Middle America, West Indies, North America, Europe, and  
 3370 Asia); B = Amazonian region; C = Atlantic Rainforest; D = Cerrado (and similar  
 3371 biomes); E = Chocoan region; F = Andean region; G = Guyana highlands.

3372  
 3373 lagrange: likelihood analysis of geographic range evolution  
 3374 Version 2 released February 2008  
 3375 This is development snapshot 20090327  
 3376 Authors: Richard Ree <rree@fieldmuseum.org>  
 3377 Stephen Smith <sasmith@nescent.org>  
 3378 <http://lagrange.googlecode.com>  
 3379

3380 Newick tree with interior nodes labeled:

3381  
 3382 (((((((Acris\_crepitans:18.01,Acris\_gryllus:18.01)N2:26.39,((((Pseudacris  
 3383 s\_bracyphona:4.67,Pseudacris\_brimleyi:4.67)N5:6.17,(((Pseudacris\_clarki  
 3384 i:0.48,Pseudacris\_triseriata\_KS:0.48)N8:0.49,Pseudacris\_maculata\_ON:0.97  
 3385 )N10:1.46,Pseudacris\_triseriata\_MI:2.44)N12:5.4,(Pseudacris\_feriarum\_KY  
 3386 :2.9,Pseudacris\_kalmi:2.9)N15:3.92,(Pseudacris\_foquettei:4.59,Pseudacris  
 3387 nigrita:4.59)N18:2.23)N19:1.02)N20:3.0)N21:14.55,(Pseudacris\_illinoens  
 3388 is:4.93,Pseudacris\_streckeri:4.93)N24:8.02,Pseudacris\_ornata:12.94)N26:1  
 3389 2.44)N27:4.61,(Pseudacris\_crucifer:18.08,Pseudacris\_ocularis:18.08)N30:1  
 3390 1.92)N31:4.52,(Pseudacris\_cadaverina:15.4,Pseudacris\_regilla:15.4)N34:19  
 3391 .12)N35:9.88)N36:10.82,((((((((Anotheca\_spinosa:18.49,Tripriion\_petasatu  
 3392 s:18.49)N39:5.87,Diaglena\_spatulata:24.36)N41:8.28,(Smilisca\_baudinii:26  
 3393 .69,((Smilisca\_cyanosticta:14.16,(Smilisca\_phaeota:10.51,Smilisca\_puma:1  
 3394 0.51)N46:3.65)N47:5.39,Smilisca\_fodiens:19.55)N49:3.57,(Smilisca\_sila:15  
 3395 .58,Smilisca\_sordida:15.58)N52:7.54)N53:3.57)N54:5.95)N55:5.83,(Isthmoh  
 3396 yla\_pseudopuma:21.31,Isthmohyla\_zeteki:21.31)N58:11.98,(Isthmohyla\_rivul  
 3397 aris:7.04,Isthmohyla\_tica:7.04)N61:26.25)N62:5.18)N63:2.38,(Tlalocohyla  
 3398 \_godmani:18.51,Tlalocohyla\_loquax:18.51)N66:9.84,(Tlalocohyla\_picta:23.0  
 3399 ,Tlalocohyla\_smithi:23.0)N69:5.34)N70:12.5)N71:1.2,(((Hyla\_andersoni:18  
 3400 .84,((Hyla\_avivoca:1.79,Hyla\_chrysoceles:1.79)N75:0.49,Hyla\_versicolor:  
 3401 2.28)N77:14.45,Hyla\_femoralis:16.72)N79:2.12)N80:2.54,(Hyla\_arenicolor:  
 3402 13.41,(((Hyla\_euphorbiacea:1.13,Hyla\_plicata:1.13)N84:2.85,Hyla\_eximia:  
 3403 3.98)N86:0.52,Hyla\_walkerii:4.49)N88:0.99,Hyla\_wrightorum:5.48)N90:7.93)N  
 3404 91:6.03,(Hyla\_immaculata:16.04,Hyla\_japonica:16.04)N94:3.4)N95:1.94)N96:  
 3405 5.85,(Hyla\_cinerea:13.38,Hyla\_gratiosa:13.38)N99:9.24,Hyla\_squirella:22  
 3406 .61)N101:4.62)N102:10.84,(((Hyla\_annectans:6.1,Hyla\_tsinlingensis:6.1)N1  
 3407 05:7.82,Hyla\_chinensis:13.91)N107:12.66,(Hyla\_arborea:12.19,Hyla\_sauvig  
 3408 nyii:12.19)N110:9.93,Hyla\_meridionalis:22.11)N112:4.46)N113:11.5)N114:3.  
 3409 97)N115:6.2,((Charadrahyla\_nephila:21.8,Charadrahyla\_taeiniopus:21.8)N118  
 3410 :17.54,Megastomatohyla\_mixe:39.34)N120:8.91)N121:2.32,(((Bromelioshyla\_b  
 3411 romeliacia:28.21,(Duellmanohyla\_rufiocolis:18.48,Duellmanohyla\_uranochr  
 3412 oa:18.48)N125:6.13,Duellmanohyla\_soralia:24.62)N127:3.59)N128:3.31,(Pty  
 3413 chohyla\_dendrophasma:21.52,(Ptychohyla\_euthysanota:11.05,(Ptychohyla\_l  
 3414 eonhardtschulzei:2.98,Ptychohyla\_zophodes:2.98)N133:6.17,Ptychohyla\_sp:9  
 3415 .15)N135:1.9)N136:2.31,Ptychohyla\_hypomycter:13.36)N138:8.16)N139:5.0,Pt  
 3416 ychohyla\_spinipollex:26.52)N141:5.0)N142:9.02,Ecnomiophyla\_miotympanum:40  
 3417 .54)N144:2.88,(Ecnomiophyla\_miliara:1.07,Ecnomiophyla\_minera:1.07)N147:42.  
 3418 35)N148:7.15)N149:1.71,(((Exerodonta\_abdivita:0.59,Exerodonta\_perkinsi:

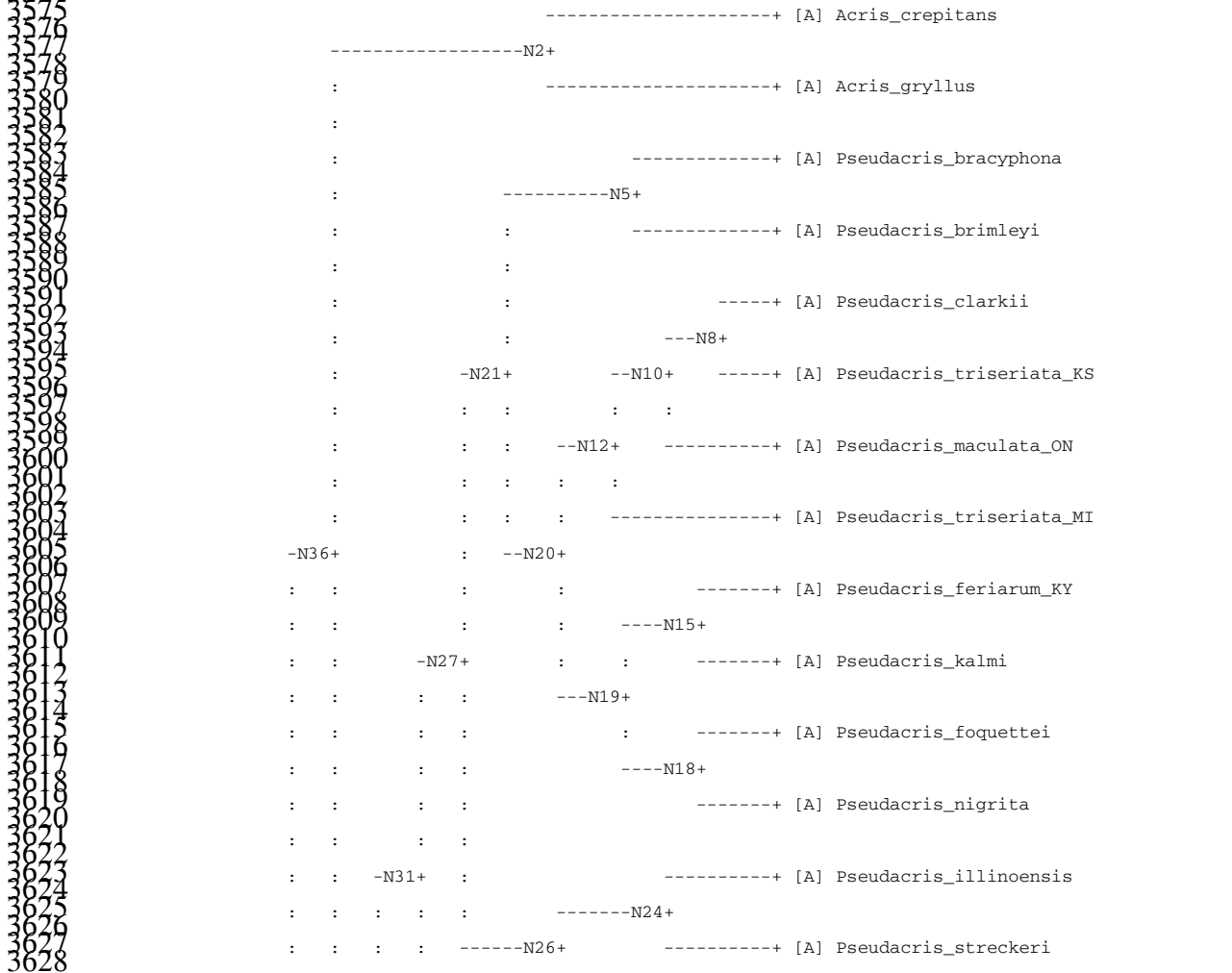
3419 0.59)N152:10.29, Exerodonta\_melanoma:10.88)N154:6.7, (((Exerodonta\_chimala  
 3420 pa:0.78, Exerodonta\_smaragdina:0.78)N157:1.16, Exerodonta\_xera:1.94)N159:1  
 3421 3.48, Exerodonta\_sumichrasti:15.42)N161:2.16)N162:16.04, (((Plectrohyla\_am  
 3422 eibothalamae:13.05, (Plectrohyla\_bistincta:7.2, (Plectrohyla\_calthula:1.97  
 3423 , Plectrohyla\_pentheter:1.97)N167:5.23)N168:5.85)N169:2.54, (((Plectrohyla  
 3424 \_arborescandens:4.15, Plectrohyla\_cyclada:4.15)N172:3.66, Plectrohyla\_siop  
 3425 ela:7.81)N174:5.38, Plectrohyla\_aff\_thorectes\_sp5:13.2)N176:2.39)N177:4.7  
 3426 , (Plectrohyla\_chrysopleura:15.03, (Plectrohyla\_glandulosa:11.85, (Plectroh  
 3427 yla\_guatemalensis:9.79, Plectrohyla\_matudai:9.79)N182:2.06)N183:3.18)N184  
 3428 :5.26)N185:13.33)N186:18.67)N187:2.94)N188:14.33, (((Aparasphenodon\_brun  
 3429 oi:22.79, (Argenteohyla\_siemersi:19.27, Nyctimantis\_rugiceps:19.27)N192:3.  
 3430 52)N193:12.64, Corythomantis\_greenengi:35.43)N195:5.45, ((Trachycephalus\_c  
 3431 oriacea:18.07, (((Trachycephalus\_hadroceps:6.51, Trachycephalus\_resinifri  
 3432 ctix:6.51)N199:0.85, Trachycephalus\_venulosus:7.35)N201:5.14, (Trachyceph  
 3433 alus\_imitatrix:9.38, Trachycephalus\_nigromaculatus:9.38)N204:3.11)N205:2.1  
 3434 , Trachycephalus\_mesophaea:14.6)N207:3.47)N208:5.68, Trachycephalus\_jordan  
 3435 i:23.75)N210:17.14)N211:4.03, ((Itapotihyla\_langsdorffii:39.01, Phyllodyte  
 3436 s\_auratus:39.01)N214:4.49, (((((Osteocephalus\_alboguttatus:14.92, (Osteoc  
 3437 ephalus\_leprieurii:4.63, Osteocephalus\_planiceps:4.63)N218:8.45, (Osteocep  
 3438 halus\_oophagus:5.62, Osteocephalus\_taurinus:5.62)N221:7.46)N222:1.84)N223  
 3439 :1.22, ((Osteocephalus\_buckleyi:4.17, Osteocephalus\_verruciger:4.17)N226:  
 3440 2.77, Osteocephalus\_cabrerae:6.94)N228:4.59, Osteocephalus\_mutabor:11.53)N  
 3441 230:4.61)N231:13.88, (Tepuihyla\_edelcaei:6.04, Tepuihyla\_sp:6.04)N234:23.97  
 3442 )N235:9.26, (((((Osteopilus\_brunneus:14.48, Osteopilus\_crucialis:14.48)N23  
 3443 8:10.45, (Osteopilus\_mariana:22.15, Osteopilus\_wilderi:22.15)N241:2.79)N2  
 3444 42:2.87, (Osteopilus\_dominicensis:24.2, Osteopilus\_pulchrilineatus:24.2)N2  
 3445 45:3.61)N246:1.65, Osteopilus\_vastus:29.46)N248:1.55, Osteopilus\_septentri  
 3446 onalis:31.02)N250:8.26)N251:2.81, (Phyllodytes\_luteolus:16.26, Phyllodytes  
 3447 \_sp:16.26)N254:25.82)N255:1.41)N256:1.42)N257:24.64)N258:4.2, (((Dendrop  
 3448 sopus\_allenorum:49.87, (((Dendropsopus\_anceps:38.43, (((Dendropsopus  
 3449 \_berthalutzae:15.59, Dendropsopus\_minusculus:15.59)N263:6.63, ((Dendrops  
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 3451 eroi:12.83)N268:7.45, (Dendropsopus\_rubicundula:10.82, Dendropsopus\_sanb  
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 3558 s\_lemur:37.74)N686:7.74,Phrynomedusa\_marginata:45.48)N688:2.24, ((Phasmah  
 3559 yla\_cochranae:13.67,Phasmahyla\_guttata:13.67)N691:29.65, (((Phyllomedusa  
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 3565 yllomedusa\_tarsius:3.98,Phyllomedusa\_trinitatus:3.98)N711:13.86,Phyllome  
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 3567 )N718:5.77,Cruziophyla\_calcarifer:53.49)N720:18.42)N721:11.06)N722;  
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Cladogram (branch lengths not to scale):



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: : : : -----+ [A] Pseudacris_ornata
: -N35+ :
: : : -----+ [A] Pseudacris_crucifer
: : -----N30+
: : -----+ [A] Pseudacris_ocularis
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: : -----+ [A] Pseudacris_cadaverina
: -----N34+
: -----+ [A] Pseudacris_regilla
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: -----+ [A] Anotheca_spinosa
: -----N39+
: -----N41+ -----+ [A] Triprion_petasatus
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: : -----+ [A] Diaglena_spatulata
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: N55+ -----+ [A] Smilisca_baudinii
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: : : : -----+ [A] Smilisca_cyanosticta
: : : : -N47+
: : -N54+ : : ----+ [AE] Smilisca_phaeota
: : : -N49+ -N46+
: : : : : ----+ [A] Smilisca_puma
: : : : :
: N63+ -N53+ -----+ [A] Smilisca_fodiens
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: : : : -----+ [AE] Smilisca_sila
: : : -----N52+
: : : -----+ [A] Smilisca_sordida
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: : : -----+ [A] Isthmohyla_pseudopuma
: : : -----N58+
: N71+ : : -----+ [A] Isthmohyla_zeteki
: : : -----N62+
: : : : -----+ [A] Isthmohyla_rivularis
: : : -----N61+
: : : -----+ [A] Isthmohyla_tica
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: : : -----+ [A] Tlalocohyla_godmani
: : : -----N66+
: : : : -----+ [A] Tlalocohyla_loquax
: : : -----N70+
: : : : -----+ [A] Tlalocohyla_picta
: : : -----N69+
: : : -----+ [A] Tlalocohyla_smithi

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:      :      -----+ [A] Hyla_andersoni
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:      :      :      -----+ [A] Hyla_avivoca
:      :      -N80+      --N75+
:      :      :      --N77+      -----+ [A] Hyla_chrysocelis
N188+  :      :      :      :
:      :      :      --N79+      -----+ [A] Hyla_versicolor
:      :      N115+      :      :
:      :      :      :      -----+ [A] Hyla_femoralis
:      :      :      :
:      :      :      N96+      -----+ [A] Hyla_arenicolor
:      :      :      :      :
:      :      :      :      :      -----+ [A] Hyla_euphorbiacea
:      :      :      :      :      -N84+
:      :      :      :      :      N91+      -N86+      -----+ [A] Hyla_plicata
:      :      :      :      :      :      :
:      :      :      :      :      :      N88+      -----+ [A] Hyla_eximia
:      :      :      :      :      :      :
:      :      :      :      N95+      N90+      -----+ [A] Hyla_walkerii
:      :      :      :      N102+      :
:      :      :      :      :      :      -----+ [A] Hyla_wrightorum
:      :      :      :      :
:      :      :      :      :      :      -----+ [A] Hyla_immaculata
:      :      :      :      :      -----N94+
:      :      :      :      :      :      -----+ [A] Hyla_japonica
:      :      :      :      :
:      :      :      :      :      :      -----+ [A] Hyla_cinerea
:      :      :      :      :      :      -----N99+
:      :      :      :      N114+      -----N101+      -----+ [A] Hyla_gratiosa
:      :      :      :      :
:      :      :      :      :      :      -----+ [A] Hyla_squirella
:      :      :      :      :
:      :      :      :      :      :      -----+ [A] Hyla_annectans
:      :      :      :      :      -----N105+
:      :      :      :      :      ---N107+      -----+ [A] Hyla_tsinlingensis
:      :      :      :      :      :
:      :      :      :      :      :      -----+ [A] Hyla_chinensis
:      :      :      :      ---N113+
:      :      :      :      :      :      -----+ [A] Hyla_arborea
:      :      :      :      :      -----N110+
:      :      :      :      ---N112+      -----+ [A] Hyla_sauvignyii
:      :      :      :
:      :      :      :      :      -----+ [A] Hyla_meridionalis
:      :      :      :
:      :      :      :      :      -----+ [A] Charadrahyla_nephila

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: : : : -----+ [A] Plectrohyla_ameibothalamae
: : --N186+ --N169+
: : : : : -----+ [A] Plectrohyla_bistincta
: : : : : --N168+
: : : : : : -----+ [A] Plectrohyla_calthula
: : : : : --N167+
: : : --N177+ -----+ [A] Plectrohyla_pentheter
: : : : :
: : : : : -----+ [A] Plectrohyla_arborescandens
: : : : : --N172+
: : : : : --N174+ -----+ [A] Plectrohyla_cyclada
: : --N185+ : : :
: : : --N176+ -----+ [A] Plectrohyla_siopela
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: : : -----+ [A] Plectrohyla_aff_thorectes_sp5
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: : : -----+ [A] Plectrohyla_chrysopleura
: : --N184+
: : : -----+ [A] Plectrohyla_glandulosa
: : --N183+
: : : -----+ [A] Plectrohyla_guatemalensis
: : --N182+
: : -----+ [A] Plectrohyla_matudai
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: : -----+ [C] Aparasphenodon_brunoi
: : -----N193+
: : : : -----+ [D] Argenteohyla_siemersi
: : -----N195+ -----N192+
: : : : -----+ [B] Nyctimantis_rugiceps
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: : : -----+ [D] Corythomantis_greenengi
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: : : -----+ [B] Trachycephalus_coriacea
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: : -N211+ : -----+ [B] Trachycephalus_hadroceps
: : : : : --N199+
: : : -N208+ --N201+ -----+ [B] Trachycephalus_resinifrictix
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: : : : : --N205+ -----+ [ABDE] Trachycephalus_venulosus
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: : : : : : : -----+ [C] Trachycephalus_imitatrix
: : : -N210+ -N207+ -----N204+
: : : : : -----+ [C] Trachycephalus_nigromaculatus
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: : : : -----+ [C] Trachycephalus_mesophaea

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3998      : : : :
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4006      : : :
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4049      : : : : -----N234+
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4051      : : : : -----+ [G] Tepuihyla_sp
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4077      : : : -N250+ :
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4372 : : : : : : : : -----+ [D] Scinax_uruguayana
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4374 : : : : : : : : -----+ [CD] Scinax_berthae
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: : : : : : : -----+ [F] Hyloscirtus_tapichalaca
: : : : : : -----N551+
: : : : : : -----+ [F] Hyloscirtus_pachus
: : : : : : -----N550+
: : : : -----N562+ -----+ [F] Hyloscirtus_pantostictus
: : : : :
: N567+ : : -----+ [A] Hyloscirtus_colymbus
: : : : : -----N555+
: : : : : -----N557+ -----+ [F] Hyloscirtus_simmonsii
: : : : : : :
: : : : -----N561+ -----+ [BF] Hyloscirtus_phyllognathus
: : : : :
: : : : -----+ [F] Hyloscirtus_lascinius
: : : : -----N560+
: : : : -----+ [AEF] Hyloscirtus_palmeri
: : : : -----+ [G] Myersiohyla_inparquesi
: : : : -----+ [G] Myersiohyla_kanaima
: : : :
: : : : -----+ [H] Cyclorana_alboguttata
: : : : ---N571+
: : : : : -----+ [H] Cyclorana_cryptotis
: : : : -N577+
: : : : : -----+ [H] Cyclorana_breviceps
: : : : : -N576+
: : : : -N579+ : -----+ [H] Cyclorana_longipes
: : : : : -N575+
: : : : : -----+ [H] Cyclorana_manya

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4733 : -N583+ :
4734 : : : -----+ [H] Cyclorana_australis
4735 : : :
4736 : : :
4737 : : :
4738 : : :
4739 : : : -----+ [H] Litoria_dahliei
4740 : : :
4741 : N587+ -----N582+
4742 : : :
4743 : : : -----+ [H] Litoria_eucnemis
4744 : : :
4745 : : :
4746 : : :
4747 : : : -----+ [H] Litoria_aurea
4748 : : :
4749 : : -----N586+
4750 : : :
4751 : N595+ -----+ [H] Litoria_cyclorhyncha
4752 : : :
4753 : : :
4754 : : :
4755 : : : -----+ [H] Litoria_genimaculata
4756 : : :
4757 : : : -----N590+
4758 : : :
4759 : : : -----+ [H] Litoria_lesueurii
4760 : : :
4761 : : -----N594+
4762 : : :
4763 : : :
4764 : : : -----+ [H] Litoria_nannotis
4765 : : :
4766 : N607+ -----N593+
4767 : : :
4768 : : : -----+ [H] Nyctimystes_dayi
4769 : : :
4770 : : :
4771 : : : -----+ [H] Litoria_caerulea
4772 : : :
4773 : : : -----N600+
4774 : : :
4775 : : : : : -----+ [H] Litoria_gilleni
4776 : : : : :
4777 : : : : -----N599+
4778 : : :
4779 : : -----N606+ -----+ [H] Litoria_splendida
4780 : : :
4781 : N611+ :
4782 : : : :
4783 : : : :
4784 : : : : -----N603+
4785 : : : :
4786 : : : -----N605+ -----+ [H] Litoria_xanthomera
4787 : : : :
4788 : : : :
4789 : : : : -----+ [H] Litoria_gracilentia
4790 : : : :
4791 : : : :
4792 : : : : -----+ [H] Litoria_phyllochroa
4793 : : : :
4794 : N627+ -----N610+
4795 : : : : -----+ [H] Litoria_subglandulos
4796 : : : :
4797 : : : :
4798 : : : : -----+ [H] Litoria_infrafrenata
4799 : : : :
4800 : : : :
4801 : : : : -----+ [H] Nyctimystes_cheesmani
4802 : : : :
4803 : : : : --N619+
4804 : : : : : : -----+ [H] Nyctimystes_foricula
4805 : : : : : :
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:      :      :      -----N622+
:      :      :      -----+ [H] Nyctimystes_narinosus
:      :      :
:      :      :      -----+ [H] Nyctimystes_papua
:      :      :
:      :      :      -----+ [H] Litoriaamboinensis
:      :      :      -----N630+
:      :      :      -----N632+      -----+ [H] Litoria_peronii
:      :      :      :      :
:      N669+      -----N636+      -----+ [H] Litoria_rothii
:      :      :      :      :
:      :      :      :      :      -----+ [H] Litoria_dentata
:      :      :      :      :      -----N635+
:      :      :      :      :      -----+ [H] Litoria_rubella
:      :      :      :      :
:      :      :      :      :      -----+ [H] Litoria_arfakiana
:      :      :      :      :      ---N639+
:      :      :      :      :      ---N641+      -----+ [H] Litoria_thesauensis
:      :      :      :      :      :      :
:      :      :      -N666+      :      -----+ [H] Litoria_modica
:      :      :      :      :      ---N647+
:      :      :      :      :      :      :      -----+ [H] Litoria_bicolor
:      :      :      :      :      :      :      ---N644+
:      :      :      :      :      --N649+      --N646+      -----+ [H] Litoria_booroolongensis
:      :      :      :      :      :      :      :
:      :      :      :      :      :      :      -----+ [H] Litoria_fallax
:      :      :      :      :      :      :
:      :      :      :      :      :      :      -----+ [H] Litoria_microbelos
:      :      :      :      :      :
:      :      :      :      :      :      :      -----+ [H] Litoria_coplandi
:      :      :      :      -N665+      ----N654+
:      :      :      :      :      :      :      -----+ [H] Litoria_watjulumensis
:      :      -N668+      :      :      ----N653+
:      :      :      :      :      :      :      -----+ [H] Litoria_wollastoni
:      :      :      :      :      -N662+
:      :      :      :      :      :      :      -----+ [H] Litoria_frecyneti
:      :      :      :      :      :      :      --N659+
:      :      :      :      :      :      :      :      -----+ [H] Litoria_inermis
:      :      :      :      -N664+      --N661+      --N658+
:      :      :      :      :      :      :      -----+ [H] Litoria_pallida
:      :      :      :      :      :
:      :      :      :      :      :      -----+ [H] Litoria_nasuta
:      :      :      :      :
:      :      :      :      :      -----+ [H] Litoria_tornieri
:      :      :
N721+      -----+ [H] Litoria_meiriana

```

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4017 :
4018 :
4019 :
4020 : -----+ [A] Agalychnis_anna
4021 :
4022 : --N672+
4023 :
4024 : --N674+ -----+ [A] Agalychnis_moreleti
4025 :
4026 : : :
4027 : : -----+ [A] Agalychnis_callidryas
4028 :
4029 : --N680+
4030 :
4031 : : : -----+ [AE] Agalychnis_litodryas
4032 :
4033 : : : --N677+
4034 :
4035 : -N682+ --N679+ -----+ [AE] Agalychnis_spurrelli
4036 :
4037 : : : :
4038 :
4039 : : : -----+ [A] Agalychnis_saltator
4040 :
4041 : -N684+ :
4042 :
4043 : : : -----+ [A] Pachymedusa_dacnicolor
4044 :
4045 : -N686+ :
4046 :
4047 : : : -----+ [C] Hylomantis_granulosa
4048 :
4049 : -N688+ :
4050 :
4051 : : : -----+ [A] Hylomantis_lemur
4052 :
4053 : : :
4054 :
4055 : : -----+ [C] Phrynomedusa_marginata
4056 :
4057 : :
4058 :
4059 : : -----+ [C] Phasmahyla_cochranae
4060 :
4061 : -N718+ -----N691+
4062 :
4063 : : : : -----+ [C] Phasmahyla_guttata
4064 :
4065 : : : :
4066 :
4067 : : : : -----+ [B] Phyllomedusa_atelopoides
4068 :
4069 : : : : ---N696+
4070 :
4071 : : : : : : -----+ [F] Phyllomedusa_duellmani
4072 :
4073 : : : : : --N695+
4074 :
4075 : : -N717+ --N702+ -----+ [F] Phyllomedusa_perinesos
4076 :
4077 : : : : :
4078 :
4079 : : : : : -----+ [D] Phyllomedusa_azurea
4080 :
4081 : : : : : --N699+
4082 :
4083 : : : -N704+ ---N701+ -----+ [BDF] Phyllomedusa_hypochondrialis
4084 :
4085 : : : : :
4086 :
4087 : : : : : -----+ [B] Phyllomedusa_palliata
4088 :
4089 : -N720+ : :
4090 :
4091 : : -N716+ -----+ [B] Phyllomedusa_tomopterna
4092 :
4093 : :
4094 :
4095 : : -----+ [B] Phyllomedusa_bicolor
4096 :
4097 : : -----N707+
4098 :
4099 : : : -----+ [B] Phyllomedusa_vaillanti
4100 :
4101 : --N715+
4102 :
4103 : : -----+ [BDF] Phyllomedusa_boliviana
4104 :
4105 : : :
4106 :
4107 : : -N714+ -----+ [B] Phyllomedusa_tarsius
4108 :

```



```

5009 : : --N711+
5010 : :
5011 : ---N713+ -----+ [BF] Phyllomedusa_trinitatus
5012 : :
5013 : :
5014 : :
5015 : -----+ [CD] Phyllomedusa_tetraploidea
5016 : :
5017 : :
5018 : :
5019 : :
5020 : -----+ [AE] Cruziohyala_calcarifer
5021 : :
5022 : :

```

```

5023 Global ML at root node:
5024 -lnL = 650.1
5025 dispersal = 0.005045
5026 extinction = 0.002908
5027

```

```

5028 Ancestral range subdivision/inheritance scenarios ('splits') at
5029 internal nodes.
5030

```

```

5031 * Split format: [left|right], where 'left' and 'right' are the ranges
5032 inherited by each descendant branch (on the printed tree, 'left' is
5033 the upper branch, and 'right' the lower branch).
5034

```

```

5035 * Only splits within 2 log-likelihood units of the maximum for each
5036 node are shown. 'Rel.Prob' is the relative probability (fraction of
5037 the global likelihood) of a split.
5038

```

```

5039
5040 At node N722:
5041 split lnL Rel.Prob
5042 [ABDE|E] -651.9 0.1576
5043 [ABE|E] -652.1 0.1293
5044 [B|BH] -652.3 0.111
5045 [ABDE|B] -652.6 0.081
5046 [ABE|B] -652.8 0.06647
5047 [BE|E] -653.1 0.05037
5048 [BFG|B] -653.1 0.04737
5049 [BDE|E] -653.2 0.04292
5050 [B|B] -653.3 0.04279
5051 [BG|B] -653.5 0.03367
5052 [BE|B] -653.8 0.02589
5053 [BDE|B] -653.9 0.02206
5054 [B|H] -653.9 0.02154
5055

```

```

5056 At node N568:
5057 split lnL Rel.Prob
5058 [ABDE|B] -651.3 0.2883
5059 [ABE|B] -651.6 0.2123
5060 [B|B] -651.9 0.1699
5061 [BE|B] -652.5 0.09427
5062 [B|BFG] -652.9 0.05899
5063 [BDE|B] -653.1 0.0521
5064 [B|BG] -653.1 0.05038
5065

```

```

5066 At node N414:
5067 split lnL Rel.Prob
5068 [ABDE|B] -651.4 0.262
5069 [ABE|B] -651.6 0.2179
5070 [B|B] -651.7 0.1985
5071 [BE|B] -652.3 0.108
5072 [AE|B] -653 0.05351
5073 [ABDE|D] -653.1 0.04815
5074

```

```

5075 At node N258:
5076 split lnL Rel.Prob
5077 [A|ABE] -651.5 0.2403

```

5078	[A BE]	-651.6	0.2304
5079	[A ABDE]	-651.7	0.2099
5080	[A BDE]	-651.7	0.2016
5081	[A AE]	-653.3	0.0398
5082			
5083	At node N188:		
5084	split lnL	Rel.Prob	
5085	[A A]	-650.1	0.9843
5086			
5087	At node N257:		
5088	split lnL	Rel.Prob	
5089	[B ABDE]	-651.7	0.202
5090	[BDE B]	-651.7	0.1969
5091	[D ABDE]	-652	0.146
5092	[B ABE]	-652.1	0.1338
5093	[D ABE]	-652.6	0.08099
5094	[BE B]	-653	0.05434
5095	[B BE]	-653.2	0.04402
5096			
5097	At node N211:		
5098	split lnL	Rel.Prob	
5099	[B B]	-651.2	0.3225
5100	[D D]	-651.8	0.1819
5101	[D BE]	-652	0.1489
5102	[D BDE]	-652.8	0.06937
5103	[BD B]	-652.8	0.06769
5104	[B BE]	-653.1	0.05201
5105			
5106	At node N195:		
5107	split lnL	Rel.Prob	
5108	[D D]	-650.9	0.4395
5109	[BD D]	-651.2	0.3184
5110	[BCD D]	-652.6	0.08251
5111			
5112	At node N193:		
5113	split lnL	Rel.Prob	
5114	[C BD]	-650.6	0.6298
5115	[C D]	-652.1	0.1336
5116			
5117	At node N192:		
5118	split lnL	Rel.Prob	
5119	[D B]	-650.4	0.7335
5120			
5121	At node N210:		
5122	split lnL	Rel.Prob	
5123	[B E]	-650.7	0.5265
5124	[BD E]	-651.4	0.2806
5125	[B B]	-652.7	0.07777
5126			
5127			
5128	At node N208:		
5129	split lnL	Rel.Prob	
5130	[B B]	-650.8	0.4956
5131	[B BC]	-651.9	0.1603
5132	[B BD]	-652	0.1449
5133	[B BCD]	-652.1	0.136
5134			
5135	At node N207:		
5136	split lnL	Rel.Prob	
5137	[BC C]	-650.6	0.6256
5138	[BCD C]	-651.3	0.3123
5139			
5140	At node N205:		
5141	split lnL	Rel.Prob	

5142 [B|C] -650.6 0.5865  
 5143 [BD|C] -651.1 0.3641  
 5144  
 5145 At node N201:  
 5146 split lnL Rel.Prob  
 5147 [B|B] -651 0.3909  
 5148 [B|BD] -651.6 0.2247  
 5149 [B|BDE] -651.7 0.1991  
 5150 [B|BE] -652.5 0.09446  
 5151 [B|ABDE] -652.9 0.06299  
 5152  
 5153 At node N199:  
 5154 split lnL Rel.Prob  
 5155 [B|B] -650.1 0.999  
 5156  
 5157 At node N204:  
 5158 split lnL Rel.Prob  
 5159 [C|C] -650.1 0.992  
 5160  
 5161 At node N256:  
 5162 split lnL Rel.Prob  
 5163 [B|B] -651.4 0.2809  
 5164 [B|ABE] -651.6 0.216  
 5165 [B|ABDE] -651.9 0.1573  
 5166 [D|ABE] -652.3 0.1092  
 5167 [D|ABDE] -652.4 0.1018  
 5168 [B|BE] -652.9 0.05997  
 5169  
 5170 At node N214:  
 5171 split lnL Rel.Prob  
 5172 [B|B] -650.9 0.4647  
 5173 [D|D] -652 0.1532  
 5174 [D|B] -652 0.1459  
 5175 [C|B] -652.3 0.1128  
 5176  
 5177 At node N255:  
 5178 split lnL Rel.Prob  
 5179 [ABE|B] -651.2 0.319  
 5180 [ABE|D] -651.5 0.2587  
 5181 [B|B] -651.6 0.2244  
 5182 [BE|B] -652.6 0.08069  
 5183 [B|C] -652.9 0.06124  
 5184  
 5185  
 5186 At node N251:  
 5187 split lnL Rel.Prob  
 5188 [B|AE] -650.9 0.4419  
 5189 [BE|A] -651.5 0.2537  
 5190 [B|E] -651.7 0.2011  
 5191  
 5192 At node N235:  
 5193 split lnL Rel.Prob  
 5194 [B|G] -650.7 0.5408  
 5195 [B|B] -651.3 0.2916  
 5196  
 5197 At node N231:  
 5198 split lnL Rel.Prob  
 5199 [B|B] -650.2 0.8631  
 5200  
 5201 At node N223:  
 5202 split lnL Rel.Prob  
 5203 [B|B] -650.1 0.9929  
 5204  
 5205 At node N222:

5206	split	lnL	Rel.Prob
5207	[B B]	-650.1	0.988
5208			
5209	At node N218:		
5210	split	lnL	Rel.Prob
5211	[B B]	-650.1	0.9986
5212			
5213	At node N221:		
5214	split	lnL	Rel.Prob
5215	[B B]	-650.3	0.8071
5216	[B BD]	-651.8	0.1857
5217			
5218	At node N230:		
5219	split	lnL	Rel.Prob
5220	[B B]	-650.4	0.7381
5221			
5222	At node N228:		
5223	split	lnL	Rel.Prob
5224	[B B]	-650.5	0.6378
5225	[BF B]	-651.3	0.3141
5226			
5227	At node N226:		
5228	split	lnL	Rel.Prob
5229	[BF F]	-650.2	0.9361
5230			
5231	At node N234:		
5232	split	lnL	Rel.Prob
5233	[G G]	-650.1	0.9515
5234			
5235	At node N250:		
5236	split	lnL	Rel.Prob
5237	[A A]	-650.5	0.6517
5238	[A AE]	-651.6	0.2289
5239			
5240	At node N248:		
5241	split	lnL	Rel.Prob
5242	[A A]	-650.2	0.9499
5243			
5244	At node N246:		
5245	split	lnL	Rel.Prob
5246	[A A]	-650.1	0.9954
5247			
5248	At node N242:		
5249	split	lnL	Rel.Prob
5250	[A A]	-650.1	0.9988
5251			
5252	At node N238:		
5253	split	lnL	Rel.Prob
5254	[A A]	-650.1	0.9992
5255			
5256	At node N241:		
5257	split	lnL	Rel.Prob
5258	[A A]	-650.1	0.9994
5259			
5260	At node N245:		
5261	split	lnL	Rel.Prob
5262	[A A]	-650.1	0.9978
5263			
5264	At node N254:		
5265	split	lnL	Rel.Prob
5266	[C C]	-650.4	0.7149
5267			
5268	At node N413:		
5269	split	lnL	Rel.Prob

5270	[B B]	-650.3	0.8434
5271			
5272	At node N355:		
5273	split lnL		Rel.Prob
5274	[B B]	-650.3	0.8324
5275			
5276	At node N327:		
5277	split lnL		Rel.Prob
5278	[B B]	-650.9	0.4497
5279	[B C]	-651.1	0.3632
5280			
5281	At node N325:		
5282	split lnL		Rel.Prob
5283	[B B]	-650.2	0.9077
5284			
5285	At node N324:		
5286	split lnL		Rel.Prob
5287	[B B]	-650.2	0.9047
5288			
5289	At node N318:		
5290	split lnL		Rel.Prob
5291	[B B]	-650.2	0.9283
5292			
5293	At node N304:		
5294	split lnL		Rel.Prob
5295	[B B]	-650.3	0.8357
5296			
5297	At node N288:		
5298	split lnL		Rel.Prob
5299	[B B]	-651	0.4163
5300	[C BC]	-651.3	0.3115
5301	[C B]	-651.9	0.1607
5302			
5303	At node N287:		
5304	split lnL		Rel.Prob
5305	[B B]	-650.7	0.5535
5306	[B BC]	-652	0.1556
5307	[BC B]	-652.6	0.08392
5308			
5309	At node N285:		
5310	split lnL		Rel.Prob
5311	[B B]	-650.5	0.6529
5312	[BC B]	-651.9	0.1664
5313			
5314	At node N275:		
5315	split lnL		Rel.Prob
5316	[BD D]	-651.2	0.3285
5317	[BC C]	-651.2	0.3246
5318	[BCD C]	-653	0.05511
5319	[BCD D]	-653.1	0.05185
5320	[B C]	-653.1	0.04777
5321			
5322	At node N273:		
5323	split lnL		Rel.Prob
5324	[B BD]	-651.3	0.3103
5325	[BC B]	-651.8	0.1799
5326	[B B]	-652.3	0.1111
5327	[BC D]	-652.6	0.08555
5328	[C BC]	-653	0.05641
5329	[B BC]	-653.1	0.05037
5330			
5331	At node N263:		
5332	split lnL		Rel.Prob
5333	[C B]	-650.4	0.7586

5334 [B|B] -652.3 0.1101  
5335  
5336 At node N272:  
5337 split lnL Rel.Prob  
5338 [B|D] -651.4 0.2833  
5339 [BD|D] -651.5 0.2479  
5340 [D|D] -652.1 0.1299  
5341 [B|C] -652.3 0.1098  
5342 [B|B] -652.6 0.08608  
5343 [B|CD] -652.6 0.07932  
5344  
5345 At node N268:  
5346 split lnL Rel.Prob  
5347 [B|B] -650.6 0.5773  
5348 [BD|B] -651 0.3892  
5349  
5350 At node N266:  
5351 split lnL Rel.Prob  
5352 [BD|B] -650.8 0.5111  
5353 [B|B] -650.9 0.4709  
5354  
5355 At node N271:  
5356 split lnL Rel.Prob  
5357 [D|D] -650.6 0.6102  
5358 [D|CD] -651.2 0.3173  
5359  
5360 At node N284:  
5361 split lnL Rel.Prob  
5362 [B|B] -650.7 0.5757  
5363 [B|BE] -651.4 0.2708  
5364 [B|ABE] -652.3 0.1131  
5365  
5366 At node N283:  
5367 split lnL Rel.Prob  
5368 [ABE|A] -650.5 0.6939  
5369 [BE|A] -651.5 0.2347  
5370  
5371 At node N279:  
5372 split lnL Rel.Prob  
5373 [ABE|B] -650.7 0.5741  
5374 [BE|B] -651.6 0.2153  
5375 [AE|B] -652.2 0.12  
5376  
5377 At node N282:  
5378 split lnL Rel.Prob  
5379 [A|A] -650.1 0.9918  
5380  
5381 At node N303:  
5382 split lnL Rel.Prob  
5383 [B|B] -650.2 0.9135  
5384  
5385 At node N299:  
5386 split lnL Rel.Prob  
5387 [B|B] -650.9 0.4473  
5388 [B|C] -651 0.4037  
5389  
5390 At node N297:  
5391 split lnL Rel.Prob  
5392 [B|E] -651.3 0.3038  
5393 [BF|F] -651.6 0.2274  
5394 [B|F] -651.9 0.1654  
5395 [B|B] -651.9 0.1624  
5396 [B|AE] -653.2 0.04687  
5397

5398 At node N295:  
5399 split lnL Rel.Prob  
5400 [B|B] -650.5 0.6986  
5401 [BF|B] -651.7 0.21  
5402  
5403 At node N291:  
5404 split lnL Rel.Prob  
5405 [B|B] -650.8 0.5079  
5406 [F|BF] -652.4 0.09787  
5407 [B|BF] -652.5 0.08658  
5408 [BF|B] -652.6 0.08588  
5409  
5410 At node N294:  
5411 split lnL Rel.Prob  
5412 [B|B] -650.3 0.8536  
5413 [BD|B] -652.2 0.1278  
5414  
5415 At node N302:  
5416 split lnL Rel.Prob  
5417 [B|B] -650.1 0.9872  
5418  
5419 At node N317:  
5420 split lnL Rel.Prob  
5421 [B|B] -650.3 0.8185  
5422  
5423 At node N309:  
5424 split lnL Rel.Prob  
5425 [B|B] -650.1 0.9636  
5426  
5427 At node N307:  
5428 split lnL Rel.Prob  
5429 [B|B] -650.2 0.9027  
5430  
5431 At node N316:  
5432 split lnL Rel.Prob  
5433 [B|B] -651.1 0.3625  
5434 [F|B] -651.1 0.3503  
5435 [B|C] -651.8 0.1901  
5436  
5437 At node N314:  
5438 split lnL Rel.Prob  
5439 [F|F] -650.3 0.8447  
5440  
5441 At node N313:  
5442 split lnL Rel.Prob  
5443 [F|F] -650.1 0.9912  
5444  
5445 At node N323:  
5446 split lnL Rel.Prob  
5447 [B|B] -650.6 0.6013  
5448 [F|B] -651.4 0.2824  
5449  
5450 At node N322:  
5451 split lnL Rel.Prob  
5452 [B|C] -650.4 0.7392  
5453 [B|B] -652.2 0.1243  
5454  
5455 At node N354:  
5456 split lnL Rel.Prob  
5457 [D|B] -651.1 0.3806  
5458 [B|B] -651.1 0.3802  
5459 [BD|B] -652.4 0.09688  
5460  
5461 At node N352:

5462	split	lnL	Rel.Prob
5463	[D D]	-650.7	0.5323
5464	[BD D]	-651	0.3888
5465			
5466	At node N336:		
5467	split	lnL	Rel.Prob
5468	[D D]	-650.7	0.5306
5469	[D BD]	-651.3	0.3118
5470			
5471	At node N332:		
5472	split	lnL	Rel.Prob
5473	[D D]	-650.3	0.8519
5474	[D BD]	-652.1	0.1384
5475			
5476	At node N331:		
5477	split	lnL	Rel.Prob
5478	[D D]	-650.5	0.702
5479	[D BD]	-651.3	0.2893
5480			
5481	At node N335:		
5482	split	lnL	Rel.Prob
5483	[BD D]	-651.5	0.236
5484	[D BD]	-651.5	0.236
5485	[BD B]	-651.5	0.2345
5486	[B BD]	-651.5	0.2345
5487	[D D]	-653.1	0.04747
5488			
5489	At node N351:		
5490	split	lnL	Rel.Prob
5491	[D D]	-650.1	0.9685
5492			
5493	At node N339:		
5494	split	lnL	Rel.Prob
5495	[D D]	-650.1	0.9983
5496			
5497	At node N350:		
5498	split	lnL	Rel.Prob
5499	[D D]	-650.1	0.9626
5500			
5501	At node N346:		
5502	split	lnL	Rel.Prob
5503	[D D]	-650.2	0.9164
5504			
5505	At node N345:		
5506	split	lnL	Rel.Prob
5507	[D D]	-650.4	0.7505
5508	[BD D]	-651.9	0.1685
5509			
5510	At node N343:		
5511	split	lnL	Rel.Prob
5512	[BDE D]	-651.2	0.3388
5513	[D D]	-651.2	0.3384
5514	[BD D]	-651.3	0.3134
5515			
5516	At node N349:		
5517	split	lnL	Rel.Prob
5518	[D D]	-650.1	0.9976
5519			
5520	At node N412:		
5521	split	lnL	Rel.Prob
5522	[B B]	-650.7	0.5545
5523	[D B]	-652.3	0.1088
5524	[C BC]	-652.7	0.07605
5525			



5526 At node N406:  
 5527 split lnL Rel.Prob  
 5528 [BD|D] -651.5 0.2536  
 5529 [D|D] -651.6 0.2156  
 5530 [BD|C] -652.6 0.0833  
 5531 [D|CD] -652.7 0.07323  
 5532 [B|B] -652.9 0.06076  
 5533 [D|C] -653 0.05523  
 5534 [B|C] -653.1 0.04935  
 5535 [BDE|D] -653.2 0.04307  
 5536 [D|BD] -653.4 0.03638  
 5537  
 5538 At node N400:  
 5539 split lnL Rel.Prob  
 5540 [D|D] -651 0.4068  
 5541 [BD|D] -651.1 0.3823  
 5542 [BDE|D] -652.5 0.09135  
 5543  
 5544 At node N398:  
 5545 split lnL Rel.Prob  
 5546 [D|D] -651.2 0.3431  
 5547 [BD|D] -651.5 0.2544  
 5548 [BDE|D] -652.2 0.126  
 5549 [B|BD] -652.9 0.05948  
 5550  
 5551 At node N366:  
 5552 split lnL Rel.Prob  
 5553 [D|BE] -650.8 0.5011  
 5554 [D|B] -651.4 0.2801  
 5555  
 5556 At node N365:  
 5557 split lnL Rel.Prob  
 5558 [BE|E] -650.5 0.6514  
 5559 [ABE|E] -652.3 0.115  
 5560 [B|E] -652.3 0.108  
 5561  
 5562 At node N363:  
 5563 split lnL Rel.Prob  
 5564 [BE|B] -650.6 0.5899  
 5565 [ABE|B] -651.8 0.1738  
 5566 [B|B] -652 0.155  
 5567  
 5568 At node N361:  
 5569 split lnL Rel.Prob  
 5570 [E|B] -650.8 0.4834  
 5571 [AE|B] -651.2 0.3388  
 5572  
 5573 At node N359:  
 5574 split lnL Rel.Prob  
 5575 [A|E] -650.2 0.8959  
 5576  
 5577 At node N397:  
 5578 split lnL Rel.Prob  
 5579 [D|D] -650.4 0.7097  
 5580 [BD|D] -652.1 0.142  
 5581  
 5582 At node N395:  
 5583 split lnL Rel.Prob  
 5584 [D|D] -650.9 0.4468  
 5585 [BD|D] -651.8 0.1753  
 5586 [B|D] -652.6 0.07812  
 5587 [BDE|D] -652.7 0.07097  
 5588  
 5589 At node N393:

5590	split	lnL	Rel.Prob
5591	[BD B]	-651	0.3965
5592	[B B]	-652.5	0.09002
5593	[BCD B]	-652.6	0.08277
5594	[BDE E]	-652.7	0.07665
5595	[BD E]	-652.7	0.07192
5596			
5597	At node N387:		
5598	split	lnL	Rel.Prob
5599	[B BD]	-651.7	0.2066
5600	[B B]	-652.1	0.1419
5601	[BD D]	-652.2	0.119
5602	[B BCD]	-652.4	0.1034
5603	[BDE D]	-652.7	0.07105
5604	[D D]	-653.1	0.04903
5605	[BD B]	-653.1	0.04807
5606	[ABDE D]	-653.3	0.03978
5607	[D BD]	-653.6	0.03161
5608	[B CD]	-653.6	0.02998
5609	[BDE B]	-653.6	0.02869
5610			
5611	At node N373:		
5612	split	lnL	Rel.Prob
5613	[B B]	-650.9	0.4337
5614	[B BD]	-651.5	0.2346
5615	[B BDE]	-652.2	0.1238
5616	[B ABDE]	-652.7	0.07472
5617			
5618	At node N372:		
5619	split	lnL	Rel.Prob
5620	[D BD]	-651	0.3866
5621	[D BDE]	-651.4	0.2858
5622	[D ABDE]	-652	0.1458
5623	[D D]	-653	0.05771
5624			
5625	At node N371:		
5626	split	lnL	Rel.Prob
5627	[D ABE]	-651	0.3974
5628	[D BE]	-651.1	0.3541
5629	[D B]	-652	0.1535
5630			
5631	At node N386:		
5632	split	lnL	Rel.Prob
5633	[BC D]	-651.5	0.2492
5634	[C D]	-651.7	0.1971
5635	[B D]	-651.7	0.1947
5636	[B B]	-652.2	0.1264
5637	[D D]	-652.6	0.08486
5638			
5639	At node N384:		
5640	split	lnL	Rel.Prob
5641	[BC C]	-650.6	0.6245
5642	[C C]	-651.7	0.2119
5643			
5644	At node N382:		
5645	split	lnL	Rel.Prob
5646	[B C]	-650.3	0.7878
5647			
5648	At node N380:		
5649	split	lnL	Rel.Prob
5650	[B B]	-650.3	0.8442
5651			
5652	At node N378:		
5653	split	lnL	Rel.Prob

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5655

5656 At node N376:

5657 split lnL Rel.Prob

5658 [B|B] -650.1 0.9923

5659

5660 At node N392:

5661 split lnL Rel.Prob

5662 [ABE|A] -650.5 0.6524

5663 [AE|A] -652.1 0.1315

5664

5665 At node N390:

5666 split lnL Rel.Prob

5667 [BE|A] -650.9 0.4575

5668 [B|AE] -651.1 0.3564

5669 [E|A] -652.6 0.08032

5670

5671 At node N405:

5672 split lnL Rel.Prob

5673 [CD|C] -650.6 0.5922

5674 [C|C] -651.5 0.2562

5675

5676 At node N404:

5677 split lnL Rel.Prob

5678 [C|C] -650.1 0.9867

5679

5680 At node N411:

5681 split lnL Rel.Prob

5682 [B|C] -650.4 0.7165

5683

5684 At node N409:

5685 split lnL Rel.Prob

5686 [B|B] -650.1 0.9548

5687

5688 At node N567:

5689 split lnL Rel.Prob

5690 [BG|G] -650.8 0.4927

5691 [BFG|G] -651.3 0.2914

5692 [B|B] -652.2 0.1232

5693

5694 At node N565:

5695 split lnL Rel.Prob

5696 [BF|G] -651 0.422

5697 [B|G] -651.2 0.3333

5698 [B|B] -652.1 0.1344

5699

5700 At node N563:

5701 split lnL Rel.Prob

5702 [B|F] -650.7 0.5609

5703 [B|B] -651.9 0.1668

5704 [BD|F] -652.3 0.1081

5705

5706 At node N541:

5707 split lnL Rel.Prob

5708 [BC|C] -650.9 0.4564

5709 [BC|D] -652 0.1498

5710 [B|D] -652.3 0.1122

5711 [B|B] -652.7 0.07664

5712 [BD|D] -652.8 0.06433

5713

5714 At node N525:

5715 split lnL Rel.Prob

5716 [C|B] -650.6 0.6205

5717 [C|BC] -652.4 0.1029

5718	[B B]	-652.4	0.0994
5719			
5720	At node N433:		
5721	split lnL		Rel.Prob
5722	[C C]	-650.3	0.8534
5723			
5724	At node N421:		
5725	split lnL		Rel.Prob
5726	[C C]	-650.1	0.9852
5727			
5728	At node N419:		
5729	split lnL		Rel.Prob
5730	[C C]	-650.1	0.9977
5731			
5732	At node N417:		
5733	split lnL		Rel.Prob
5734	[C C]	-650.1	0.9996
5735			
5736	At node N432:		
5737	split lnL		Rel.Prob
5738	[C C]	-650.2	0.8886
5739			
5740	At node N428:		
5741	split lnL		Rel.Prob
5742	[C C]	-650.1	0.9976
5743			
5744	At node N427:		
5745	split lnL		Rel.Prob
5746	[C C]	-650.1	0.9994
5747			
5748	At node N425:		
5749	split lnL		Rel.Prob
5750	[C C]	-650.1	0.9995
5751			
5752	At node N431:		
5753	split lnL		Rel.Prob
5754	[C C]	-650.4	0.7762
5755	[C CD]	-651.7	0.2091
5756			
5757	At node N524:		
5758	split lnL		Rel.Prob
5759	[B B]	-650.4	0.7462
5760			
5761	At node N452:		
5762	split lnL		Rel.Prob
5763	[B B]	-650.5	0.6666
5764	[B G]	-651.7	0.1957
5765	[B BG]	-652.4	0.09742
5766			
5767	At node N438:		
5768	split lnL		Rel.Prob
5769	[B B]	-650.7	0.5705
5770	[E B]	-651.3	0.305
5771			
5772	At node N437:		
5773	split lnL		Rel.Prob
5774	[B B]	-650.5	0.6489
5775	[B BC]	-652.1	0.1294
5776	[B BD]	-652.4	0.1004
5777			
5778	At node N451:		
5779	split lnL		Rel.Prob
5780	[BG G]	-650.5	0.6585
5781	[G G]	-651.7	0.2074

5782  
 5783 At node N449:  
 5784 split lnL Rel.Prob  
 5785 [G|BG] -650.5 0.6959  
 5786 [G|G] -651.6 0.2191  
 5787  
 5788 At node N443:  
 5789 split lnL Rel.Prob  
 5790 [G|G] -650.1 0.9536  
 5791  
 5792 At node N441:  
 5793 split lnL Rel.Prob  
 5794 [G|G] -650.4 0.7688  
 5795 [BG|G] -651.6 0.2161  
 5796  
 5797 At node N448:  
 5798 split lnL Rel.Prob  
 5799 [B|G] -650.3 0.7893  
 5800  
 5801 At node N446:  
 5802 split lnL Rel.Prob  
 5803 [B|B] -650.2 0.9314  
 5804  
 5805 At node N523:  
 5806 split lnL Rel.Prob  
 5807 [B|B] -650.3 0.7987  
 5808  
 5809 At node N517:  
 5810 split lnL Rel.Prob  
 5811 [B|B] -650.6 0.6034  
 5812  
 5813 At node N503:  
 5814 split lnL Rel.Prob  
 5815 [B|B] -651.5 0.2345  
 5816 [BD|E] -652 0.1466  
 5817 [B|E] -652 0.145  
 5818 [C|B] -652.5 0.09435  
 5819 [D|B] -652.9 0.05817  
 5820 [BE|E] -653 0.05352  
 5821 [D|BE] -653.3 0.04203  
 5822  
 5823 [BDE|E] -653.4 0.03749  
 5824  
 5825 At node N499:  
 5826 split lnL Rel.Prob  
 5827 [BD|D] -651.4 0.2616  
 5828 [BCD|D] -652.2 0.1179  
 5829 [D|D] -652.4 0.1007  
 5830 [BDE|D] -652.5 0.09466  
 5831 [BC|C] -652.6 0.08598  
 5832 [C|C] -652.6 0.08529  
 5833 [BC|D] -653.2 0.04316  
 5834 [B|D] -653.3 0.0415  
 5835  
 5836 At node N463:  
 5837 split lnL Rel.Prob  
 5838 [C|BCD] -651.6 0.2266  
 5839 [C|C] -652 0.1499  
 5840 [C|BC] -652 0.1496  
 5841 [C|BD] -652.1 0.1339  
 5842 [C|CD] -652.6 0.08131  
 5843 [D|BDE] -653.1 0.04856  
 5844 [B|BDE] -653.2 0.0442  
 5845

5846 At node N462:  
5847 split lnL Rel.Prob  
5848 [BD|D] -652.1 0.1385  
5849 [C|C] -652.1 0.1339  
5850 [BC|C] -652.3 0.1096  
5851 [BDE|D] -652.4 0.09972  
5852 [BCD|C] -652.8 0.06742  
5853 [BD|C] -652.8 0.06512  
5854 [BCD|D] -652.9 0.06137  
5855 [D|D] -652.9 0.05785  
5856 [B|CD] -653.2 0.04407  
5857 [D|CD] -653.2 0.04406  
5858 [ABDE|D] -653.5 0.03198  
5859 [B|C] -653.6 0.0295  
5860  
5861 At node N456:  
5862 split lnL Rel.Prob  
5863 [CD|B] -651.5 0.2498  
5864 [C|B] -651.9 0.1598  
5865 [D|BE] -652.1 0.1325  
5866 [BD|E] -652.2 0.1274  
5867 [D|ABE] -652.3 0.106  
5868 [D|B] -653 0.05638  
5869  
5870 At node N461:  
5871 split lnL Rel.Prob  
5872 [D|D] -651.2 0.3467  
5873 [C|C] -651.2 0.3335  
5874 [C|CD] -652.3 0.1098  
5875 [D|CD] -652.4 0.104  
5876  
5877 At node N460:  
5878 split lnL Rel.Prob  
5879 [D|C] -650.4 0.7533  
5880  
5881 At node N498:  
5882 split lnL Rel.Prob  
5883 [D|D] -650.5 0.6949  
5884 [CD|D] -651.5 0.2462  
5885  
5886 At node N496:  
5887 split lnL Rel.Prob  
5888 [D|D] -650.6 0.5813  
5889 [D|CD] -652.2 0.1279  
5890 [CD|C] -652.3 0.1083  
5891 [CD|D] -652.3 0.107  
5892  
5893 At node N484:  
5894 split lnL Rel.Prob  
5895 [D|D] -650.7 0.532  
5896 [D|CD] -651.1 0.3822  
5897  
5898 At node N470:  
5899 split lnL Rel.Prob  
5900 [D|D] -650.3 0.7788  
5901  
5902 At node N466:  
5903 split lnL Rel.Prob  
5904 [D|D] -651.2 0.3252  
5905 [DF|F] -651.9 0.1621  
5906 [F|DF] -651.9 0.1621  
5907 [DF|D] -651.9 0.1606  
5908 [D|DF] -651.9 0.1606  
5909

5910 At node N469:  
 5911 split lnL Rel.Prob  
 5912 [D|D] -650.5 0.6584  
 5913  
 5914 At node N483:  
 5915 split lnL Rel.Prob  
 5916 [C|CD] -650.4 0.7336  
 5917 [C|D] -651.7 0.1932  
 5918  
 5919 At node N475:  
 5920 split lnL Rel.Prob  
 5921 [C|C] -650.1 0.9891  
 5922  
 5923 At node N473:  
 5924 split lnL Rel.Prob  
 5925 [C|C] -650.1 0.9978  
 5926  
 5927 At node N482:  
 5928 split lnL Rel.Prob  
 5929 [CD|D] -650.8 0.475  
 5930 [D|D] -651.7 0.2022  
 5931 [C|CD] -652.4 0.1045  
 5932 [D|CD] -652.4 0.1009  
 5933  
 5934 At node N481:  
 5935 split lnL Rel.Prob  
 5936 [D|D] -650.7 0.5739  
 5937 [D|CD] -651.3 0.29  
 5938 [DF|D] -652.5 0.09218  
 5939  
 5940 At node N479:  
 5941 split lnL Rel.Prob  
 5942 [F|D] -650.2 0.8813  
 5943  
 5944 At node N495:  
 5945 split lnL Rel.Prob  
 5946 [D|D] -650.9 0.4401  
 5947 [D|CD] -652.1 0.1391  
 5948 [C|CD] -652.2 0.1271  
 5949 [CD|C] -652.4 0.1047  
 5950 [CD|D] -652.4 0.09643  
 5951 [C|C] -652.6 0.08373  
 5952  
 5953 At node N489:  
 5954 split lnL Rel.Prob  
 5955 [D|C] -650.3 0.8332  
 5956  
 5957 At node N487:  
 5958 split lnL Rel.Prob  
 5959 [D|D] -650.1 0.9867  
 5960  
 5961 At node N494:  
 5962 split lnL Rel.Prob  
 5963 [C|D] -650.4 0.7203  
 5964 [D|D] -652 0.1558  
 5965  
 5966 At node N492:  
 5967 split lnL Rel.Prob  
 5968 [C|C] -650.1 0.9855  
 5969  
 5970 At node N502:  
 5971 split lnL Rel.Prob  
 5972 [E|A] -650.4 0.7769  
 5973 [BE|A] -652.1 0.1406

5974  
 5975 At node N516:  
 5976 split lnL Rel.Prob  
 5977 [B|E] -651 0.3993  
 5978 [B|B] -651.4 0.2728  
 5979 [B|AE] -652.4 0.09779  
 5980 [BE|A] -652.6 0.07995  
 5981  
 5982 At node N514:  
 5983 split lnL Rel.Prob  
 5984 [B|B] -650.3 0.8094  
 5985  
 5986 At node N512:  
 5987 split lnL Rel.Prob  
 5988 [B|B] -650.1 0.9696  
 5989  
 5990 At node N508:  
 5991 split lnL Rel.Prob  
 5992 [B|B] -650.3 0.8576  
 5993  
 5994 At node N506:  
 5995 split lnL Rel.Prob  
 5996 [B|B] -650.4 0.7583  
 5997  
 5998 At node N511:  
 5999 split lnL Rel.Prob  
 6000 [B|B] -650.1 0.9947  
 6001  
 6002 At node N522:  
 6003 split lnL Rel.Prob  
 6004 [B|B] -651.1 0.3633  
 6005 [B|BC] -651.6 0.2212  
 6006 [B|BCD] -652.3 0.1065  
 6007 [BE|B] -652.4 0.1011  
 6008 [B|BD] -652.9 0.06128  
 6009  
 6010 At node N521:  
 6011 split lnL Rel.Prob  
 6012 [BCD|C] -650.8 0.506  
 6013 [BC|C] -651.1 0.3687  
 6014  
 6015 At node N540:  
 6016 split lnL Rel.Prob  
 6017 [CD|D] -650.8 0.5137  
 6018 [D|D] -651.2 0.3472  
 6019  
 6020 At node N536:  
 6021 split lnL Rel.Prob  
 6022 [C|D] -650.3 0.8329  
 6023  
 6024 At node N534:  
 6025 split lnL Rel.Prob  
 6026 [C|C] -650.1 0.9725  
 6027  
 6028 At node N533:  
 6029 split lnL Rel.Prob  
 6030 [C|C] -650.1 0.9975  
 6031  
 6032 At node N532:  
 6033 split lnL Rel.Prob  
 6034 [C|C] -650.1 0.9996  
 6035  
 6036 At node N530:  
 6037 split lnL Rel.Prob



6038	[C C]	-650.1	0.9999
6039			
6040	At node N539:		
6041	split lnL		Rel.Prob
6042	[D D]	-650.1	0.9776
6043			
6044	At node N562:		
6045	split lnL		Rel.Prob
6046	[F F]	-650.3	0.8164
6047			
6048	At node N552:		
6049	split lnL		Rel.Prob
6050	[F F]	-650.1	0.9778
6051			
6052	At node N544:		
6053	split lnL		Rel.Prob
6054	[F F]	-650.1	0.9933
6055			
6056	At node N551:		
6057	split lnL		Rel.Prob
6058	[F F]	-650.1	0.993
6059			
6060	At node N547:		
6061	split lnL		Rel.Prob
6062	[F F]	-650.1	0.9963
6063			
6064	At node N550:		
6065	split lnL		Rel.Prob
6066	[F F]	-650.1	0.9952
6067			
6068	At node N561:		
6069	split lnL		Rel.Prob
6070	[F F]	-650.3	0.7932
6071			
6072	At node N557:		
6073	split lnL		Rel.Prob
6074	[F F]	-650.4	0.7555
6075			
6076	At node N555:		
6077	split lnL		Rel.Prob
6078	[E F]	-651.2	0.3377
6079	[A EF]	-651.3	0.2905
6080	[AE F]	-651.6	0.2318
6081	[F F]	-652.8	0.0666
6082			
6083	At node N560:		
6084	split lnL		Rel.Prob
6085	[F F]	-650.2	0.8657
6086			
6087	At node N721:		
6088	split lnL		Rel.Prob
6089	[H B]	-651.2	0.3326
6090	[H E]	-651.2	0.3204
6091	[E E]	-652.5	0.09348
6092	[B B]	-652.8	0.06578
6093			
6094	At node N669:		
6095	split lnL		Rel.Prob
6096	[H H]	-650.3	0.8386
6097			
6098	At node N720:		
6099	split lnL		Rel.Prob
6100	[BE E]	-651.6	0.2191
6101	[E E]	-652	0.1541

6102	[B E]	-652.1	0.1362
6103	[ABE A]	-652.6	0.08338
6104	[B B]	-652.6	0.08215
6105	[ABE E]	-653	0.05687
6106	[B AE]	-653.4	0.0387
6107			
6108	At node N718:		
6109	split	lnL	Rel.Prob
6110	[BE B]	-651.2	0.3341
6111	[ABE B]	-651.5	0.241
6112	[B B]	-651.5	0.241
6113	[ABDE B]	-652.9	0.06087
6114			
6115	At node N688:		
6116	split	lnL	Rel.Prob
6117	[BE B]	-651.4	0.2678
6118	[ABE B]	-651.6	0.2337
6119	[B B]	-651.7	0.2106
6120	[B C]	-652.8	0.06488
6121	[ABE D]	-652.9	0.06322
6122	[AE B]	-652.9	0.05797
6123			
6124	At node N686:		
6125	split	lnL	Rel.Prob
6126	[ABE A]	-650.3	0.7945
6127			
6128	At node N684:		
6129	split	lnL	Rel.Prob
6130	[AE B]	-650.6	0.6109
6131	[A BE]	-651.5	0.236
6132			
6133	At node N682:		
6134	split	lnL	Rel.Prob
6135	[A A]	-650.8	0.515
6136	[AE A]	-651	0.387
6137			
6138	At node N680:		
6139	split	lnL	Rel.Prob
6140	[A A]	-650.6	0.6007
6141	[A AE]	-651.1	0.3802
6142			
6143	At node N674:		
6144	split	lnL	Rel.Prob
6145	[A A]	-650.1	0.998
6146			
6147	At node N672:		
6148	split	lnL	Rel.Prob
6149	[A A]	-650.1	0.9995
6150			
6151	At node N679:		
6152	split	lnL	Rel.Prob
6153	[A A]	-650.6	0.5868
6154	[AE A]	-651.1	0.3752
6155			
6156	At node N677:		
6157	split	lnL	Rel.Prob
6158	[AE A]	-651.6	0.2187
6159	[A AE]	-651.6	0.2187
6160	[AE E]	-651.6	0.2157
6161	[E AE]	-651.6	0.2157
6162	[A A]	-652.2	0.1201
6163			
6164	At node N717:		
6165	split	lnL	Rel.Prob

6166	[B B]	-650.7	0.5612
6167	[C B]	-651.2	0.3211
6168			
6169	At node N691:		
6170	split lnL		Rel.Prob
6171	[C C]	-650.3	0.8546
6172			
6173	At node N716:		
6174	split lnL		Rel.Prob
6175	[B B]	-650.2	0.9071
6176			
6177	At node N704:		
6178	split lnL		Rel.Prob
6179	[B B]	-650.2	0.9361
6180			
6181	At node N702:		
6182	split lnL		Rel.Prob
6183	[B B]	-650.3	0.8274
6184			
6185	At node N696:		
6186	split lnL		Rel.Prob
6187	[B F]	-650.3	0.8068
6188			
6189	At node N695:		
6190	split lnL		Rel.Prob
6191	[F F]	-650.2	0.9231
6192			
6193	At node N701:		
6194	split lnL		Rel.Prob
6195	[B B]	-650.6	0.6136
6196	[BD B]	-651.8	0.1872
6197			
6198	At node N699:		
6199	split lnL		Rel.Prob
6200	[D BD]	-650.6	0.601
6201	[D BDF]	-651.6	0.2255
6202			
6203	At node N715:		
6204	split lnL		Rel.Prob
6205	[B B]	-650.3	0.8361
6206			
6207	At node N707:		
6208	split lnL		Rel.Prob
6209	[B B]	-650.1	0.9913
6210			
6211	At node N714:		
6212	split lnL		Rel.Prob
6213	[B B]	-650.6	0.5998
6214	[B BC]	-652.6	0.08404
6215			
6216	At node N713:		
6217	split lnL		Rel.Prob
6218	[B C]	-651.5	0.2573
6219	[B D]	-651.5	0.2566
6220	[B B]	-651.9	0.1687
6221	[B CD]	-652.1	0.1374
6222	[BF D]	-652.8	0.06962
6223			
6224	At node N711:		
6225	split lnL		Rel.Prob
6226	[B B]	-650.7	0.5485
6227	[B BF]	-650.9	0.4353
6228			
6229			